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Review

Developmental Origins of Disease - Crisis Precipitates Change

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Key Words

Nutrition • Thrifty phenotype • Developmental programming • Paternal, maternal, sex differences • Epigenetics

Abstract

The concept of developmental origins of diseases has gained a huge interest in recent years and is a constantly emerging scientific field. First observations hereof originated from epidemiological studies, linking impaired birth outcomes to adult chronic, noncommunicable disease. By now there is a considerable amount of both epidemiological and experimental evidence highlighting the impact of early life events on later life disease susceptibility. Albeit far from being completely understood, more recent studies managed to elucidate underlying mechanisms, with epigenetics having become almost synonymous with developmental programming. The aim of this review was to give a comprehensive overview of various aspects and mechanisms of developmental origins of diseases. Starting from initial research foci mainly centered on a nutritionally impaired intrauterine environment, more recent findings such as postnatal nutrition, preterm birth, paternal programming and putative interventional approaches are summarized. The review outlines general underlying mechanisms and particularly discusses mechanistic explanations for sexual dimorphism in developmental programming. Furthermore, novel hypotheses are presented emphasizing a non-mendelian impact of parental genes on the offspring's phenotype.

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Introduction

Throughout the entire life of an individual, environmental factors play an important role for its state of health. However, at no stage in life the surrounding environment has a bigger impact than during embryonic and fetal life. Growth and development in utero are complex

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and dynamic processes which require an orchestration of a variety of maternal, paternal and fetal factors for an optimal outcome. This complex interaction between mother, father, placenta and embryo/fetus ensures an optimal supply of nutrients, oxygen and endocrine signals, all fundamental elements for normal development. Disruptions in this supply system may not only have a direct impact on altering fetal growth patterns, but, as evidence suggests, can be associated with the occurrence of diseases in the later life of the offspring. In the current review, we will discuss exogenous (environmental) as well as endogenous factors (both parental and offspring genes) contributing to the complex interaction between mother, father, placenta and embryo/fetus.

Undernutrition

Epidemiological data unequivocally indicate that there is a connection between early life conditions, anthropometric measures at birth and disease susceptibility in later life [1, 2]. The "Barker Hypothesis", also called the "Fetal Programming Hypothesis" or the theory of the "Developmental Origins of Health and Diseases (DOHaD)", has become the foundation for this increasingly popular scientific field [2]. Barker et al. were not the first investigating this subject, but it was their groundbreaking epidemiological studies in England and Wales in the late 1980ies that inspired research worldwide. Barker et al. initially demonstrated a geographical relationship between cases of ischemic heart disease in the years 1968-1978 and child mortality rates between 1921-1925 [3]. A follow-up study showed that individuals born with a reduced birth weight had an increased risk for coronary heart disease in their adult life [4, 5]. Hales et al. demonstrated in another follow-up study that there is a similar inverse correlation between birth weight and later life glucose tolerance or insulin resistance [6]. In addition, they revealed that individuals with the lowest birth weight, in comparison to heavier newborns, displayed a six fold increased risk for impaired glucose tolerance or diabetes mellitus type 2 in late adulthood [6]. Until now, these findings have been replicated in several different study populations and in different ethnic groups [7]. Based on their observations, Hales and Barker formulated the "Thrifty Phenotype Hypothesis", a more detailed hypothesis trying to outline a putative mechanism of fetal programming. According to this explanation model, gestational under nutrition induces a series of adaptive processes in the fetus, trying to maximize the chances of survival in the given nutrient-poor environment. However, if a mismatch between pre- and postnatal nutrient supply exists fetal adaptation can be deleterious, increasing the risk for diseases later in life [7, 8].

The Thrifty Phenotype Hypothesis

Research stimulated by the thrifty phenotype hypothesis has improved the understanding of the plasticity of early human development, emphasizing an important role of developmental plasticity as a possible contributing factor to later human disease [9]. Until now, the thrifty phenotype hypothesis was confirmed by a number of human studies. The link between a poor intrauterine environment, restricted fetal growth and increased adult disease risk was well demonstrated in the "Dutch famine study". In this retrospective study, children born during a war-inflicted famine between December 1944 and April 1945 were analyzed [10, 11]. During the famine, the daily caloric intake for the general population was restricted to 400 - 800 kcal per day, about half as much as before and after this period. The comparison of individuals that were in utero during this time with individuals born a year before or after the famine showed that gestational caloric restriction was associated with a decreased birth weight and an increased prevalence of impaired glucose tolerance at an age of 50 years [10]. Moreover, twin studies were able to confirm the Thrifty Phenotype Hypothesis. A Danish study examined mono- and dizygotic twins which were discordant for the occurrence of type 2 diabetes. Results of the study revealed that the diabetic



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twin was born with a significantly lower birth weight compared to the euglycemic twin sibling [12]. Other twin studies, especially studies on monozygotic twins, highlighted the importance of the intrauterine environment and developmental plasticity, regardless of the underlying genotype [13]. In addition to the connection between a sub-optimal intrauterine environment, disturbed fetal growth patterns and disease predisposition in adulthood, several more recent studies showed that postnatal nutrition is to be regarded as another critical component of the thrifty phenotype hypothesis. Crowther et al. investigated the impact of postnatal weight gain in a cohort of 7-year-old South African children [14]. Results of this study showed that children born with a reduced birth weight and rapid postnatal weight gain, displayed an impaired glucose tolerance already at an age of 7 years [14]. Further studies in Finland and India replicated these findings [15–17]. Various studies have demonstrated that rapid postnatal weight gain in newborns with an initial low birth weight is mainly due to fat accumulation and not due to an increase in muscle mass [18–20]. This specific phenotype was observed in several cohorts of small for gestational age newborns [21, 22]. In addition it was demonstrated that fat accumulation is more prominent in visceral than in subcutaneous fat depots [23, 24].

