

SHORT GENOME REPORT

Open Access



Complete genome sequence of *Paenibacillus yonginensis* DCY84^T, a novel plant Symbiont that promotes growth via induced systemic resistance

Yeon-Ju Kim^{1*†}, Johan Sukweenadhi^{1†}, Ji Woong Seok², Chang Ho Kang³, Eul-Su Choi¹, Sathiyamoorthy Subramaniyam¹ and Deok Chun Yang^{1*}

Abstract

This article reports the full genome sequence of *Paenibacillus yonginensis* DCY84^T (KCTC33428, JCM19885), which is a Gram-positive rod-shaped bacterium isolated from humus soil of Yongin Forest in Gyeonggi Province, South Korea. The genome sequence of strain DCY84^T provides greater understanding of the *Paenibacillus* species for practical use. This bacterium displays plant growth promotion via induced systemic resistance of abiotic stresses.

Keywords: *Paenibacillus yonginensis* DCY84^T, Genome, PacBio, Plant growth promoting rhizobacteria (PGPR)

Introduction

Various *Paenibacillus* species constitute a large group of facultative anaerobic endospore-forming Gram-positive bacteria that are extensively distributed in nature. Ash et al. proposed that members of 'group 3' within the genus *Bacillus* should be transferred to the genus *Paenibacillus*, for which they proposed *Paenibacillus polymyxa* as the type species [1]. Since that time, 174 different type species have been described.

Members of the genus *Paenibacillus* are well known as PGPR, together with *Azotobacter*, *Azospirillum*, *Pseudomonas*, *Acetobacter*, and *Burkholderia* [2]. While many new species from the genus *Paenibacillus* have been reported [3], the type species *Paenibacillus polymyxa* [4] is considered a PGPR that is widely used in sustainable agriculture and environmental remediation because of its multiple functions [2, 5]. Coupled with many plant species, some *Paenibacillus* species have been developed as biofertilizers or biocontrol agents and have been used effectively in the control of plant-pathogenic fungi, bacteria, and nematodes [5–7]. *P. yonginensis* DCY84^T

was isolated from a decomposed humus mixture in South Korea and its plant growth promotion traits have been characterized in vitro [8]. This strain is capable of inducing the defense response of *Arabidopsis* against several abiotic stresses [9]. Genome sequencing of *P. yonginensis* DCY84^T was conducted to obtain additional insights into the physiological characteristics involved in microbe-plant interactions and to facilitate better understanding of the molecular basis of these traits.

Organism information

Classification and features

Paenibacillus yonginensis DCY84^T was isolated from a decomposed humus mixture collected from Yongin province. It is a Gram-positive bacterium that can grow on Tryptic soy broth agar at 28 °C. Cells of strain DCY84^T are rod-shaped with a diameter ranging from 0.7–0.9 μm and length ranging from 3.4 to 4.7 μm. Growth occurs under aerobic conditions with an optimum growth temperature at 25–30 °C and a temperature range of 15–40 °C, general features of strain DCY84^T were presented in Table 1. Phylogenetic tree highlighting the position of *Paenibacillus yonginensis* DCY84^T and phylogenetic inferences were obtained using the maximum-likelihood method (Fig. 1). Cell

* Correspondence: yeonjukim@khu.ac.kr; deokchunyang@yahoo.co.kr

†Equal contributors

¹Graduate School of Biotechnology, College of Life Science, Kyung Hee University, Yongin 446-701, South Korea

Full list of author information is available at the end of the article

Table 1 Classification and general features of *Paenibacillus yonginensis* DCY84^T

