Impact of instant-controlled pressure drop treatment on thermal properties and microbial decontamination of banana flour

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Impact of Instant-controlled Pressure Drop Treatment on Thermal Properties and Microbial Decontamination of Banana Flour

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Abstract. Banana flour was produced from unripe banana fruit. During the production process, an instant-controlled pressure drop (IPD) treatment was applied to small slices of banana fruit. The treatment was basically a combination of heat and mechanical treatment. The treatment was performed to investigate its impact on the thermal properties and the microbial decontamination of the banana flour. The analysis of differential scanning calorimetry showed that the IPD treatment resulted in a higher gelatinization temperatures of the banana flour product. The gelatinization temperatures of the IPD treated banana are the initial temperature 12.53 °C, the peak temperature 63.10 °C, and the final temperature 155.41 °C. It was found that IPD treatment was superior in reducing the number of bacteria in the banana. It was also found that the addition of IPD treatment time will reduce the number of microbial colonies in banana.

INTRODUCTION

Banana is a common name encompassing many species in the genus *Musa* from *Musaceae* family. Bananas are plants that can grow easily in tropical and subtropical regions. Banana cultivation requires low investment yet it quickly brings in an income. The banana plantation can be developed in both the lowlands and highlands. In Indonesia, banana cultivation is practiced in almost all rural agricultural areas. Because of its natural characteristics, banana quality will deteriorate quickly during storage. Storage at low temperatures is initially considered as a promising method for extending the durability of the freshly harvested bananas. But apparently, by applying such method, bananas experience chilling injuries, including prevented ripening, when stored at low temperatures [1]. In the ripening period, which is also called as climacteric stage, the biochemical composition of the banana fruit is changed due the occurrence of simultaneous biochemical processes, including degradation of the starch, starch hydrolysis which produces sugar, and synthesis of bioactive molecules e.g., polyphenol compound [2].

Nowadays, there is growing awareness from researchers to develop technology or process that can avoid the losses in the post-harvest period of banana fruit. Producing banana flour from unripe banana fruit is perceived as one of the reliable method to avoid losses in post-harvest. Recently, the use of unripe banana flour as functional food ingredient is growing rapidly. It is because of its attractive nutritional value, especially its rich in indigestible carbohydrates and mainly its high content of resistant starch and dietary fiber. Another advantageous are the banana

flour rich in bioactive compounds, e.g., phenolic compounds, which will act as antioxidant and have a much longer shelf life than the ripe banana fruit [3-6]. Banana flour has been used as one of the ingredients in several commercial food products, including slowly digestible cookies [7]; pasta, especially spaghetti [8]; yoghurt, fruit bar, infant foods, noodles, chips, ice cream, muffins, confectioneries, and jam [9].

Various technology processing have been developed to produce banana flour, including pretreatment using organic acids, i.e., ethylene diamine tetra acetic acid, benzoic acid and sorbic acid, to inhibit browning due to enzyme activity, and other biochemical and microbial degradation during flour production [10]; production of high resistant starch content of unripe banana flour by controlling the process temperature below the gelatinization temperature [11]; agglomeration process in a pulsed-fluidized bed agglomeration to increase the flow ability of the green banana flour product dietary fiber and phytochemical bioactivity enrichment of unripe banana flour by mechanically fractionation [12]; and improvement of functional properties, pasting properties and digestibility of green banana flour by application of heat moisture treatment, annealing and retro gradation [13]. In the present work, an instant-controlled pressure drop (IPD) treatment, which is basically a simultaneous mechanical and heat treatment, is applied on thin slices of unripe banana before the final dehydration process. The goal is to observe the impact of the IPD treatment on the thermal properties, namely its gelatinization temperatures and the microbial decontamination level of the banana flour product.