Parallel to the epidemiological studies outlined above, various animal studies have been conducted over the years in order to investigate the underlying mechanisms of developmental programming more thoroughly. Preclinical results substantiated findings from observational studies and gave more insight into involved mechanisms, also substantiating the thrifty phenotype hypothesis [25, 26]. It was demonstrated that restricting the maternal diet during gestation does not just result in low birth weight but induces disproportional growth. At the expense of organs such as liver, kidney, pancreas, lung and skeletal muscle, the development of brain, heart and adrenal gland is prioritized [25, 27]. Caloric restriction during gestation was shown to reduce pancreatic beta cell mass formation in the offspring, leading to a decreased production of insulin [28]. In a postnatal calorie-rich life, this lack of insulin production can predispose for the development of diabetes [29].

Overnutrition

As worldwide obesity rates are constantly rising, the focus of research has moved from maternal undernutrition as a predisposing factor for reduced birth weight and adult disease susceptibility, to the impact of maternal overnutrition on offspring health. Interestingly, it was shown that maternal overnutrition and an increased birth weight of the newborn elicits similar effects on offspring health as observed in low birth weight offspring [30, 31]. Being born small for gestational age (SGA) usually is associated with deficits in placental function, placental blood flow and adverse environmental influences, such as maternal undernutrition, particularly if the diet lacks sufficient protein levels [32–35]. Furthermore, literature suggests a complex genetic association, as SGA offspring more commonly occurs in women themselves born SGA [32, 36]. Increases in birth weight are typically associated with maternal obesity and gestational or pre-gestational diabetes [37–39]. Moreover, a very recent study provided genetic evidence for a causal relationship between maternal obesityrelated traits and offspring birth weight [37]. Large for gestational age (LGA) offspring usually displays an increased body fat mass and an increased risk for metabolic disease in later life [37, 40, 41]. Current evidence suggests that either being born with a reduced or an increased birth weight increases disease risk in later life. Meta-analysis have underlined this by demonstrating an U-shaped relationship between later life metabolic diseases and birth weight [42–44]. The overlap of the adult phenotype in SGA and LGA offspring raises the important question which mechanisms are affected in these conditions, and vice versa how these mechanisms can be triggered by conditions producing extremely disparate early life phenotypes [32]. Interestingly, specific overnutrition by feeding an isocaloric highprotein diet to rats, was shown to elicit no effects on birth weight but cause an impaired phenotype in adult animals [45]. Furthermore, it was demonstrated in animal and human



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models that the simultaneous presence of two gestational insults, maternal obesity and maternal stress, are associated with increased rates of both, SGA and LGA offspring [31, 32, 46]. Literature on this subject suggests that the variability in the observed outcomes is likely due to different dosage, timing and downstream effects of a maternal insult. Maternal obesity and pre-pregnancy high-fat intake was on one hand shown to increase the risk for SGA and preeclampsia. On the other hand, this combination of maternal insults was associated with maternal gestational overeating which is associated with gestational diabetes and LGA [30, 32, 47]. Animal stress models revealed that the timing of the maternal stressor during gestation is a key factor for the offspring to be born SGA or LGA [48]. Furthermore, placental development and function, especially an impairment of the placental barrier which normally limits fetal exposure to maternal stress hormones, is thought to play a central role in the severity of maternal stress effects [49, 50]. Taken together, both over and undernutrition are associated with developmental programming. Future studies will give a more precise picture of the complex interaction between nutrition and developmental programming. More recent studies already aimed to discern the role of micronutrient deficiency or excess in regards to developmental origins of disease [51–55]. Furthermore there are also novel approaches to integrate the role of the microbiome and its interaction with nutrition into developmental programming studies [56].