MIGS ID	Property	Term	Evidence Code
	Classification	Domain Bacteria	TAS [17]
		Phylum <i>Firmicutes</i>	TAS [18, 19]
		Class <i>Bacilli</i>	TAS [20]
		Order <i>Bacillales</i>	TAS [21, 22]
		Family <i>Paenibacillaceae</i>	TAS [21, 23]
		Genus <i>Paenibacillus</i>	TAS [15]
		Species <i>Paenibacillus yonginensis</i>	TAS [8, 9]
		Strain DCY84 ^T	TAS [8, 9]
	Gram stain	positive	IDA
	Cell shape	rod	IDA
	Motility	motile	IDA
	Sporulation	spore production	IDA
	Temperature range	15–40 °C	IDA
	Optimum temperature	30 °C	IDA
	pH range; Optimum	5–9; 8	IDA
	Carbon source	D-Xylose, D-ribose, D-glucose and others	TAS [8]
MIGS-6	Habitat	humus soil	IDA
MIGS-6.3	Salinity	0.5–4.5% NaCl	IDA
MIGS-22	Oxygen requirement	Aerobic	IDA
	Carbon source	glucose, lactose	TAS [8]
MIGS-15	Biotic relationship	Free-living	IDA
MIGS-14	Pathogenicity	Non-pathogenic	NAS
MIGS-13	Source material identifiers	KCTC 33428 ^T , JCM 19885 ^T	TAS [8]
MIGS-4	Geographic location	South Korea: Gyeonggi province	IDA
MIGS-5	Sample collection	September 2013	IDA
MIGS-4.1	Latitude	37.314 N	IDA
MIGS-4.2	Longitude	127.268 W	IDA
MIGS-4.4	Altitude	131.37 m	IDA

Evidence codes: *IDA* inferred from direct assay, *TAS* traceable author statement (i.e., a direct report exists in the literature), and *NAS* non-traceable author statement (i.e., not directly observed for the living, isolated sample, but based on a generally accepted property for the species, or anecdotal evidence). These evidence codes are from the Gene Ontology project [24]

morphology was examined using scanning electron microscopy (Fig. 2).

Genome sequencing information

Genome project history

P. yonginensis DCY84^T was selected for genome sequencing because we observed the presence of a unique compatible solute for plant protection from biotic stress and potential plant growth promoting activity with rice in reclaimed paddy soil and *Panax ginseng* C.A.Mey, respectively. The complete genome sequence has been deposited in the NCBI sequencing read archive under NCBI BioProject PRJNA306396 with BioSample SAMN04419545 and

overall sequencing project information was presented in Table 2. Sequencing, annotation, and analysis were performed at LabGenomics (Seongnam, Republic of Korea).

Growth conditions and genomic DNA preparation

For growth and genomic DNA preparation, *P. yonginensis* DCY84^T (KCTC 33428^T=JCM 19885^T) was grown in DSMZ medium 1 (Nutrient Agar) at 28 °C. DNA was isolated from 0.5–1 g of cell paste using the JetFlex genomic protocol as recommended by the manufacturer. For genome sequencing and assembly, the draft genome of *P. yonginensis* DCY84^T was generated using the PacBio platform following the manufacturer's instructions.

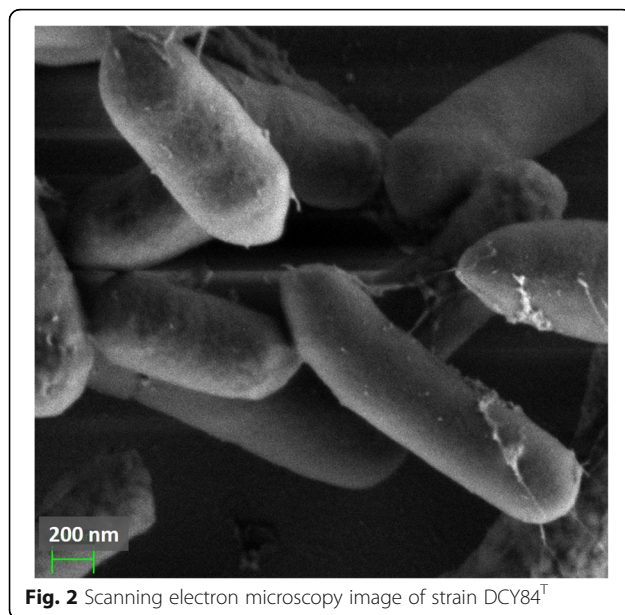
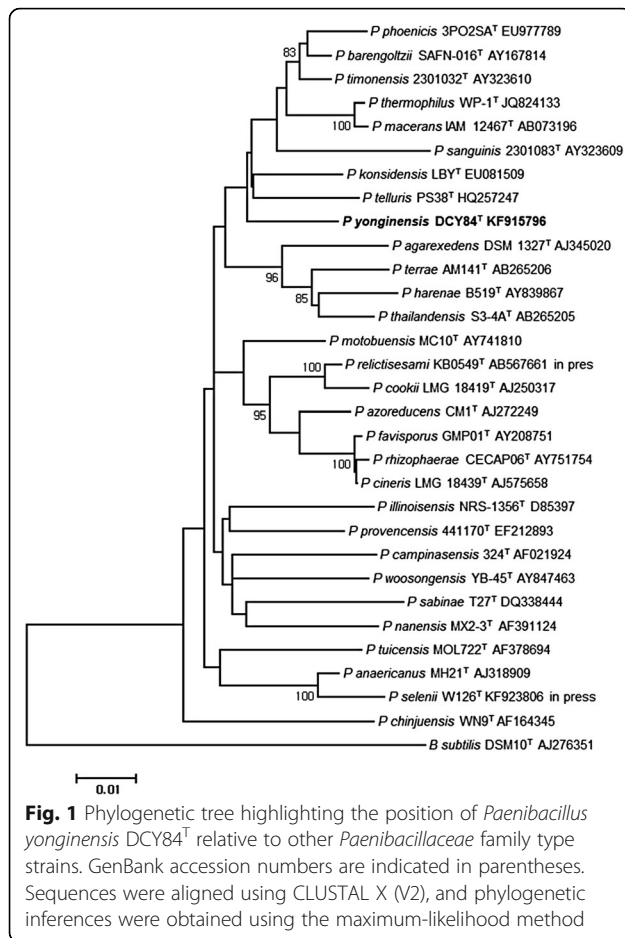


