

# Pertanika Journal of TROPICAL AGRICULTURAL SCIENCE

VOL. 41 (2) MAY 2018



A scientific journal published by Universiti Putra Malaysia Press

## JUTAS Journal of Tropical Agricultural Science AN INTERNATIONAL PEER-REVIEWED JOURNAL

EDITOR-IN-CHIEF Mohd. Zamri-Saad, Malaysia Veterinary Pathology

#### CHIEF EXECUTIVE EDITOR

Nayan Deep S. Kanwal Environmental Issues – Landscape Plant Modelling Applications

UNIVERSITY PUBLICATIONS COMMITTEE Zulkifli Idrus, Chair

#### EDITORIAL STAFF

Journal Officers: Chai Sook Keat, ScholarOne Kanagamalar Silvarajoo, ScholarOne Tee Syin-Ying, ScholarOne Ummi Fairuz Hanapi, Publication Officer

**Editorial Assistants:** Florence Jiyom Rahimah Razali Zulinaardawati Kamarudin

<u>COPY EDITORS</u> Crescentia Morais Doreen Dillah Pooja Terasha Stanslas

PRODUCTION STAFF Pre-press Officers:

Kanagamalar Silvarajoo Nur Farrah Dila Ismail Wong Lih Jiun

Layout & Typeset: Lilian Loh Kian Lin Wong Wai Mann

WEBMASTER Mohd Nazri Othman

PUBLICITY & PRESS RELEASE Magdalene Pokar (ResearchSEA) Florence Jiyom

#### EDITORIAL OFFICE

JOURNAL DIVISION Office of the Deputy Vice Chancellor (R&I) 1<sup>st</sup> Floor, IDEA Tower II UPM-MTDC Technology Centre Universiti Putra Malaysia 43400 Serdang, Selangor Malaysia. Gen Enq.: +603 8947 1622 | 1616 E-mail: executive\_editor.pertanika@upm.my URI: <u>www.journals.jd.upm.edu.my</u>

#### PUBLISHER

UPM PRESS Universiti Putra Malaysia 43400 UPM, Serdang, Selangor, Malaysia. Tel: +603 8946 8855, 8946 8854 Fax: +603 8941 6172 E-mail: penerbit@upm.edu.my URL: http://penerbit.upm.edu.my



EDITORIAL BOARD 2018-2020

Baharuddin Salleh Plant pathologist / Mycologist, Universiti Sains Malaysia, Malaysia.

David Edward Bignell Soil biology and termite biology, University of London, UK.

Eric Standbridge Microbiology, Molecular genetics, Universiti of California, USA.

**Ghizan Saleh** Plant breeding and genetics, Universiti Putra Malaysia, Malaysia.

Idris Abd. Ghani Entomology Insect taxonomy and biodiversity, Integrated pest management, Biological control, Biopesticides, Universiti Kebangsaan Malaysia, Malaysia.

Jamilah Bakar Food Science and Technology, Food Quality / Processing and Preservation, Universiti Putra Malaysia, Malaysia. Kadambot H.M. Siddique, FTSE Crop and environment physiology, Germplasm enhancement, The University of Western Australia, Australia.

Leng-Guan Saw Botany and Conservation, Plant Ecology, Forest Research Institute Malaysia (FRIM), Kepong, Malaysia.

Mohd. Azmi Ambak Fisheries, Universiti Malaysia Terengganu, Malaysia.

Nor Aini Ab-Shukor Tree improvement, Forestry genetics & biotechnology, Universiti Putra Malaysia, Malaysia.

Richard T. Corlett Biological Sciences, Terrestrial Ecology, Climate Change, Conservation Biology, Biogeography, National University of Singapore, Singapore. Shamshuddin Jusop Soil science, Soil mineralogy, Universiti Putra Malaysia, Malaysia.

Son Radu Food safety, Risk assessment, Molecular biology, Universiti Putra Malaysia, Malaysia.

Srini Kaveri Veterinary, Immunology, INSERM, Centre de Recherche Cordeliers, Paris, France.

Suman Kapur Biological Sciences, Agricultural and Animal Biotechnology, Birla Institute of Technology and Science BITS-Pilani, Hyderabad, India.

Wen-Siang Tan Molecular biology, Virology, Protein chemistry, Universiti Putra Malaysia, Malaysia.

Zora Singh Horticulture, Production technology and post-handling of fruit crops, Curtin University, Australia.

#### INTERNATIONAL ADVISORY BOARD 2018-2021

Alexander Salenikovich Forestry, Wood and Forest Sciences, Université Laval, Canada.

**Banpot Napompeth** 

Entomology, Kasetsart University, Thailand.

Pest Management, Imperial College London, UK.

Graham Matthews

Pest Management, Imperial College London, UK.

Denis J. Wright

Jane M. Hughes Genetics, Griffith University, Australia.

> Malcolm Walkinshaw Biochemistry, University of Edinburgh, Scotland.