MATERIALS AND METHODS

Raw Material

Unripe banana (*Musa sp*) was purchased from a local market. Bananas were then peeled and cut into thin slices at 16x16x2 mm mm³. *Rappaport Soy Broth* (RV), *Selenite Cystine Broth* (SC), *Lactose Broth* (LB), *Xylose Lysine Deoxycholate* agar (XLD agar), *Triple Sugar Iron* agar (TSI agar) and *Lysine Iron* agar (LIA agar) were purchased from Merck, Germany; *Brilliant Green* agar (BGA agar) was purchased from Pronadisa, Spain; and Peptone was purchased from Himedia, India.

Dehydration Method

There are two stages of dehydration, namely initial and final dehydration. Both stages of dehydration were carried out on the same device: a convective hot air dryer (Memmert UFE 600). The banana slices were spread in 1-layer above the dryer tray. The air flow, which was maintained at a temperature of 50 °C, flew through the tray creating convective mass and heat transfer. In the initial dehydration stage, the banana slices were dehydrated to reduce the water content up to 20% (wet basis). This initial dehydration was carried out to make the banana slices physically strong enough against the mechanical and heat action experienced during the IPD treatment. Meanwhile, in the final dehydration, the processed banana slices was dehydrated until the moisture content reaches 7% (wet basis) before being crushed into flour. As a comparison, there were samples that were not treated with the IPD, so the dehydration was only completed at one stage. For these samples, the dehydration was carried out using the oven or air-dried outside under the Sun.

Instant-controlled Pressure Drop Treatment

The IPD treatments were carried out using a set of apparatus as shown in Fig. 1 (a). Basically, the apparatus consisted of treatment vessel, pressure drop valve, vacuum tank, vacuum pump and steam boiler. The treatment vessel has 1 liter volume, while the vacuum tank has around 60 fold volume of the treatment vessel to facilitate an instant pressure drop. A certain quantity of sample was placed in a perforated sample holder then it was inserted in the treatment vessel, while the pressure drop valve was closed. Water steamed from the boiler was then injected to the treatment vessel and held at a certain pressure for a few seconds. The pressure drop valve was then suddenly opened and produced an instant pressure drop in the treatment vessel from high pressure to vacuum pressure in less than one second. This cycle was repeated as needed. The pressure fluctuation in the treatment vessel is typically

presented in Fig. 1 (b). In the present work, one cycle of IPD was applied. The pressure of the steam applied in the treatment sample was 2.7 bar, while the treatment time was varied at 10, 20, 30, 40, 50 and 60 seconds. The instant pressure dropped suddenly, bringing down the pressure at the treatment sample from 2.7 bar to 0.005 bar.

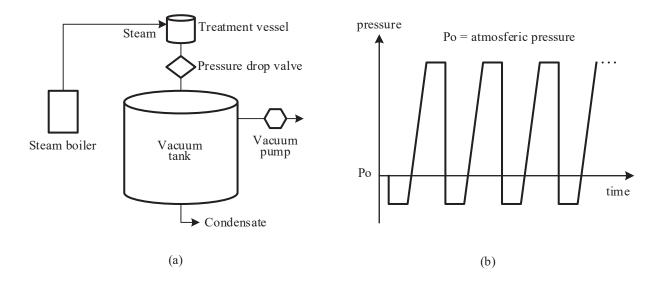


FIGURE 1. (a) The schematic diagram of the apparatus to create instant-controlled pressure drop; (b) Typical pressure fluctuation cycles in the treatment vessel.

Thermal Properties Measurement

The thermal properties were measured using a Differential Scanning Calorimetry (DSC) Mettler Toledo Star System. The measured parameters were the initial, peak, and completion/end set gelatinization temperatures as well as the gelatinization enthalpy.