Table 2 Genome sequencing project information for *Paenibacillus yonginensis* DCY84^T

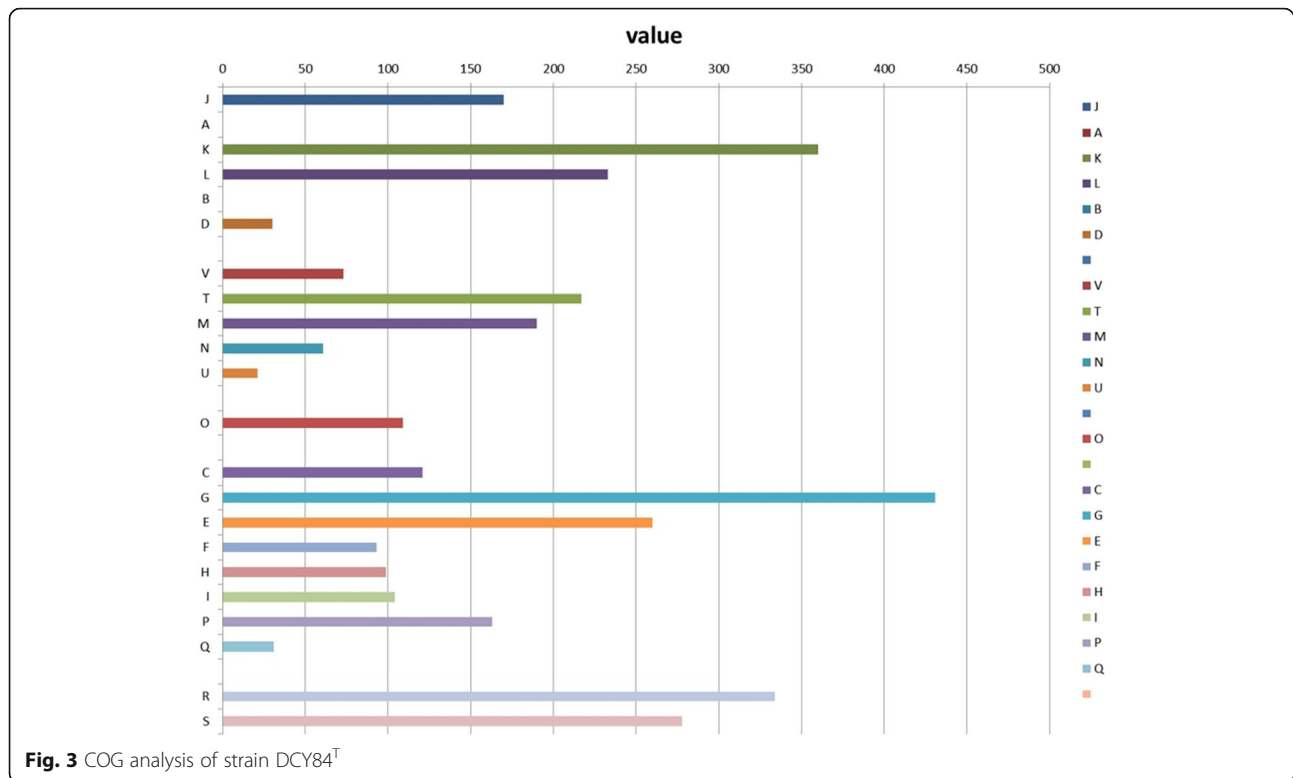
MIGS ID	Property	Term
MIGS 31	Finishing quality	Finished
MIGS-28	Libraries used	Pacbio SMRTbell™ library
MIGS 29	Sequencing platforms	PacBio RS
MIGS 31.2	Fold coverage	130X
MIGS 30	Assemblers	SMRT Analysis v2.3.0 HGAP.2
MIGS 32	Gene calling method	Glimmer 3.02 ex: Prodigal, GenePRIMP
	Locus Tag	AWM70
	GenBank ID	CP014167
	GenBank Date of Release	January 28, 2016
	GOLD ID	Gp0177323
	BIOPROJECT	PRJNA306396
MIGS 13	Source Material Identifier	KCTC 33428 ^T , JCM 19885 ^T
	Project relevance	Taxonomy, agriculture, plant-microbe interactions