Manjit S. Kang Plant Breeding and Genetics, Louisiana State University Agric. Center, Baton Rouge, USA.

Peter B. Mather Ecology and Genetics, Queensland University of Technology, Australia. Syed M. Ilyas Project Director, National Institute of Rural Development, Post Harvest Engineering and Technology, Indian Council of Agricultural Research, Hydrerbad, India.

Tanveer N. Khan Plant Breeding and Genetics, The UWA Institute of Agriculture, The University of Western Australia, Australia.

#### ABSTRACTING AND INDEXING OF PERTANIKA JOURNALS

Pertanika is almost 40 years old; this accumulated knowledge has resulted in the journals being abstracted and indexed in SCOPUS (Elsevier), Clarivate Analytics [formerly known as Thomson (ISI)] Web of Science<sup>®®</sup> Core Collection- Emerging Sources Citation Index (ESC). Web of Knowledge [BIOSIS & CAB Abstracts], EBSCO and EBSCOhost, DOAJ, ERA, Google Scholar, TIB, MyCite, Islamic World Science Citation Center (ISC), ASEAN Citation Index (ACI), Cabell's Directories & Journal Guide.

The publisher of Pertanika will not be responsible for the statements made by the authors in any articles published in the journal. Under no circumstances will the publisher of this publication be liable for any loss or damage caused by your reliance on the advice, opinion or information obtained either explicitly or implied through the contents of this publication. All rights of reproduction are reserved in respect of all papers, articles, illustrations, etc., published in Pertanika. Pertanika provides free access to the full text of research articles for anyone, web-wide. It does not charge either its authors or author-institution for refereeing/publishing outgoing articles or user-institution for accessing incoming articles. No material published in Pertanika may be reproduced or stored on microfilm or in electronic, optical or magnetic form without the written authorization of the Publisher. **Convricht © 2018 Universiti Putra Malavia Press. All Rights Reserved.** 

### Pertanika Journal of Tropical Agricultural Science Vol. 41 (2) May 2018

#### Contents

Foreword	
Nayan Deep S. Kanwal	i
Tribute	
Tributes In Memory of the Life of Professor Emeritus Dr. CHIN Hoong-Fong <i>Chief Executive Editor, Pertanika Journals</i>	505
Review Articles	
Toxicity Effects of Fish Histopathology on Copper Accumulation Siti Nadzirah Padrilah, Mohd Khalizan Sabullah, Mohd Yunus Abd Shukor, Nur Adeela Yasid, Nor Aripin Shamaan and Siti Aqlima Ahmad	519
Mammary Gland Tumours in the Dog, a Spontaneous Tumour Model of Comparative Value to Human Breast Cancer Kabiru Sahabi, Sujey Kumar Rajendren, Jia Ning Foong and Gayathri Thevi Selvarajah	541
Changes in Rice Physiology and Soil Conditions during Low-Water-Input Rice Production System - A Short Review Jahan, M. S.	575
A Mini Review on Phytochemical Constituents and Pharmacological Activities of Adenium obesum Mohamed Shafiq, S., Ling, A. P. K., Lim, C. L., Chye, S. M. and Koh, R. Y.	591
Regular Articles	
Gram-positive Bacteria with Commercial Potential from the Gastrointestines of Holothuria (Mertensiothuria) Leucospilota (Timun Laut) and Stichopus Horrens (Gamat) from Malaysian Waters Kamarul Rahim Kamarudin and Maryam Mohamed Rehan	605
The Analysis of Arbutin in Mao ( <i>Antidesma thwaitesianum</i> Muell. Arg.) Extracts Thongjuta Suwanprasert	621
Rice Ratooning Using the Salibu System and the System of Rice Intensification Method Influenced by Physiological Traits <i>Pinta Omas Pasaribu, Triadiati and Iswandi Anas</i>	637