Critical Periods for Nutritional Programming

Experimental and epidemiological data show that effects of developmental programming can be triggered throughout gestation. However, the nature of adult disease can vary according to the timing of a gestational insult [57]. Analysis of the "Dutch Hunger Winter" cohort demonstrated that offspring exposed to famine during early periods of gestation displayed an increased risk for coronary heart disease in later life, which could not be observed if famine exposure happened during later stages of pregnancy [58]. Interestingly, caloric restriction during late gestation was associated with disturbances in glucose-insulin homeostasis, clinically reflected by an increased risk of type 2 diabetes [58]. This finding could be replicated in animal models [59, 60]. Gardner et al. showed that maternal undernutrition in sheep in late gestation also led to impaired glucose-insulin homeostasis, highlighting the importance of late gestational periods in regards to programming effects on intermediary metabolism [60]. According to developmental processes in respective gestational periods, current literature suggests that nutritional insults during early gestation may influence organ development, altering fetal physiology in late gestation, and postnatal function, yet often without a measurable effect on birth weight [59, 61-64]. This was demonstrated by various studies investigating nutritional alteration in the periconceptual period, which is characterized by fertilisation, blastocystogenesis and the implantation process [61–65]. Nutritional insults set in later embryonic and early fetal life, a period which comprises intense organogenesis and placental development, display similar patterns of developmental programming [66–70]. Dietary modifications in later stages of gestation, characterized by very pronounced fetal growth and placental maturation, were shown to have a strong impact on birth weight and organ maturation with pronounced programming effects on intermediary metabolism and hormonal systems [71–75]. Furthermore, also postnatal nutrition plays an important role in developmental programming. A large body of studies have demonstrated that postnatal catch-up growth in low birth weight newborns is a crucial factor for increasing the risk of adult metabolic disturbances [20, 22, 76]. Intriguingly, also in postnatal developmental programming timing is of importance. It was shown that catch-up growth restricted to the first postnatal year did not have an effect on insulin levels, but sustained catch-up growth was associated with higher insulin levels in seven year old- and insulin resistance in eight year old children [77–79]. Next to postnatal periods, newer evidence highlights the importance of preconceptual nutrition of both the mother and the father on offspring health.



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There is a growing amount of evidence, demonstrating that dietary challenges during oozyte development or spermatogenesis can induce permanent phenotypic changes in the offspring [80–84]. Taken together, available evidence highlights the importance of different time frames before, during and after gestation in regards to developmental origins of health and disease. However, given species differences in physiology, metabolism, placental structure and function, cautious interpretation of the available studies is warranted, especially when extrapolating to human situations [72].

Prematurity

More recent data showed that fetal adaptation causing persistent functional and structural changes of the fetal organism can be induced by factors that go beyond gestational nutrition and do not necessarily have to impact on anthropometric measures at birth [85]. A very important factor in this regard is preterm birth. As outlined before, initial studies on fetal programming focused on a 'deprived' intrauterine environment as a cause for low birth weight or SGA [10, 86]. Such anthropometric measurements were then used as surrogate parameters for association analyses with later life disease [10, 86]. However, many of these epidemiologic studies based their investigations on old birth records, sometimes assessing birth weight without considering gestational age at birth [87, 88]. Thus, it is possible that a considerable amount of individuals included in these epidemiologic studies, were preterm individuals and not small for gestational age [87]. De Jong et al. demonstrated in a systematic literature review followed by a meta-analysis, that preterm birth is associated with higher blood pressure in adulthood, suggesting a relevant role of gestational age in fetal programming [89]. There is an increasing body of evidence that prematurity is associated with an increased risk for various disease in adult life [53, 87, 89-91]. It is known that preterm birth causes an interruption of normal organogenesis, especially in organ systems that display a branching morphogenesis like kidney, lung, pancreas, and the vascular system [87, 92–94]. The developing kidney is particularly vulnerable to preterm birth which causes considerable deficits in organ structure and function [93]. Prematurity was shown to be associated with a lower nephron endowment [95, 96] potentially increasing the risk of hypertension, proteinuria and kidney disease in later life [97, 98]. Although underlying mechanisms of increasing adult disease susceptibility by prematurity on first glance seem to be more direct, there are similarities between being born SGA and preterm. Preterm birth cannot be simply seen as an abrupt termination of gestation, but rather as a pathologic, stressful and inflammatory event, influenced by numerous factors, ranging from ethnicity and socioeconomic status to dysfunctions in hormonal systems and gestational micronutrient deficiencies [53, 88, 99–101]. Similar to SGA infants preterm infants suffer from an adverse intrauterine environment and, by being born prematurely, are additionally exposed to an adverse neonatal environment [102]. Both, SGA and preterm born infants display similar postnatal growth patterns with about 80% of both groups exhibiting catch up growth [102]. Resembling observations in SGA newborns, prematurity predisposes to childhood adiposity, with data indicating a shift in adipose tissue distribution towards visceral fat depots [102]. Furthermore, similar to observations in SGA cohorts, a more rapid postnatal catch-up growth was shown to be associated with greater reductions in insulin sensitivity [103, 104]. A recent systematic review and meta-analysis showed that there is also an association between preterm birth and insulin sensitivity throughout life. However, the data in this regard are conflicting and observed associations might be affected by the overall heterogeneity of the study designs and analyzed populations [105]. Albeit conflicting findings, prematurity putatively increases the risk for insulin resistance which, at least in part, appears to be regulated by postnatal growth. This highlights the importance of an optimal nutritional strategy for preterm infants which yet remains to be determined [104, 106].