Genome sequencing and assembly

Sequencing produced 74,264 reads with an average length of 7828 bp, which was assembled using *the de novo* HGAP implemented within the analysis pipeline SMRT Analysis 2.2 (Pacific Biosciences, CA, USA). Ambiguous base and inserted/deleted regions between the PacBio assembled and preassembled high quality draft sequences were manually corrected using consensus sequences for final assembly. Long reads were selected as the seed sequences for constructing preassemblies, and the other short reads were mapped to the seeds using BLASTR software for alignment, which corrected errors in the long reads and thus increased the accuracy rating of bases. The sequencing run yielded 581,398,217 filtered and sub-read bases and a total of 113,985,693 pre-assembled bases were used for deep sequencing. tRNA and rRNA genes were identified by tRNAscan-SE version 1.3 [10] and RNAmmer version 1.2 [11]. The ORFs were predicted using Glimmer 3.02 and the annotation of predicted genes was conducted using Blastall 2.2.26. Protein coding genes were annotated based on the COGs database.

Genome annotation

The purpose of the present study was to develop a better understanding of the *P. yonginensis* DCY84^T genetic background to develop more effective utilization of the strain. COGs analysis of strain DCY84^T is shown in Fig. 3 and the number of genes associated with the 22 general COGs functional categories presented in Table 3. The analysis of the full *P. yonginensis* DCY84^T genome in



comparison with other related *Paenibacillus* strains is included in Additional file 1: Table S1.

The *iaaM* gene, also gene responsible for IAA synthesis, siderophores production, phosphate transporter, phosphonate cluster, antimicrobial production, and synthesis of the volatile organic compound *bdhA* are present in the *P. yonginensis* DCY84^T genome. These genes corroborate with our physiological results demonstrating plant growth promotion and induced systemic resistance in the plant symbiont [9, 10].

Insights from the genome sequence

The completed *P. yonginensis* DCY84^T genome consists of a single circular chromosome of 4,985,901 bp, with a GC content of 51.01%, which is similar to most *Paenibacillus* strains (45 – 54%) as reported previously [12] (Fig. 4). The genome size of the strain DCY84^T (4.985 Mb) is smaller than the other sequenced members of genus *Paenibacillus* including *P. polymyxa* CF05 (5.76 Mb), and *P. mucilaginosus* 3016 (8.74 Mb) [13]. Full genome of DCY84^T was annotated by following NCBI prokaryotic genome annotation pipeline [14]. A total of 4498 genes were predicted for the genome, including 4233 coding sequences (94.1% of total genes) and 147 pseudo genes. Nucleotide content and gene count levels of the chromosome were summarized in Table 4. More detail annotation of the strain