Effect of Foliar Fertiliser on Banana Noor Asma' Mohd Anuar Mushoddad, Nurul Syaza Abdul Latif and Suhaimi Othman@Osman	655
Effect of Conventional and Superheated Steam Roasting on the Total Phenolic Content, Total Flavonoid Content and DPPH Radical Scavenging Activities of Black Cumin Seeds <i>Liang, L. C., Zzaman, W., Yang, T. A. and Easa, A. M.</i>	663
Effect of Antimicrobial Activities on the Various Solvents Extracts of Leaves of <i>Scurrula Ferruginea</i> (Jack) Danser (Loranthaceae) Vanielie Terrence Justine, Muskhazli Mustafa and Rusea Go	677
Effect of Various Composting Methods on the Concentration and Viability of <i>Ascaris suum</i> Eggs in Organic Fertilisers <i>Arianne L. Andes and Vachel Gay V. Paller</i>	687
Use of Bio-Chemical Surfactant Producing Endophytic Bacteria Isolated from Rice Root for Heavy Metal Bioremediation <i>Arun Karnwal</i>	699
Plant Growth Promoting Rhizobacteria (PGPR) Application with Different Nitrogen Fertilizer Levels in Rice ( <i>Oryza sativa</i> L.) <i>Hamid Ghaffari, Abdollatif Gholizadeh, Abbas Biabani, Alireza</i> <i>Fallah and Mohammad Mohammadian</i>	715
Effects of Non-Medicated and Medicated Urea Molasses Multinutrient Blocks on Dry Matter Intake, Growth Performance, Body Condition Score and Feed Conversion Ratio of Saanen Lactating Does Fed Conventional Diets <i>Mira, P., Wan Zahari, M., Rusli, N. D. and Mat, K.</i>	729 741
Deficit Irrigation for Improving the Postharvest Quality of Lowland	, 11
Tomato Fruits Mohammed, H. N., Mahmud, T. M. M. and Puteri Edaroyati, M. W.	759
Natural Products from Stem Bark of Calophyllum andersonii Keng Hong Tee, Gwendoline Cheng Lian Ee, Ka Woong Wong, Thiruventhan Karunakaran, Vivien Yi Mian Jong and Soek Sin Teh	
Influence of Maternal Dietary Energy and Protein on the Embryonic Development of FUNAAB – Alpha Chickens B. Saleh, S. T. Mbap, D. J. U. Kalla and U. D. Doma	769
Investigative Baseline Reference on the Status of Pork pH, Shear Force, Colour, Drip and Cooking Loss in RYR1 Mutation Free, Commercial 3-way Crosses in Malaysia	777

Michelle-Fong, W. C., Ooi, P. T., Awis, Q. S. and Goh, Y. M.

Development and Validation of an Unsaturated Soil Water Flow Model for Oil Palm <i>Teh, C. B. S.</i>	787
Anther Dehiscence, Pollen Viability and Stigma Receptivity Study on Cultivars of Black Pepper ( <i>Piper nigrum</i> L.) <i>Chen, Y. S., Dayod, M. and Tawan, C. S.</i>	801
Gene Action Mechanism for Drought Tolerance in Extra-Early Yellow Maize Inbreds Shaibu, A. S.	815
Assessment of Soybean Resistance to Whitefly ( <i>Bemisia tabaci</i> Genn.) Infestations Kurnia Paramita Sari and Apri Sulistyo	825
Balance of Nitrogen in Plant-Soil System with the Presence of Compost+Charcoal Erry Purnomo, Franky Sinaga, Indri P Amanda and Riverina DP Putra	833
Technology Assessment of Growing Superior Mungbean (Vigna radiata L.) Varieties on a Dryland in North Lombok I Komang Damar Jaya, Sudirman, Aris Budianto, Abdurachman Hanafi and I Nyoman Soemeinaboedhy	845
Effect of Cytokinins on <i>In Vitro</i> Growth of Hypocotyl and Cotyledon of Tomato ( <i>Lycopersicon esculentum</i> ) Wina Dian Savitri, Popy Hartatie Hardjo, Leonardo Tejo Gunawan Putra Hardianto and Steven Sutanto	855
Effect of Phytase Enzyme on Growth, Nutrient Digestibility and Survival Rate of Catfish ( <i>Pangasius hypothalamus</i> ) Fingerlings Diana Rachmawati and Istiyanto Samidjan	865
Preserving Blue Swimming Crab ( <i>Portunus pelagicus</i> ): Its Conservation using Trap Modifications in Betahwalang, Demak Herry Boesono, Dhian Meita Hapsari, Aristi Dian Purnama Fitri and Kukuh Eko Prihantoko	879
Growth Pattern of Barb ( <i>Barbodes balleroides</i> ) at the Period of Inundation in Jatigede Reservoir, Sumedang Regency, West Java <i>Titin Herawati, Atikah Nurhayati and Sona Yudha Diliana</i>	889
Effect of Proteolytic Plant-Derived Enzyme on Gourami (Osphronemus goramy Lac.) Growth Rate Yuli Andriani, Yeni Mulyani, Irfan Zidni, Muhammad Yusra Sadri and Putra Nur Wicaksono	897

Bioinsecticide Entomopathogenic Nematodes as Biological Control Agent for Sustainable Agriculture Didik Sulistyanto, Ralf-Udo Ehlers and Bachtiar H.Simamora	907
Short Communication	
Prevalence of Avian Polyomavirus in Psittacine Birds in the Klang Valley Z. Zanon, N. Najihah, J. Abu and A. R. Mariatulqabtiah	917



#### **TROPICAL AGRICULTURAL SCIENCE**

Journal homepage: http://www.pertanika.upm.edu.my/

## Effect of Cytokinins on *In Vitro* Growth of Hypocotyl and Cotyledon of Tomato (*Lycopersicon esculentum*)

#### Wina Dian Savitri\*, Popy Hartatie Hardjo, Leonardo Tejo Gunawan Putra Hardianto and Steven Sutanto

Department of Biology, Faculty of Biotechnology, University of Surabaya, Jawa Timur, 60292, Indonesia

#### ABSTRACT

Study of regeneration from different tissues or organs of plants is important as it gives information on how a piece of a plant can transform into its whole form. This process is even substantial when we talk about genetic engineering in plants, since no genetic engineering is valuable without knowing first the standard protocol for regenerating the transformed tissue or organs to become a whole plant. This experiment used hypocotyl and cotyledon of tomato cv. Tymoti as the explants was used to study how different concentrations (1.5-3 ppm) of cytokinins (Kinetin (Kin), 6-benzylaminopurine (BAP), thidiazuron (TDZ) and Zeatin (Zn)) affect its growth. As many as 16 explants were used for each treatment. The growth of both explants in the Murashige and Skoog (MS) media + vitamins showed that Zn and TDZ were superior among the other treatments in inducing calli and primordia organ.