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The Role of Insulin in Fetal Development and Adult Diseases

The developmental origins theory can be applied to all early life events including low birth weight and/or prematurity [107–109]. Adverse environmental exposures during fetal and neonatal life are thought to trigger compensatory persistent physiological responses. Such adaptations may modify set points of physiological systems involved in sustaining homeostasis. This can become maladaptive if a mismatch between anticipated and actual environment occurs. In this regard, a lot of research was focused on insulin resistance as the main culprit for both, altered anthropometric measurements at birth, and later life disease susceptibility. Divergent to the developmental origins theory first postulated by Barker et al., Hattersley and Tooke proposed in their "fetal insulin hypothesis" that genetically determined insulin resistance results in impaired insulin-mediated growth in the fetus as well as in an insulin resistant phenotype in adult life [110]. It is known that type 2 diabetes has a strong genetic component. Furthermore, insulin acts as key factor in fetal growth. Thus, any genetic variant that impairs insulin secretion and/or insulin sensitivity may reduce birth weight and concomitantly result in adult life type 2 diabetes. Put differently, the "fetal insulin hypothesis" postulated that the genotype, not low birth weight, increases the risk of adult diabetes [110, 111]. The hypothesis is supported by genetic evidence showing that single nucleotide polymorphisms associated with an increased risk for type 2 diabetes were associated with low birth weight [112]. Moreover, a study in Caucasian mothers revealed that there is a negative correlation between total glycated hemoglobin in cord blood (fetal) and birth weight [113]. The relationship between cord blood glycemia and birth weight is diametrically opposed to the well described positive correlation between maternal glycemia and birth weight which was also observed in this study [113]. When subjected to similar degrees of maternal glycemia (reflected by maternal total glycated hemoglobin), lighter fetuses appear incapable of lowering their blood glucose concentrations (reflected by the newborn's total glycated hemoglobin), as do heavier fetuses. Meanwhile, the findings of an inverse relationship between cord blood glycemia and birth weight were replicated in an Asian cohort, highlighting their validity [114]. Until now, fetal blood glucose concentrations were regarded as a passive reflection of maternal glycemia. However, the observed inverse correlation between cord blood and birth weight showed that the fetal response to similar maternal glucose levels might not behave as uniform as previously thought [113, 114]. From a hypothetical point of view such findings can be explained by both, genetics and the fetal environment, underlining that future research, integratively applying genetic and epigenetic methodology, is still needed to better characterize the association between early life and adult disease susceptibility.

Epigenetics

Although the underlying molecular mechanisms are incompletely understood so far, there is convincing evidence that developmental plasticity is mediated by epigenetic modifications of the DNA. Important epigenetic mechanisms are histone modifications, non coding RNAs and DNA methylation [115, 116]. These tools generally affect how accessible DNA is to transcription factor complexes, how efficiently transcription proceeds, and how stable already transcribed mRNA is [104]. Histone modifications consist of chemical alterations such as acetylation, phosphorylation and methylation [116] which can modulate chromatin structure, thus influencing the accessibility of the transcription machinery to the gene [117]. Non coding RNAs can trigger RNAse activity by RNA interference eliminating mRNA transcribed by target genes [38, 118]. The currently best studied epigenetic mechanism is DNA methylation [119]. DNA methylation is the addition of a methyl group at the C5 position of the cytosine pyrimidine ring via DNA methyl transferase activity [120]. Methylated cytosines are generally located in cytosine-phosphate-guanine (CpG) sequences [121, 122]. Although about 70% of all genomic CpGs are methylated, there are clusters of



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CpGs, termed CpG islands, that remain unmethylated [102, 120, 123]. Such unmethylated CpG islands are associated with about 60% of all human genomic promoters [124]. Methylating a CpG site attracts methyl-binding proteins that trigger chromatin remodeling, leading to a more condensed chromatin, thus restricting access for the transcription machine [125]. Therefore, promoter regions of translated genes usually display low methylated CpG islands, whereas un-translated genes are heavily methylated.

In mammalian development, there are two main periods characterized by extensive epigenetic modifications. During the course of gametogenesis, genome-wide demethylation takes place followed by remethylation before fertilization. In early phases of embryogenesis, extensive epigenetic modifications occur, with phases of total demethylation alternating with phases of remethylation, ensuring the totipotency of the developing embryo [126]. Additionally, de- and remethylation processes after fertilization are thought to play a role in the removal of acquired epigenetic modifications, especially those acquired during gametogenesis [127-129]. However, some parental epigenetic modifications seem to escape the second wave of demethylation, underlining a potential inheritance of epigenetic modifications set during gametogenesis [102, 130].

Epigenetic mechanisms are not only important in early phases of pregnancy but throughout gestation [131, 132]. Current literature suggests that epigenetic modifications acquired during early developmental phases can be permanent [133]. It was demonstrated in a variety of experimental models and clinical studies of fetal programming that environmental conditions during gestation or shortly after birth can induce epigenetic alterations, stably changing the degree of promoter methylation and thereby permanently altering gene expression [54, 133]. Rat offspring of dams fed a low-protein diet during pregnancy exhibit decreases in promoter methylation of the glucuronid receptor and the peroxisome proliferator-activated receptor α (PPAR- α) gene in the liver [134]. Similar epigenetic changes were shown for p53 in the kidney [135], the suprarenal angiotensin II type-1b receptor [136], and for the hypothalamic glucocorticoid receptor [137]. More recent data underlined that an alteration of DNA methylation triggered by maternal undernutrition is not tissue specific but a global phenomenon, associated with widespread changes of gene expression [138]. It is not exactly known yet for how long the time window for stable epigenetic changes is opened, but current evidence suggests that the timeframe spans from conceptional to early postnatal stages [137, 139]. It has also been demonstrated that DNA methylation patterns can be transmitted from one generation to the following [140]. Moreover, it was shown that a gestational low-protein diet fed to F0 dams can still alter promoter methylation and gene expression of the F2 generation without any nutrient restriction in the F1 generation [141]. Another study even described a significant impact of a gestational/lactational low protein diet administered only to the F0 generation on the phenotype of F3 generation offspring [142].