DCY84^T was available in Additional file 2: Table S5. Most of selected *Paenibacillus* strain was reported to have plant growth promoting factor traits. The summary features of DCY84^T and referred strains are showed on Additional file 1: Table S1 below, including the genome accession number, genome size, GC content, annotation information, protein, Gene, Pseudo gene. The COGs analysis of strain DCY84^T and other closely related *Paenibacillus* strains was provided on Additional file 1: Table S2 (direct plant growth promoting factors) and Additional file 1: Table S3 (indirect plant growth promoting factors). The genome of *P. yonginensis* DCY84^T and *P. polymyxa* M1 were visualized in Additional file 3: Figure S1 by the comparison using the Artemis software and ACT [15]. Strain DCY84^T increased nutrient availability by producing several hydrolyzing enzymes, amino acid transporter proteins (Additional file 1: Table S4). Moreover, Strain DCY84^T treatment can induce plant defense mechanism mediated by ABA signal under salinity stress.

Extended insights

Genome analysis showed that *P. yonginensis* DCY84^T contained many genes related to the stress response, such as IAA, choline, glutamate decarboxylase and malate transporters, potassium uptake protein, heat shock proteins, chaperone proteins, and sugar transporters.

Table 3 Number of genes associated with the 22 general COG functional categories

Code	Value	%age ^a	Description
J	170	4.02	Translation, ribosomal structure and biogenesis
A	0	0.00	RNA processing and modification
K	360	8.50	Transcription
L	233	5.50	Replication, recombination and repair
B	0	0.00	Chromatin structure and dynamics
D	30	0.71	Cell cycle control, cell division, chromosome partitioning
V	73	1.72	Defense mechanisms
T	217	5.13	Signal transduction mechanisms
M	190	4.49	Cell wall/membrane/envelope biogenesis
N	61	1.44	Cell motility
U	21	0.50	Intracellular trafficking, secretion, and vesicular transport
O	109	2.58	Posttranslational modification, protein turnover, chaperones
C	121	2.86	Energy production and conversion
G	431	10.18	Carbohydrate transport and metabolism
E	260	6.14	Amino acid transport and metabolism
F	93	2.20	Nucleotide transport and metabolism
H	99	2.34	Coenzyme transport and metabolism
I	104	2.46	Lipid transport and metabolism
P	163	3.85	Inorganic ion transport and metabolism
Q	31	0.73	Secondary metabolites biosynthesis, transport and catabolism
R	334	7.89	General function prediction only
S	278	6.57	Function unknown
–	1372	32.41	Not in COGs
^b Total	4750	112.21	

^aThe percentage is based on the total number of protein coding genes in the annotated genome

^bThe total does not correspond to 4498 CDS because some genes are associated with more than one COG functional categories

These genes most likely allow the strain to cope with different environmental stresses. Experimentation and additional analysis of these genes may help to elucidate the mechanisms mediating the stress response and facilitate the development of *P. yonginensis* DCY84^T as a biofertilizer. When the strain DCY84^T was used as a treatment for early sprouting rice seeds, several genes responsible for primary metabolism were upregulated in the rice root, which could be related to PGPR. These results indicate that *P. yonginensis* DCY84^T might have the potential for application in industrial biotechnology as a producer of miscellaneous hydrolases.

This is the first report describing the genome sequence of *P. yonginensis* DCY84^T. When coated on sprouting rice seeds or seedlings directly on paddy soil, strain DCY84^T and silica zeolite complex were shown to enhance rice yield and also increase GABA content in brown rice. Treatment was also shown to induce systemic stress resistance responses in rice and *Arabidopsis* under heavy metal and salty conditions. Furthermore,

the sequence of *P. yonginensis* DCY84^T provides useful information and may contribute to agricultural applications of *Paenibacillus* genera in practical biotechnology. Rice yield was affected by the amount of strain DCY84^T administered during the early sprouting stage. Silica zeolite complex and strain DCY84^T treatment inhibited the occurrence of fungal infection, and also enhanced rice quality. Silica zeolite complex and two treatments with strain DCY84^T resulted in the highest head rice levels (86.8%) compared to a one-time treatment of DCY84^T (67.9%), and without strain DCY84^T treatment (46.4%). The PGPR treatment enhanced head rice levels by 40.4% [16]. Strain treatment also enhanced nitrogen uptake and increased levels of stored nitrogen in the rice grain, indicating that the strain DCY84^T enhanced plant nitrogen utilization with less nitrogen fertilizer application. The most important parameters for economic rice value are head rice rate and good appearance; strain DCY84^T treatment enhanced both the rice quality and reduced commercial nitrogen fertilizer usage.