Keywords: Cotyledon, cytokinins, hypocotyls, in vitro growth, tissue regeneration

#### INTRODUCTION

Cytokinins, theoretically, are plant growth regulators (PGRs) that trigger the differentiation of shoots. This PGR is primarily produced in root caps and then distributed into shoots

ARTICLE INFO Article history: Received: 18 September 2017 Accepted: 30 April 2018

*E-mail addresses:* winasavitri@staff.ubaya.ac.id (Wina Dian Savitri) poppy\_hardjo@staff.ubaya.ac.id (Popy Hartatie Hardjo) leonardo.tejogunawan@gmail.com (Leonardo Tejo Gunawan Putra Hardianto) ssutanto2@gmail.com (Steven Sutanto) \*Corresponding author (Aloni et al., 2005). Several kinds of cytokinins have been discovered, namely thidiazuron (TDZ), 6-benzylaminopurine (BAP),  $6-\gamma-\gamma$ -dimethylaminopurine (2-ip), kinetin and zeatin. Among all the cytokinins that have been mentioned above, 2-ip and zeatin are naturally occurring, while the rest are derived synthetically (Razdan, 2002).

Yet, some plant species showed a different responses toward cytokinins. For example, less than 25% up to 50% of callus occurred on muskmelon's cotyledon explants cultured on MS media + vitamins incorporated with 1-2 ppm BAP, although 11.11% (1 out of 9 explants) and 44.44% (4 out of 9 explants) shoots were also produced from 1 ppm BAP and 2 ppm BAP respectively (Ishak, 2015). Our preliminary data on tomato cv. Tymoti showed that 0.5-3 ppm BAP applied on cotyledon and hypocotyls generated low to high callus structure on each explant. Savitri (2015) suggested that cotyledon explants of tomato cv. Tymoti cultured on MS medium + vitamins with the adding of 1-3 ppm BAP in combination with 0.1 ppm TDZ produced not only shoots but also calli that ranged from 18.75-56.25%. In addition, 0.5-2.5 ppm BAP or TDZ mixed with 0.1 ppm indole acetic acid (IAA) yielded 100% callus structure when applied to 10-week-old leaf discs of tomato cv. Tymoti cultured in dark condition (Savitri et al., 2016). Those findings represent that in relatively low concentration, cytokinins could also give rise to callus formation instead of shoot differentiation. Tomato cv. Tymoti is a hybrid that has already been sold commercially. This product is unique because it is suitable to be cultured on lowland, such as in Surabaya. Additionally, this product is resistant to Geminivirus and Pseudomonas solanacearum. This cultivar seems more promising than the others

because it can be planted in lowland, so that the hybrid can be used in the experiment as a sample to learn about the tomato regeneration by in vitro culture.

The current experiment is aimed at studying the effect of four different cytokinins, i.e. TDZ, BAP, kinetin and zeatin on four concentrations; these are 1.5, 2, 2.5 and 3 ppm for each cytokinin. The results could be beneficial to give information about tomato regeneration through indirect pathway. The indirect pathway is very useful to produce a new traits, because the callus can divide verv fast without certain direction. This can lead to cell mutation where some of the daughter cells are different from the parent cell. The ultimate aim of this research is to find a new trait from tomato cv. Tymoti (crop improvements), such as shorter reproduction cycle and greater vields.

#### MATERIALS AND METHODS Plant Materials

The seeds of tomato cv. Tymoti were collected and surface sterilised by double dipping methods using sodium hypochlorite (NaOCl) solution, namely 2.63% (5 minutes) and 1.8% (15 minutes) respectively. These method were followed by rinsing it with sterile distilled water three times. The surface-sterile seeds (10-15) were cultured on ½ MS medium for 14 days. The hypocotyl and cotyledon were collected after that.

#### **Culture Media**

Half strength MS medium was prepared to culture the surface-sterile seeds. Each culture bottle contained 25 mL 1/2 MS medium. MS media + vitamins (Phytotech) were prepared for the treatments. Zeatin (Zn), Thidiazuron (TDZ), Benzylaminopurine (BAP), and Kinetin (Kin), at a concentration of 1.5, 2, 2.5 and 3 ppm respectively were added to the MS media + vitamins. Each bottle contained 25 mL MS medium + vitamins each enriched with cytokinin in a certain concentration. As much as 3% sucrose was added to the media. Before the adding of 1.2% agar, the pH was set at 5.6 for both media. Four cotyledon or hypocotyl were cultured on each culture bottle. Each treatment was repeated four times.