Paternal Programming

Until now, the focus regarding fetal programming was mostly set on maternal programming. However, there is accumulating evidence that the father also plays a relevant role in epigenetic modifications of the offspring's phenotype [143, 144]. Epidemiological data showed that the grandchildren of men, who were exposed to a restricted caloric intake during the slow growth phase just before reaching puberty, lived significantly longer than grandchildren of men, who experienced overnutrition during this phase [82]. In a more detailed analysis of these data, it was demonstrated that an excess caloric intake of the paternal grandfather was associated with a fourfold increased risk of dying from diabetes associated disease in the grandchildren's generation [83]. There is also data from animal experiments confirming an influence in terms of fetal programming on the offspring. In a study by Anderson et al. it was shown that paternal fasting before mating was associated with reduced serum glucose levels in the F1 generation [145]. Ng et al. were able to demonstrate



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that a preconceptional high fat diet of the father causes beta cell dysfunction of the pancreas in the F1 generation [146]. Apart from dietary influences, Bakke et al. demonstrated in a pioneer study that hypothyroidism of male rats before mating resulted in significant phenotypic changes of the F1 generation [147]. Paternal hypothyroidism was induced either by radiothyroidectomy or by large doses of neonatally injected thyroxine. Offspring of hypothyroid fathers displayed a slower postnatal development, reduced weaning weights and increased final body weights, and had enlarged pituitary and thyroid glands. Furthermore, female offspring from thyroidectomized fathers developed significantly smaller uteri and enlarged ovaries, whereas testes of male offspring were significantly smaller [147].

Sex Differences in Developmental Origins of Disease

The existence of sex specific differences in animal models of developmental programming is well described in currently available literature. The vast majority of non communicable diseases, which in many cases have developmental origins, often display some degree of sex bias [148]. Most developmental programming studies have shown that the same stimulus can elicit different long term effects, depending on the sex of the offspring. The underlying mechanism of this sexual dimorphism is not well understood [149]. It was demonstrated that gene expression shows sex specific differences, which are already detectable in the preimplanted embryo, long before any gonadal development and sex hormone production [150–152]. Thus, such early phenotypic differences can only be attributed to transcriptional differences resulting from different sex chromosomes, i.e. to Y-chromosomal genes and X-chromosomal genes that to a smaller or bigger extent escape X-chromosome inactivation [150]. Moreover, it was shown that sex chromosomal differences in gene expression can influence the transcription of autosomal genes, resulting in prominent sex specific transcriptional differences [150]. Analysis of bovine blastocysts demonstrated that one third of genes, most of them of autosomal origin, displays sex specific differences in expression [153]. Mechanistically, the imprinting of X-linked genes may be involved in sex specific expression differences. In females specific imprinting ensures that the paternal allele is uniquely or preferentially expressed. As male embryos are missing the paternally inherited X-chromosome, synergistic effects of double X dosage plus imprinting mechanisms may be responsible for sex specific transcriptional differences [150]. Early gestational sexual dimorphism in protein expression may influence several molecular pathways, including glucose and protein metabolism and impact on epigenetic mechanisms, particularly DNA methylation. This might be one underlying reason for a sex specific different susceptibility to environmental stressors, leading to distinct long-term effects in the offspring [150]. Furthermore, there is evidence in literature that the fetal sex as a major genetic variant of the fetal genome may influence maternal physiology during gestation in genetically susceptible pregnant women. It was demonstrated that depending on fetal sex certain maternal genetic variants (ACE I/D; PPARG2 Pro 12 Ala; PROGINS progesteron receptor polymorphism) are associated with different outcomes in regards to maternal glycemia and blood pressure regulation, both very influential factors in fetal development [154–156]. Another crucial factor for sexual dimorphism in developmental programming is the placenta [148, 149, 151]. Being the functional link between the maternal environment and the fetus, the placenta plays a central role as a buffer for environmental effects and is capable of modulating effects of adverse intrauterine conditions [151]. As this organ derives from embryonic trophoblast cells, it bears the same sex as the embryo/fetus [151]. Depending on the sex of the fetus, the placenta displays sexual dimorphism, with different growth rates and a varying responsiveness to fetal hormones [157]. In many species male placentas usually are larger or distinctively shaped, an observation that, at least in mice, seems to be independent of androgen effects [151, 158]. More importantly, current literature suggests that female and male placentas are characterized by different molecular mechanisms to optimize the outcome of the offspring, with distinct transcriptomes, perfectly