Determination of the Number of Microbial Colony by Pour Plate Counting

Twenty-five grams of the banana slices were fed to 225 ml of 0.85% w/v NaCl solution and 0.1% peptone. This mixture was blended for approximately 2 (two) minutes to crush the banana into pieces. Subsequently, a series of dilutions was carried out by taking 1 ml of the liquid sample and diluting to 10 ml in a diluents tube. The dilution was continued by taking 1 ml of liquid from the first diluents tube and diluting again to 10 ml in a second diluents tube. Dilution was carried out until the concentration of the sample 10^{-6} times the initial concentration. From each diluents tube, 0.1 ml of liquid was taken and transferred to a petri dish. Furthermore, as much as 10-15 ml of the molten agar medium (plate count agar) at a temperature of $\pm 45^{\circ}$ C was poured into the petri dish that had filled with the sample liquid. Rotate all the plates to achieve a better mixing.

The remaining medium was then poured into an empty cup as a blank. The blank was incubated together with the sample. If there was a growth of microorganisms on the blank, it means that the agar had been contaminated and all results could not be used. After solidifying, the cup was reversed, and incubated at 35°C for 48 hours. After 48 hours incubation, the number of microorganism colonies was calculated. To find out whether there was a fungus on the banana samples, the petri dish was incubated again for 48 hours. The presence of fungal or mold growth was observed after 96 hours incubation.

Isolation and Identification of Salmonella

Twenty-five grams of banana flour samples were fed to 225 ml of *Lactose Broth* (LB) containing pancreatic digestion gelatin, lactose monohydrate, and beef extract. This mixture was continuously stirred and incubated at

35°C for one day. As much as 1 ml of the incubated mixture was taken and dissolved in *Rappaport Soy Broth* (RV) and Selenite Cystine Broth (SC), then incubated at 35°C for one day. Each of incubated RV and SC solution was taken and then scratched on a *Brilliant Green* agar (BGA) and XLD agar using a loop. BGA and XLD media were used as selective media for *Salmonella* bacteria. The results of the scratches were incubated at 35°C for one day. After incubation, the results of this scratch were grouped based on the color of media and colony. Microbes that grew on XLD and BGA were taken using a needle and then inserted into *Triple Sugar Iron*agar (TSI agar) and *Lysine Iron*agar (LIA agar). The TSI agar and LIA agar were incubated at 35°C for one day.

After incubation for 24 hours, an analysis was carried out by observing the color changes in TSI agar and LIA agar. On the TSI agar, the occurrence of glucose, lactose, and/or sucrose fermentation, and production of H₂S was examined. The red color showed the base reaction (B), while the yellow color showed the acid reaction (A). Formation of H₂S was marked by the appearance of black color. Red color (B) on the surface and yellow (A) at the bottom indicated glucose fermentation, but not lactose and sucrose. The yellow color on the surface and bottom showed the occurrence of glucose, lactose and/or sucrose fermentation. The reaction that occurred in TSI agar by Salmonella bacteria was characterized by the occurrence of red color on the slant (inclined agar) and the butt (the base of the inclined agar), and there were black spots on the slant. Whereas in the LIA agar, the occurrence of Salmonella was marked by the change in media from color to red and the appearance of black spots.

RESULTS AND DISCUSSION

The Thermal Properties

Starch, when heated in the presence of excess water, will undergo the phase of transformation from order to disorder arrangement. Such transition is known as gelatinization that will happen over a temperature range, which is typical of starch sources. This phase is associated with diffusion of water into the granule, water uptake by the amorphous background region, hydration and radial swelling of the starch granules, loss of birefringence, loss of crystalline order, uptake of heat, uncoiling and dissociation of double helices in the crystalline regions, and amylose leaching [14]. Gelatinization is followed by pasting. The gelatinization process, which is a function of the starch's water ratio, can be detected by a differential scanning calorimetry (DSC). Initial (To), peak (Tp), and completion/endset (Te) gelatinization temperatures can be obtained from DSC thermograms.