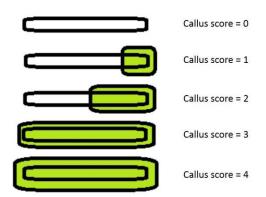
#### **Incubator Condition**

Incubator room was set at 25°C with 80-85% humidity, white fluorescent lamps were used to provide light, approximately equalling to 2000 lux. The photoperiod was regulated at 16 hours light/ 8 hours dark.

#### **Data Analysis**

Data was collected after eight weeks of culture. The callus and shoot formation data were derived from the number of explants that produced callus or shoot, compared with all the explants on each culture bottle and converted into a percentage. Because each treatment was repeated 4 times, percentage average was

used. Data of every explant was noted from the average of callus score (Figure 1) for every 16 explants in each treatment. Data related to friable callus, compact callus, 'friable callus with nodule' and 'compact callus with nodule' were derived from number of callus matched with each type of callus compared with total number of explants that produced callus in each treatment. This data was converted into percentage. Data of 'number of shoots per explant' was calculated from the average number of shoots produced by every 16 explants in each treatment. The Kruskal-Wallis test (Minitab 17) was used to analyse data of 'callus score', 'number of shoots' and comparison between hypocotyl and cotyledon on both data. Correlation coefficients between callus formation (%) vs. callus score and vs. shoot formation (%) were performed using Microsoft Excel 2007 program.



*Figure 1.* Illustration of callus score. 0, no callus formation; 1, quarter of explant formed callus; 2, half of explant formed callus; 3, entire explant formed callus; 4, callus size is twice of the initial explant Green indicates the growth of callus

#### **RESULTS AND DISCUSSION**

#### Effect of Cytokinins on Hypocotyls' Development

Based on Table 1, callus formation on hypocotyls, after being exposed to different kinds and concentrations of cytokinins, ranges from 43.75-100%. The lowest callus formation was produced by 3 ppm Kinetin, while the highest was produced by 1.5-2.5 ppm TDZ and 2.5 ppm Zeatin. This finding shows that TDZ and Zeatin are the best among the treatments. Even though 3 ppm TDZ, 1.5-2 ppm Zeatin, and 3 ppm Zeatin were not the highest, they are still higher among other treatments (93.75%). However, for Kinetin and BAP, callus formation varied between 43.75% and 62.5%.

The callus score is shown in Table 1 while the different letters show the significant differences among the treatments. Callus formation was the highest (93.75%-100%) when hypocotyls is exposed to 2.5 ppm TDZ and 1.5-3 ppm Zn. Given the situation, 2.5 ppm TDZ was chosen because

Table 1Effect of cytokinins on hypocotyl's development

it contributed to the highest shoot formation (31.25%), although the number of shoot per explant was low. This was probably because the explants were not sub-cultured in a new fresh media, as the explants' age was already 8 weeks old when data was collected. The longer the usage of culture medium, the lower the nutrients. There are not enough nutrients on the media to produce more shoots. Moreover, TDZ is much cheaper than Zn. Osman et al. (2010) reported that the 8-week-old hypocotyls and cotyledon tomato explants transferred to  $\frac{1}{2}$  MS + 1 ppm Indole acetic acid (IAA) produced plantlets with fine roots. The experiments also suggested that 0.5-3 ppm TDZ was suitable to produce 5-6 shoots from a cotyledon explant. Razdan (2002) proposed that a low concentration of auxins and cytokinins induce production of shoot and axillary buds while the high levels lead to callus and root formation. Yet in this experiment, a relatively low concentration of cytokinins (1.5-3 ppm) led to callus formation.

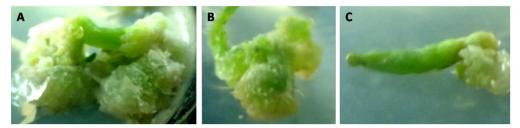
Treatment	Callus	Callus	Friable Callus	Compact	Friable Callus	Compact Callus	Shoot Formation	No. of
(ppm)	Formation (%)	Score <sup>v</sup>	(%)	Callus (%)	with Nodule (%)	with Nodule (%)	(%)	Shoots
BAP 1.5	56.25	1 c <sup>w</sup>	22.22	33.33	0	33.33	25	1×
BAP 2	62.5	1 c	20	50	0	30	18.75	1
BAP 2.5	50	1 c	75	12.5	0	12.5	6.25	0
BAP 3	68.75	1 bc	9.09	36.36	0	54.55	37.5	1
Kin 1.5	43.75	1 c	28.57	57.14	0	14.29	6.25	0
Kin 2	56.25	1 c	0	55.56	11.11	33.33	25	1
Kin 2.5	50	1 c	37.5	50	0	12.5	6.25	0
Kin 3	43.75	1 c	0	28.57	0	71.43	6.25	0
TDZ 1.5	100	3 b	100	0	0	0	6.25	0
TDZ 2	100	4 a	81.25	0	18.75	0	18.75	1
TDZ 2.5	100	3 a	0	0	0	100	31.25	1
TDZ 3	93.75	3 a	0	86.67	0	13.33	18.75	1
Zn 1.5	93.75	2 bc	0	0	0	100	25	1
Zn 2	93.75	3 b	0	0	0	100	25	2
Zn 2.5	100	3 ab	0	0	0	100	25	1
Zn 3	93.75	2 b	0	0	0	100	31.25	1