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shaped for a proper development of the given sex [151, 159]. Concomitantly, this results in a different susceptibility to perturbations in the intrauterine environment and the ability to cope with them. The molecular mechanisms underlying this sexually distinct adaptive responses are largely unknown, however, current data indicates that the sex specific genome and epigenome are key factors [150, 151]. Finally, regarding disease susceptibility in later life, the impact of sex hormones during development and over the course of life has to be taken into account. However, only a few studies so far have examined the contribution of sex hormones in later life disease susceptibility due to developmental programming. Ojeda et al. demonstrated in a rat model of placental insufficiency that intrauterine growth retardation (IUGR) was associated with hypertension in male offspring [160, 161]. Furthermore, serum testosterone levels were twofold higher in IUGR males than in healthy controls, indicating a connection to the observed hypertension in male IUGR offspring. Castration at an age of 10 weeks abolished hypertension in male IUGR offspring with intrauterine growth retardation. No effects of castration on blood pressure were observed in healthy controls [161]. Female growth retarded offspring also developed hypertension, however this increase in blood pressure returned to normotensive values once the animals reached puberty and displayed increasing levels of estradiol [162]. Ovariectomy at an age of 10 weeks blunted this decrease in blood pressure compared to intact IUGR females. In a third group that received 17β-estradiol replacement, ovariectomy induced increases in blood pressure were attenuated [162]. Results from these and similar studies indicate that sex hormones can influence the long term consequences of developmental programming [161–163]. However, until now there is a lack of studies that evaluated this matter in different animal models with other outcomes than hypertension in a similar extensive fashion as Ojeda et al. Taken together, sexual dimorphism is tightly connected to developmental programming. The influence of sex hormones and differences in placental function are important factors in this regard. Moreover, disparities in the sex specific genome and epigenome, leading to a transcriptional sexual dimorphism which is already present in the preimplanting embryo, may play a relevant role in the varying susceptibility to environmental stressors among the sexes.

Interaction of Parental Genes, Parental Environment and Fetal Programming

As outlined before, maternal and paternal environmental factors can influence the phenotype of the offspring by inducing epigenetic adaptive mechanisms. Another factor responsible for developmental programming during intrauterine life might be related to parental genes that impact on the fetal phenotype independent of their presence in the fetal genome [164, 165]. About 25 years ago, Parkhurst et al. described a wimp mutation in Drosophila that resulted in a lethal phenotype, although the mutation was not transmitted to the offspring [166]. Hocher et al. translated this finding to mammalian/human development. They showed that a single nucleotide polymorphism (SNP) in the maternal G protein beta3subunit gene, which is involved in regulation of blood supply to the uterus, is associated with a substantial reduction in birth weight without actually being transmitted to the offspring (Fig. 1) [167]. Other studies later demonstrated similar independent associations between specific maternal genes and offspring phenotype without any transmission of the particular gene [168–179]. It was shown that maternal mutations of relevant genes involved in folate metabolism are associated with an increased risk for neural tube or congenital heart defects [170–172]. Similar findings were demonstrated for maternal polymorphisms involved in glucose metabolism [173]. Such drastic teratogenic consequences highlight the possible impact maternal genetic deficiencies can have, regardless of any transmission to the offspring. Not as drastic alterations on the offspring phenotype, that better fit to the concept of the developmental origins hypothesis were observed for maternal polymorphisms in the monoamine oxidase A, the peroxisome proliferator activated receptor gamma and cytochrome P enzyme genes controlling sex steroid biosynthesis and metabolism [174-176]. Additionally, it was demonstrated that the maternal genotype plays an important role



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Fig. 1. The Advanced Fetal Programming Hypothesis. (A) Maternal gene dysfunction may influence ovary function. Physiological ovarian function, oocyte development and maturation, including the establishment of epigenetic patterns could potentially be affected by altered maternal gene function [193-196]. (B) Maternal gene dysfunction can impact on the placenta and the embryonic/ fetal environment by altering decidual function [181, 182, 197-199]. (C) Maternal gene dysfunction may alter weaning behavior and lactation performance and thus influence the early postnatal environment [200-203]. (D) The impact of maternal gene dysfunction on embryonal/fetal/neonatal environmental factors listed in A-C may trigger stable, long lasting epigenetic adaptation in the offspring or result in developmental toxicity [204-206]. Regardless of the underlying mechanism, the phenotype of the offspring may be modified by altered maternal gene function without any transmission of the affected maternal gene.



in modifying adverse intrauterine conditions [180]. Studies showed that polymorphisms of genes encoding for xenobiotic metabolizing enzymes, such as *GSTT1*, and *GSTM1* gene or phase I/phase II enzymes such as *CYP1A1* or *EPHX1* can elicit a modifying effect on birth weight among actively or passively smoking mothers [177–179]. Taken together, there is an increasing amount of evidence indicating that parental genes may influence offspring physiology independent of the inheritance of these genes.