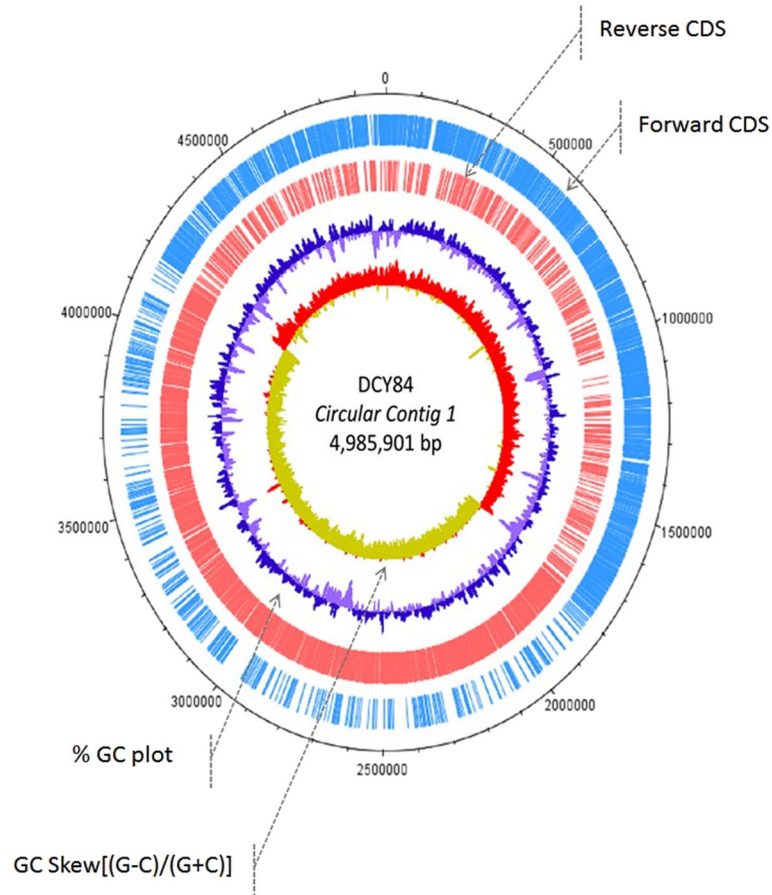


Fig. 4 Graphical circular map of the chromosome. From the outside to the center, genes on the forward strand are colored by COG categories (only genes assigned to COG), genes on the reverse strand are colored by COG categories (only genes assigned to COG), RNA genes (tRNAs green, rRNAs red), G + C content, and GC skew. Purple and olive colors indicate negative and positive values, respectively

Table 4 Genome statistics

Attribute	Value	% of Total
Genome size (bp)	4,985,901	100.0
DNA coding (bp)	4,267,050	85.6
DNA G + C (bp)	2,543,529	51.0
DNA scaffolds	1	100.0
Total genes	4498	100.0
Protein coding genes	4233	94.1
RNA genes	118	2.6
Pseudo genes	121	2.7
Genes in internal clusters	792	17.6
Genes with function prediction	4380	97.4
Genes assigned to COGs	3378	75.1
Genes with Pfam domains	2661	59.2
Genes with signal peptides	295	6.6
Genes with transmembrane helices	1197	26.6
CRISPR repeats	4	–

Conclusion

The DCY84^T strain was isolated from a decomposed humus mixture. Phylogenetic analysis based on the 16S rRNA gene confirmed its affiliation to the genus *Paenibacillus*. G + C content, COGs, and average nucleotide identities are presented. The genomic features of strain DCY84^T are consistent with the plant growth promoting activity of this strain, including IAA production, phosphate solubilizing activity, and siderophores production. In addition, DCY84^T induced systemic stress resistance mechanisms in rice and *Arabidopsis* under heavy metal and salty conditions.