*Note.* \*Callus Score: 0, no callus formation; 1, quarter of explant formed callus; 2, half of explant formed callus; 3, entire explant formed callus; 4, callus size is twice of the initial explant; \*Mean values with the same letter are not significantly different at  $P \ge 0.05$ ; \*Mean values are not significantly different at  $P \ge 0.05$ .

Pertanika J. Trop. Agric. Sci. 41 (2): 855 - 864 (2018)

Callus scores were used to describe how much calli were formed from a single explant. The scores ranged from 0 to 4. Each score shows the size of callus descriptively from 'no callus formation' to 'the size of callus as twice the initial explant'. This descriptive data was then analysed using the Kruskal-Wallis test after being converted into scores. Figure 2 shows the callus formed from hypocotyls explants. The callus score 4, 3 and 1 are as shown on Figure 2A, 2B and 2C respectively. The nodules that occur on callus indicate the sign of organogenic callus, meaning that it will develop into organ primordia which usually are shoot buds rather than root. Later, the nodules or the organogenic calli will form calli with partial organ regeneration. Ikeuchi et al. (2013) categorised these calli as shooty, rooty and embryonic, based on the adventitious organ's type that regenerated from the callus. The nodules formed from compact callus are shown in Figure 3, while nodules formed from friable callus are described in Figure 4.

There is a positive correlation between callus formation (%) and its score in hypocotyl (Figure 7), and between callus formation (%) and shoot formation (%) (Figure 8). These data indicate that the higher the percentage of callus formation, the higher the callus score and shoot formation.

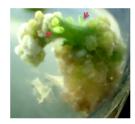


*Figure 2.* Callus formation on the hypocotyls explant as the effect of cytokinins after 8 weeks of culture on MS medium + vitamins. A, 2 ppm TDZ (callus score: 4); B, 3 ppm Zn (callus score: 3); C, 1.5 ppm Kin (callus score: 1)



*Figure 3*. Shoot formation on hypocotyl explant after 8 weeks of culture on MS medium + vitamins enriched with 2 ppm BAP. The arrows show the nodules that later will develop into shoot buds

Wina Dian Savitri, Popy Hartatie Hardjo, Leonardo Tejo Gunawan Putra Hardianto and Steven Sutanto



*Figure 4*. Friable callus with nodules formed on hypocotyl explant after 8 weeks of culture on MS medium + vitamins incorporated with 2 ppm TDZ. The arrows show the nodules that later will develop into shoot buds

## Effect of cytokinins on cotyledon's development

The callus formation in all treatments achieved by cotyledon explants was relatively lower than those by hypocotyl (Table 2). The callus score per explant was also lower than hypocotyl. In terms of shoot growth, 1.5 ppm TDZ, 2.5 Zn and 2.5-3 ppm BAP gave a higher number compared with hypocotyl by 2, 2 and 2-3 number of shoot, respectively. Yet, Table 3 shows there is no significant difference in the number of shoot per explant produced by hypocotyl and cotyledon. This finding is not supported by Wayase and Shitole (2014) on tomato cv. Dhanashri. They concluded that cotyledonary explants were better than hypocotyl in producing shoots. If the statistical data can be ignored, it is likely that 2.5 ppm BAP can be chosen because BAP is cheaper than TDZ and Zn. The BAP is the most commonly used cytokinin (Bhojwani & Dantu, 2013), and TDZ is the most active cytokinin (Huetteman & Preece, 1993). Zeatin is naturally occurring cytokinin in plants (Mok et al., 2002).

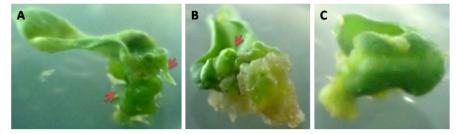
Table 2Effect of cytokinins on cotyledon's development