In a very recent study, Hocher et al. aimed to better characterize this biological phenomenon in an animal experiment. Therefore, female heterozygous endothelial nitric oxide synthase (eNOS) knockout mice were mated with male wildtype mice and their wildtype offspring was compared to wildtype offspring from wildtype mice. A heterozygous knock out in the *eNOS* gene was chosen because of the central role *eNOS* plays in controlling vascular and placental function, negatively affecting the interuterine environment [181, 182]. The partial lack of the maternal *eNOS* gene resulted in a reduced birth weight and a steatotic liver phenotype of wildtype offspring. Sex specific differences regarding the phenotype were observed [183]. Using a similar study design, Costantine et al. had previously also demonstrated a sex specific transgenerational effect of a maternal heterozygous eNOS deficiency on the vascular phenotype of wildtype offspring [184]. Results of both studies suggest a non-environmentally mediated mechanism of developmental programming driven by altered parental gene function. Without any transmittance of the specific gene, maternal and putatively also paternal gene dysfunction might influence oocyte or sperm maturation and later embryonic and fetal development by the induction of epigenetic modifications or developmental toxicity, finally resulting in an altered phenotype ("The Advanced Fetal Programming Hypothesis"; see Fig. 1) [165, 183]. Next to the general implications of these findings for the field of developmental programming, they also suggest to reassess murine knock out or transgenic animal models, one of the most important tools currently used in studying gene function. The presumed causality between a genetic manipulation and a resulting phenotype should be reconsidered in regards to potential confounding by



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an induction of epigenetic changes due to parental gene dysfunction that is absent in the offspring [183].

Interventional Approaches

Some studies already investigated if it is possible to ameliorate deleterious effects of fetal programming by interventional approaches. Lillycrop et al. showed in a rat model of fetal programming that a high-protein diet of pregnant dams lead to a hypomethylation of different genes in the offspring. However, if folate was supplemented simultaneous to the low protein diet, the previously observed epigenetic modifications were absent [134]. Until now these or similar results were observed by other studies [138, 185]. It was demonstrated that the DNA methylation machinery relies on ingested methyl group donors and other essential micronutrients [185]. A restriction of these factors may have far-reaching consequences on the phenotype of offspring [139, 185].

An interesting explanation model of epigenetic modifications due to environmental influences provides the "Free Radical Theory of Developement" [186, 187]. This theory is based on the biochemical link between redox buffer systems, such as glutathione, and the methyl group metabolism. It was demonstrated that unfavorable intrauterine conditions, triggered for example by gestational protein restriction, can impact on the capacity of redox buffer systems and expose the organism to increased oxidative stress [186]. As an opposing measure, the production of glutathione can be increased which requires methyl group metabolites. This increased demand for methyl group donors can result in a reduced availability of the essential methyl group donator S-adenosylmethionine (SAM), thus affecting epigenetic mechanisms including DNA and histone methylation [186]. Camboine et al. were able to show in a protein restriction model that the simultaneous feeding of a lipidperoxidation inhibitor during pregnancy and lactation is able to prevent the effects of the low protein diet on blood pressure regulation, vascular function and microvascular rarefaction and counteracts a reduction of glutathione [188]. Similar results were generated employing other antioxidants [189–192]. In summary, nutritional interventions during pregnancy affecting the methyl group metabolism might be able to prevent or alter a developmentally programmed phenotype. Future studies are needed to assess, whether such approaches could be translatable therapeutic options targeting the developmental origins of diseases.

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Disclosure Statement

The authors have nothing to disclose.

References

1 Gillman MW: Prenatal famine and developmental origins of type 2 diabetes. Lancet Diabetes Endocrinol 2015;3:751–752.



Cell Physiol Biochem 2016;39:919-938 DOI: 10.1159/000447801 © 2016 The Author(s). Published by S. Karger AG, Basel and Biochemistry Published online: August 12, 2016 www.karger.com/cpb