Additional files

Additional file 1: Table S1. Genome comparison of strain DCY84^T and closest *Paenibacillus* strains. **Table S2.** COGs analysis of direct plant growth promoting traits. **Table S3.** COGs analysis of indirect plant growth promoting traits. **Table S4.** Some important genes annotated on strain DCY84^T genome. (DOCX 81 kb)

Additional file 2: Table S5. Annotation of the *Paenibacillus yonginensis* DCY84^T genome. (XLSX 443 kb)

Additional file 3: Figure S1. Comparative genome analysis of *P. yonginensis* DCY84^T and *P. polymyxa* M1 using the Artemis software and ACT. (TIFF 16717 kb)

Abbreviations

bdhA: 2,3-butanediol synthesis; COGs: Clusters of Orthologous Groups of proteins; HGAP: Hierarchical Genome Assembly Process; IAA: Indole-3-acetic acid; *iaaM*: Tryptophan monooxygenase; ORFs: Open Reading Frames; PGPR: Plant Growth Promoting Rhizobacteria; SMRT: Single Molecule, Real-Time

Acknowledgements

We appreciated to the company, Saturn Bio Tech for rice field trials, they supported for application of the strain DCY84 as bio fertilizer in reclaimed paddy soil.

Funding

This study was supported by a grant from the Next-Generation BioGreen 21 ("PJ012034"), Rural Development Administration, in Republic of Korea.

Authors' contributions

YJK designed the study, carried out the genome analysis, and drafted the manuscript. JS performed DNA isolation, electron microscopy, the phylogenetic analysis for taxonomic study and corrected the manuscript. JWS and CHK carried out the sequencing and helped to draft the manuscript. ESC and SS participated in the study design. DCY coordinated. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Author details

¹Graduate School of Biotechnology, College of Life Science, Kyung Hee University, Yongin 446-701, South Korea. ²Lab Genomics Co. Ltd, Jinju, South Korea. ³Division of Applied Life Science and PMBBRC, Gyeongsang National University, Jinju, South Korea.