Treatment	Callus	Callus	Friable	Compact	Friable Callus	Compact Callus	Shoot	No. of
(ppm)	Formation (%)	Score <sup>v</sup>	Callus (%)	Callus (%)	with Nodule (%)	with Nodule (%)	Formation (%)	Shoots
BAP 1.5	25	1×	0	100	0	0	0	0×
BAP 2	56.25	1	11.11	88.89	0	0	0	0
BAP 2.5	31.25	0	40	40	0	20	6.25	2
BAP 3	56.25	1	22.22	33.33	0	44.44	25	3
Kin 1.5	12.5	0	0	50	0	50	12.5	1
Kin 2	12.5	0	0	50	0	50	6.25	2
Kin 2.5	18.75	0	33.33	66.67	0	0	0	0
Kin 3	25	0	0	50	0	50	12.5	2
TDZ 1.5	31.25	1	0	40	0	60	25	2
TDZ 2	37.5	1	0	83.33	0	16.67	0	0
TDZ 2.5	43.75	2	71.42	0	28.57	0	12.5	2
TDZ 3	56.25	1	0	0	0	100	6.25	1
Zn 1.5	31.25	0	0	0	0	100	0	0
Zn 2	6.25	0	0	0	100	0	0	0
Zn 2.5	62.5	1	0	0	20	80	25	2
Zn 3	43.75	1	0	0	14.29	85.71	0	0

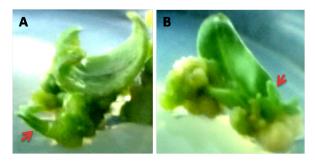
*Note.* <sup>v</sup>Callus Score: 0, no callus formation; 1, quarter of explant formed callus; 2, half of explant formed callus; 3, entire explant formed callus; 4, callus size is twice of the initial explant; "Mean values with the same letter are not significantly different at  $P \ge 0.05$ ; <sup>x</sup>Mean values are not significantly different at  $P \ge 0.05$ .

Pertanika J. Trop. Agric. Sci. 41 (2): 855 - 864 (2018)

Just like the hypocotyl, in cotyledon, four callus types and 5 kinds of callus score (0, 1, 2, 3 and 4) were present. Figure 5A shows compact callus with nodules and Figure 5B shows friable callus with nodules. They both scored 1 and 3 respectively based on the callus size. Figure 5C shows 0 callus score (i.e. no callus is formed on the cotyledon explant). Figure 6A and 6B show the buds on this explant. The correlation between callus formation (%) and shoot formation (%) was also analysed (Figure 7 and 8). As in the cotyledons, correlation between callus formation (%) and callus score is also clearly shown by coefficient correlation (r) 0.67. Furthermore, a lower positive correlation was shown by callus formation (%) versus shoot formation (%) (r = 0.35).



*Figure 5*. Callus formation on the cotyledon explants as the effect of cytokinins after 8 weeks of culture on MS medium + vitamins. A, 1.5 ppm Kin (callus score: 1); B, 3 ppm TDZ (callus score: 2); C, 1.5 ppm Zn (callus score: 0). The arrows show the nodules that later will develop into shoot buds



*Figure 6.* Cotyledon explants formed callus after 8 weeks of culture on MS medium + vitamins. A, Compact callus was produced after exposed to 1.5 ppm BAP; B, Friable callus was produced after being exposed to 2.5 ppm TDZ. The arrows show the shoot buds

Wina Dian Savitri, Popy Hartatie Hardjo, Leonardo Tejo Gunawan Putra Hardianto and Steven Sutanto

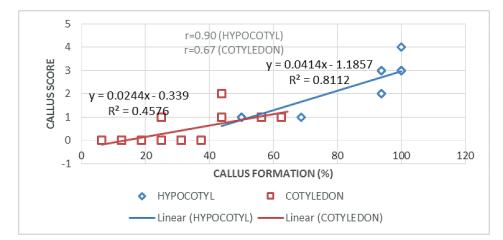


Figure 7. Positive correlation between callus formation (%) and callus score on hypocotyl and cotyledon

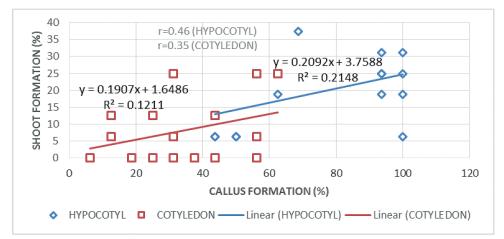


Figure 8. Positive correlation between callus formation (%) and shoot formation (%) on hypocotyl and on cotyledon

#### Comparing the two explants

In terms of shoot production from callus (indirect pathways), cotyledon explants showed better result. This is a common cultivar specific result. Genetic and environmental conditions are two major causes that effect regeneration. Moghaieb et al. (1999) reported the opposite finding, that hypocotyl explants in tomato cv. Pontaroza produced greater number of shoots compared with cotyledons.

Therefore, hypocotyl produces a higher callus formation and a higher callus score per explant. This finding was supported by correlation data between callus formation versus callus score and callus formation (%) versus shoot formation (%), that both showed a positive relationship. The comparison test performed by the Kruskal-Wallis showed no significant difference between two groups of data (Table 3). The additional experiment, such as sub-culturing the incubated explants into fresh medium, is needed to prove that cotyledon produces greater number of shoots than those that are not sub-cultured.