The thermograms of DSC analysis of the banana flour, both without IPD treatment - sun dried sample, without IPD treatment - oven dried sample, and with IPD treatment - oven dried sample, were summarized on Table 1 in terms of the gelatinization temperature and enthalpy. Without IPD treatment - oven dried banana samples are the samples which were conventionally dried in oven at 50°C without insertion of IPD technology during the dehydration process. Meanwhile, without IPD treatment - sun dried banana samples were obtained by exposed the banana slice to the sun outside, and it need three days exposure to reach the desired final moisture content. As expressed on the table, it can be seen that the banana with IPD treatment had higher gelatinization temperatures, with an initial granule gelatinization temperature (To) of 12.53 °C, a peak temperature (Tp) of 63.10 °C and a final temperature (Te) of 155.41 °C. These temperatures are higher than result of banana flour without IPD treatment, i.e.: for sun dried sample 6.28, 47.60 and 110.59 °C. It means that the banana flour with IPD treatment required higher temperatures to ensure complete gelatinization and pasting than two other products. The lower reducing sugar content due to enzyme deactivation during IPD treatment can be a reason of the higher gelatinization temperature. Flour with higher gelatinization temperature is preferred for producing food products in which delayed pasting is desired, such as in retorted canned foods.

Furthermore, banana flour with IPD treatment required more energy to gelatinize (122.95 J/g gelatinization enthalpy) than without IPD - sun dried banana flour (184.16 J/g gelatinization enthalpy) and required less energy to gelatinize than without IPD - oven dried banana flour (235.97 J/g gelatinization enthalpy).

TABLE 1. Gelatinization temperature and enthalpy of banana flour with different treatments

Gelatinization Temperature &Gelatinization Enthalpy	Without IPD - Sun Dried	Without IPD - Oven Dried	with IPD - Oven Dried		
Onset To (°C)	6.28	5.83	12.53		
Peak Tp (°C)	47.60	42.74	63.10		
End set Te (°C)	110.59	105.92	155.41		
Enthalpy (ΔH , J/g)	184.16	235.97	122.95		

Microbial Decontamination

The impact of the IPD treatment in regard with microbial decontamination was investigated by measuring the microbial content in the sample before and after IPD treatment. Total plate count method was used to perform the measurement. The result of microbial content measurement, expressed in CFU (Colony Forming Unit)/25 g of sample, for banana sample with IPD treatment – oven dried, without IPD treatment – oven dried, and without IPD treatment - sun dried were presented on Table 2. For banana with IPD treatment, the treatment time was varied to be 10, 20, 30, 40, 50 and 60 seconds.

TABLE 2. Comparison of the microbial content at different treatment methods

	Microbial Content(CFU/25 g) at Various IPD Treatment Time						
Treatment Method	10 (s)	20 (s)	30 (s)	40 (s)	50 (s)	60 (s)	
With IPD treatment (130 °C -2.7 bar) – Oven Dried	600	200	100	50	0	0	
Without IPD treatment – Oven Dried	16,000						
Without IPD treatment – Sun Dried	2,000						

The table exhibits the effect of treatment time of the IPD technology on the microbial content of the banana flour. The results above show that the longer the IPD treatment time, the lesser the microbial colony number of banana flour. The long contact time of the food material (banana) to the steam increases the efficacy of the IPD technology in regard with microbial decontamination. The table also shows the superiority of the IPD treatment over the two other treatment methods in reducing the food microbial content. The low microbial content of banana flour with IPD treatment would help prolong the shelf time of banana product.

The banana flour obtained from the three applied treatment was analyzed for *Salmonella* identification. This analysis used Xylose Lysine Deoxycholate (XLD) agar selective medium in which *Salmonella* will de-carboxylate lysine to reach neutral pH of 7.4. Over this neutral pH, *Salmonella* can produce H₂S from thiosulphate reduction. The decarboxylation reaction is as follows:

$$C_6H_{14}N_2O_2 \rightarrow C_5H_{14}N_2 + CO_2$$
 (1) (lysine) (cadaverine)

The XLD agar contains three sugars (lactose, sucrose and xylose) and amino acid lysine. Certain bacteria, such as *Salmonella* and *Citrobacter* that are grown on this XLD agar can degrade the sugars to produce acid, which will exhibit color change of indicator from red to yellow. Lysine decarboxylation into cadaverine will cause the medium surrounding the growing colony to be reddish purple because of the increase of pH. The alkaline reaction of this lysine decarboxylation can then be neutralized by the acid produced from xylose metabolism. Normally, in the BGA agar, *Salmonella* cannot ferment sucrose or lactose, and result in red colonies surrounded by red zone.