Received: 7 March 2017 Accepted: 27 September 2017

Published online: 13 October 2017

References

- Ash C, Priest FG, Collins MD. Molecular identification of rRNA group 3 bacilli (Ash, Farrow, Wallbanks and Collins) using a PCR probe test. Proposal for the cr (heading level 1) eation of a new genus *Paenibacillus*. Antonie Van Leeuwenhoek. 1993;64(3):253–60.
- Hayat R, Ali S, Amara U, Khalid R, Ahmed I. Soil beneficial bacteria and their role in plant growth promotion: a review. *Ann Microbiol*. 2010;60:579–98.
- Tindall BJ. What is the type species of the genus *Paenibacillus*? Request for an opinion. 2005;50:939–40.
- Kwak YY, Shin JH. Complete genome sequence of *Paenibacillus beijingsensis* 7188^T (=DSM 24997^T), a novel rhizobacterium from jujube garden soil. *J. Biotechnol*. 2016;206:75–6.
- Judicial Commission of the International Committee on Systematics of Prokaryotes. The type species of the genus *Paenibacillus* Ash et al. 1994 is *Paenibacillus polymyxa*. *Opinion 77*. *IJSEM*. 2005;55:513.
- Kwon YS, Lee DY, Rakwal R, Baek SB, Lee JH, Kwak YS, Seo JS, Chung WS, Bae DW, Kim SG. Proteomic analyses of the interaction between the plant-growth promoting rhizobacterium *Paenibacillus polymyxa* E681 and *Arabidopsis thaliana*. *Proteomics*. 2016;16(1):122–35.
- Lapidot D, Dror R, Vered E, Mishli O, Levy D, Helman Y. Disease protection and growth promotion of potatoes (*Solanum tuberosum* L.) by *Paenibacillus dendritiformis*. *Plant Pathol*. 2015;64(3):545–51.
- Goswami D, Parmar S, Vaghela H, Dhandhukia P, Thakker JN. Describing *Paenibacillus mucilaginosus* strain N3 as an efficient plant growth promoting rhizobacteria (PGPR). *Cogent Food Agric*. 2015;1:1000714. doi:10.1080/23311932.2014.1000714.
- Sukweenadhi J, Kim YJ, Lee KJ, Koh SC, Hoang VA, Nguyen NL, Yang DC. *Paenibacillus yonginensis* sp. nov., a potential plant growth promoting bacterium isolated from humus soil of Yongin forest. *Antonie Van Leeuwenhoek*. 2014;106(5):935–45.
- Sukweenadhi J, Kim YJ, Choi ES, Koh SC, Lee SW, Kim YJ, Yang DC. *Paenibacillus yonginensis* DCY84^T induces changes in *Arabidopsis thaliana* gene expression against aluminum, drought, and salt stress. *Microbiol Res*. 2015;172:7–15.
- Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. *Nucleic Acids Res* 25:955–964. <http://dx.doi.org/10.1093/nar/25.5.0955>.
- Lagesen K, Hallin P, Rødland EA, Staerfeldt HH, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. *Nucleic Acids Res* 35:3100–3108. <http://dx.doi.org/10.1093/nar/gkm160>.
- XF H, Li SX, JG W, Wang JF, Fang QL, Chen JS. Transfer of *Bacillus mucilaginosus* and *Bacillus edaphicus* to the genus *Paenibacillus* as *Paenibacillus mucilaginosus* comb. nov. and *Paenibacillus edaphicus* comb. nov. *Int J Syst Evol Microbiol*. 2010;60:8–14.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. NCBI prokaryotic genome annotation pipeline. *Nucleic Acids Res*. 2016;24:1–11.
- Carver TJ, Rutherford KM, Berriman M, Rajandream MA, Barrell BG, Parkhill J. ACT: the Artemis Comparison Tool. *Bioinformatics*. 2005;21(16):3422–3.
- Choi ES, Sukweenadhi J, Kim YJ JKH, Koh SH, Kang CH, Hoang VA, Yang DC. The effects of rice seed dressing with *Paenibacillus yonginensis* and Silicon on crop development on South Korea's reclaimed tidal land. *Field Crop Res*. 2016;188:121–32.
- Woese CR, Kandler O, Wheelis ML. Towards a natural system of organisms: proposal for the domains Archaea, Bacteria, and Eucarya. *Proc Natl Acad Sci*. 1990;87:4576–9.
- Garrity GM, Lilburn TG, Cole JR, Harrison SH, Euzebey J, Tindall BJ. 2007. Taxonomic Outline of Bacteria and Archaea (TOBA) Release 7.7. Michigan: Michigan State University Board of Trustees; 2001-2007. p. 1–5.
- Gibbons NE, Murray RGE. Proposals Concerning the Higher Taxa of Bacteria. *Int J Syst Bacteriol*. 1978;28:1–6.
- Garrity GM, Holt JG. The Road Map to the Manual. In: Garrity GM, Boone DR, Castenholz RW, editors. *Bergey's Manual of Systematic Bacteriology*, vol. 1. Second ed. New York: Springer; 2001. p. 119–66.
- Murray RGE. The Higher Taxa, or, a Place for Everything...? In: Holt JG, editor. *Bergey's Manual of Systematic Bacteriology*, First Edition, Volume 1, The Williams and Wilkins Co. Baltimore; 1984. p. 31–4.
- Ludwig W, Schleifer KH, Whitman WB. Class I. *Bacilli* class. nov. In: De Vos P, Garrity G, Jones D, Krieg NR, Ludwig W, Rainey FA, Schleifer KH, Whitman WB, editors. *Bergey's Manual of Systematic Bacteriology*, vol. 3. Second ed. New York: Springer-Verlag; 2009. p. 19–20.
- Skerman VBD, McGowan V, Sneath PHA. Approved Lists of Bacterial Names. *Int J Syst Bacteriol*. 1980;30:225–420.
- Ashburner M, Ball CA, Blake JA, Botstein D, Butler H, Cherry JM, et al. Gene ontology: tool for the unification of biology. The Gene Ontology Consortium. *Nat Genet*. 2000;25(1):25–9.

Submit your next manuscript to BioMed Central and we will help you at every step:

- We accept pre-submission inquiries
- Our selector tool helps you to find the most relevant journal
- We provide round the clock customer support
- Convenient online submission
- Thorough peer review
- Inclusion in PubMed and all major indexing services
- Maximum visibility for your research

Submit your manuscript at
www.biomedcentral.com/submit