Table 3

Treatment	Callus Score per Explant*	Number of Shoot per Explant*
BAP 1.5	NS**	NS
BAP 2	NS	NS
BAP 2.5	NS	NS
BAP 3	NS	NS
Kin 1.5	NS	NS
Kin 2	Sig	NS
Kin 2.5	NS	NS
Kin 3	NS	NS
TDZ 1.5	Sig	NS
TDZ 2	Sig	NS
TDZ 2.5	NS	NS
TDZ 3	Sig	NS
Zn 1.5	Sig	NS
Zn 2	Sig	NS
Zn 2.5	Sig	NS
Zn 3	Sig	NS

Comparing hypocotyl and cotyledon explants in callus score and number of shoots per explant

*Note.* \* Data of hypocotyl and cotyledon's comparisons were analysed using Kruskal-Wallis Test by significance level of 0.05; \*\* NS: not significantly different; \*\*\*Sig: significantly different.

#### CONCLUSION

BAP, TDZ, Kinetin and Zeatin induced the production of callus on hypocotyl and cotyledon of tomato cv. Tymoti. The shoots were also produced but in a very low percentage because the explants had not been sub-cultured in a new fresh MS medium. There was positive correlation between percentage of callus formation and callus score and shoot formation in both hypocotyl and cotyledon. In spite of the fact there was no significant difference between hypocotyl and cotyledon in producing shoots, using a hypocotyl explant and exposing it to 1.5-3 ppm TDZ or Zeatin may lead to a higher probability in producing callus.

#### ACKNOWLEDGEMENT

Authors would like to thank Lembaga Penelitian dan Pengabdian kepada Masyarakat (LPPM) Ubaya for funding this research, and the Faculty of Biotechnology Ubaya for providing the facilities to complete these experiments.

#### REFERENCES

- Aloni, R., Langhans, M., Aloni, E., Dreieicher, E., & Ullrich, C. I. (2005). Root-synthesized cytokinin in Arabidopsis is distributed in the shoot by the transpiration stream. *Journal of Experimental Botany*, 56(416), 1535–1544.
- Bhojwani, S. S., & Dantu, P. K. (2013). Plant tissue culture: An introductory text. New Delhi, India: Springer.
- Huetteman, C. A., & Preece, J. E. (1993). Thidiazuron: a potent cytokinin for woody plant tissue culture. *Plant Cell, Tissue and Organ Culture, 33*(2), 105 – 119.
- Ikeuchi, M., Sugimoto, K., & Iwase, A. (2013). Plant Callus: Mechanisms of induction and repression. *The Plant Cell*, 25(9), 3159–3173.
- Ishak, J. (2015). Studi regenerasi dan seleksi melon 'action 434' untuk transformasi genetik menggunakan Agrobacterium tumefaciens [Study of regeneration and selection of '434' melon action for genetic transformation using Agrobacterium tumefaciens]. Skripsi (p. 33). Surabaya, Indonesia: Universitas Surabaya.
- Moghaieb, R. E. A., Saneoka, H., & Fujita, K. (1999). Plant regeneration from hypocotyl and cotyledon explant of tomato (*Lycopersicon esculentum* Mill.). Soil Science and Plant Nutrition, 45(3), 639-646.

- Mok, M. C., Martin, R. C., & Mok, D. W. S. (2000). Cytokinins: Biosynthesis, Metabolism and Perception. *In Vitro Cellular & Developmental Biology - Plant*, 36(2), 102 – 107.
- Osman, M. G., Elhadi, E. A., & Khalafalla, M. M. (2010). Callus formation and organogenesis of tomato (*Lycopersicon esculentum* Mill, C.V. Omdurman) induced by thidiazuron. *African Journal of Biotechnology*, 9(28), 4407 – 4413.
- Razdan, M. K. (2002). Introduction to plant tissue culture. Enfield (NH), USA: Science Publishers, Inc.
- Savitri, W. D. (2015). Direct adventitious shoot formation from tomato hypocotyls and cotyledons. In Badruzsaufari, H., Suryajaya, Uripto, T. S., & Rodiansono (Eds.), Proceeding of International Conference on Natural and Environmental Sciences for Sustainable Development 2015 (pp. 77 – 81). Banjarbaru, Indonesia: Fakultas Matematika dan Ilmu Pengetahuan Alam Universitas Lambung Mangkurat.
- Savitri, W. D., Ferina, A. B., Octavia, Y., Muliawan, E., & Effendi, E. E. (2016). Calluses from tomato cv. Tymoti and their morphological characteristics as supporting material for plant tissue culture lesson. In N. Ducha (Ed.) *Prosiding Seminar Nasional Biologi 2016* (pp. 293 – 297). Surabaya, Indonesia: Universitas Negeri Surabaya.
- Wayase, U. R., & Shitole. M.G. (2014). Effect of plant growth regulators on organogenesis in tomato (*Lycopersicon esculentum* Mill.) cv. Dhanashri. *International Journal of Pure and Applied Sciences and Technology*, 20(2), 65–71.

## Pertanika Journal of Tropical Agricultural Science