In the present research, the *Salmonella* identification of the banana flour using both XLD and BG agar show negative results. The 48 hour incubated sample showed yellowish white opaque colonies (Fig. 2 and 3). These colonies may be *E. coli*, *Enterobacter*, *Serratia*, or *Klebsiella*.



FIGURE 2. Colonies on the XLD agar (*Salmonella* negative).

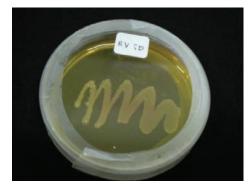


FIGURE 3. Colonies on the BGA agar (*Salmonella* negative).

In order to strengthen the results from XLD and BG agar incubation, the colonies from those selective media were transferred into slant differential agar of *Triple Sugar Iron*agar (TSI agar) and deep *Lysine Iron*agar (LIA agar). Over growing on the TSI medium, *Salmonella* will ferment glucose so that acid is produced at the bottom of the slant agar called butt. This acid production will cause the medium color to change into red. *Salmonella* will also produce H₂S gas, which is marked by black dots on the slant. The result obtained from this analysis shows the yellow colonies on the slant and the butt (Fig. 4). These colonies may be the result of glucose and lactose or sucrose fermentation by *Hafnia*, *Klebsiella*, *Staphilococus*, *Streptococus*, and *Serratia* bacteria. The growth of *Salmonella* on the LIA medium resulted in the color change into red with black dots as a result of lysine decarboxylation activity. In this research, the result of analysis using LIA indicated the medium color change into purple and yellow, which proved that the sample did not contain *Salmonella* (Fig. 5).



FIGURE 4. Result of *Salmonella* identification on the TSI agar (*Salmonella* negative).



FIGURE 5. Result of *Salmonella* identification on LIA agar (*Salmonella* negative).

The results of *Salmonella* identification showed that there were no *Salmonella* colonies found on the banana sample both with and without IPD treatment. Such results may ne also supported by clean environment and negligible load of *Salmonella* on the raw material.

CONCLUSION

The impact of the instant-controlled pressure drop on the melting properties and microbial decontamination of banana flour has been investigated. The analysis of banana flour thermal properties by a differential scanning calorimetry showed that the banana flour with the treatment of instant-controlled pressure drop (IPD) had higher gelatinization temperatures. The banana flour with IPD treatment requires more energy to gelatinize than without IPD treatment and sun dried, and required less energy to gelatinize than without IPD treatment and oven dried. It found the superiority of the instant-controlled pressure drop treatment over those without instant-controlled pressure drop treatment, in terms of reducing the food microbial content. It also found that that the longer the instant-controlled pressure drop treatment duration, the lesser the microbial colony number of banana flour. Furthermore, *Salmonella* identification also indicates that the produced banana flour do not contain any *Salmonella*.

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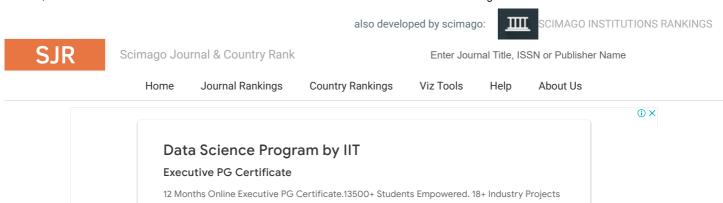
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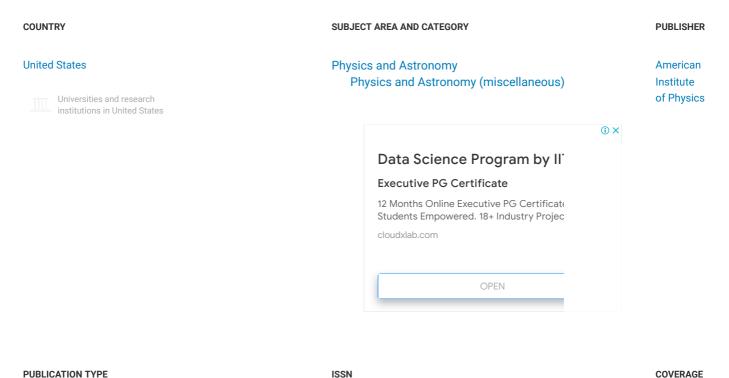


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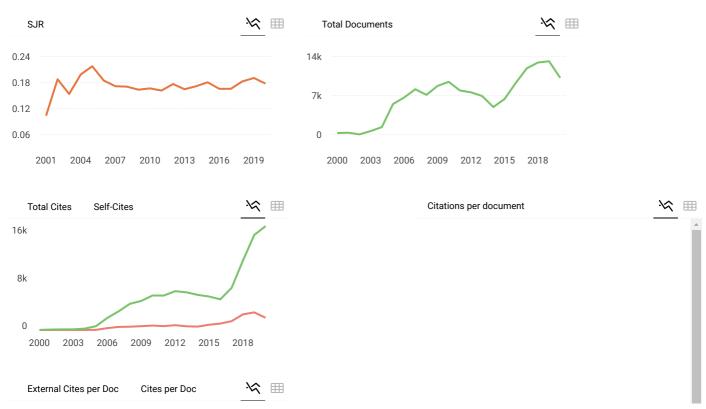
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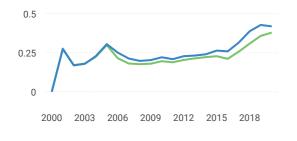
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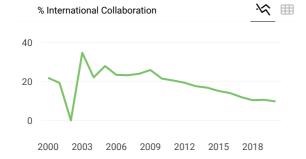
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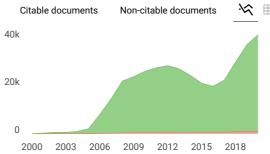


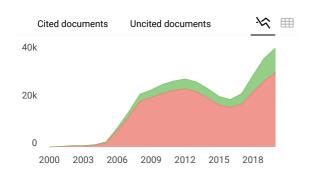
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A Akshya Sekar 2 years ago

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Thanks

reply

Melanie Ortiz 2 years ago

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D **Duha Ahmed** 2 years ago

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https://www.scopus.com/sourceid/26916

I hope the AIP Conference Proceeding is still in the Scopus for 2020 with my best wishes Miss Duha

reply

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Melanie Ortiz 2 years ago

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reply



Melanie Ortiz 2 years ago

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Best Regards, SCImago Team

Thanh Quang Khai Lam 2 years ago

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i don't see in Scimago.

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Teo Jin Chuan 2 years ago

Dear Admin,

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Regards

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H Hassan Abdulhadi 3 years ago

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Best regards

TArik AlOmran

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Best regards,

Budi

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Conference date: 12-13 December 2018

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Editors: Anto Budi Listyawan, Nurul Hidayati, Wisnu Setiawan, Tri Widodo Besar

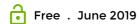
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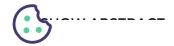


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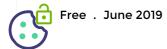
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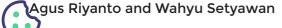
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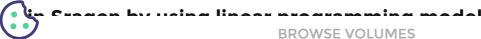
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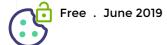


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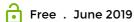
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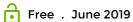
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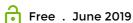
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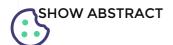


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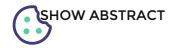


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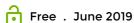
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