- 2 Olsen J: David Barker (1938-2013) -- a giant in reproductive epidemiology. Acta Obstet Gynecol Scand 2014:93:1077-1080.
- 3 Barker DJ, Osmond C: Infant mortality, childhood nutrition, and ischaemic heart disease in England and Wales. Lancet 1986;1:1077-1081.
- Osmond C, Barker DJP, Winter PD, Fall CHD, Simmonds SJ: Early growth and death from cardiovascular 4 disease in women. BMJ 1993;307:1519-1524.
- 5 Barker DJ, Osmond C, Simmonds SJ, Wield GA: The relation of small head circumference and thinness at birth to death from cardiovascular disease in adult life. BMJ 1993;306:422.
- 6 Hales CN, Barker DJ, Clark PM, Cox LJ, Fall C, Osmond C, Winter PD: Fetal and infant growth and impaired glucose tolerance at age 64. BMJ 1991;303:1019.
- 7 Hales CN, Barker DJ.: The thrifty phenotype hypothesis: Type 2 diabetes. Br Med Bull 2001;60:5.
- 8 Hales CN, Barker DJP: Type 2 (non-insulin-dependent) diabetes mellitus: the thrifty phenotype hypothesis. Diabetologia 1992;35:595-601.
- 9 Lindsay RS, Bennett PH: Type 2 diabetes, the thrifty phenotype – an overview. Br Med Bull 2001;60:21–32.
- 10 Ravelli ACJ, van der Meulen JH., Michels RPJ, Osmond C, Barker DJP, Hales CN, Bleker OP: Glucose tolerance in adults after prenatal exposure to famine. Lancet 1998;351:173-177.
- 11 Ravelli AC, van Der Meulen JH, Osmond C, Barker DJ, Bleker OP: Obesity at the age of 50 y in men and women exposed to famine prenatally. Am J Clin Nutr 1999;70:811-816.
- 12 Poulsen P, Vaag AA, Kyvik KO, Jensen DM, Beck-Nielsen H: Low birth weight is associated with NIDDM in discordant monozygotic and dizygotic twin pairs. Diabetologia 1997;40:439-446.
- 13 Poulsen P, Esteller M, Vaag A, Fraga MF: The Epigenetic Basis of Twin Discordance in Age-Related Diseases. Pediatr Res 2007;61:38R-42R.
- 14 Crowther NJ, Cameron N, Trusler J, Gray IP: Association between poor glucose tolerance and rapid post natal weight gain in seven-year-old children. Diabetologia 1998;41:1163-1167.
- Forsen T, Eriksson J, Tuomilehto J, Reunanen A, Osmond C, Barker D: The fetal and childhood growth of 15 persons who develop type 2 diabetes. Ann Intern Med 2000;133:176.
- 16 Yajnik C: Interactions of perturbations in intrauterine growth and growth during childhood on the risk of adult-onset disease. Proc Nutr Soc 2008;59:257-265.
- 17 Eriksson JG, Forsen T, Tuomilehto J, Osmond C, Barker DJP: Early adiposity rebound in childhood and risk of type 2 diabetes in adult life. Diabetologia 2003;46:190–194.
- 18 Gale CR: Intrauterine Programming of Adult Body Composition. J Clin Endocrinol Metab 2001;86:267–272.
- Law CM, Barker DJ, Osmond C, Fall CH, Simmonds SJ: Early growth and abdominal fatness in adult life. J 19 Epidemiol Community Health 1992;46:184-186.
- 20 Okada T, Takahashi S, Nagano N, Yoshikawa K, Usukura Y, Hosono S: Early postnatal alteration of body composition in preterm and small-for-gestational-age infants: implications of catch-up fat. Pediatr Res 2015;77:136-142.
- 21 Jornayvaz FR, Selz R, Tappy L, Theintz GE: Metabolism of oral glucose in children born small for gestational age: evidence for an impaired whole body glucose oxidation. Metabolism 2004;53:847-851.
- 22 Ibanez L: Early Development of Adiposity and Insulin Resistance after Catch-Up Weight Gain in Small-for-Gestational-Age Children. J Clin Endocrinol Metab 2006;91:2153-2158.
- 23 Harrington TAM, Thomas EL, Frost G, Modi N, Bell JD: Distribution of Adipose Tissue in the Newborn. Pediatr Res 2004;55:437-441.
- 24 Modi N, Thomas EL, Harrington TAM, Uthaya S, Doré CJ, Bell JD: Determinants of Adiposity during Preweaning Postnatal Growth in Appropriately Grown and Growth-Restricted Term Infants. Pediatr Res 2006:60:345-348.
- 25 Desai M, Crowther NJ, Lucas A, Hales CN: Organ-selective growth in the offspring of protein-restricted mothers. Br J Nutr 1996;76:591-603.
- Hoppe CC, Evans RG, Bertram JF, Moritz KM: Effects of dietary protein restriction on nephron number in 26 the mouse. Am J Physiol Regul Integr Comp Physiol 2007;292:R1768-1774.
- 27 Kikuchi T, Uchiyama M: Epidemiological studies of the developmental origins of adult health and disease in Japan: a pediatric perspective in present day Japan. Clin Pediatr Endocrinol 2010;19:83–90.
- Bertin E, Gangnerau M-N, Bellon G, Bailbé D, Arbelot De Vacqueur A, Portha B: Development of beta-cell 28 mass in fetuses of rats deprived of protein and/or energy in last trimester of pregnancy. Am J Physiol Regul Integr Comp Physiol 2002;283:R623-630.



Cell Physiol Biochem 2016;39:919-938 DOI: 10.1159/000447801 © 2016 The Author(s). Published by S. Karger AG, Basel and Biochemistry Published online: August 12, 2016 www.karger.com/cpb

Reichetzeder et al.: Developmental Origins of Disease

- 29 Inadera H: Developmental origins of obesity and type 2 diabetes: molecular aspects and role of chemicals. Environ Health Prev Med 2013;18:185-197.
- Alfaradhi MZ, Ozanne SE: Developmental programming in response to maternal overnutrition. Front Genet 30 2011;2:27.
- Heerwagen MJR, Miller MR, Barbour LA, Friedman JE: Maternal obesity and fetal metabolic programming: 31 a fertile epigenetic soil. Am J Physiol Regul Integr Comp Physiol 2010;299:R711-722.
- 32 Grissom NM, Reyes TM: Gestational overgrowth and undergrowth affect neurodevelopment: similarities and differences from behavior to epigenetics. Int J Dev Neurosci 2013;31:406-414.
- 33 Godfrey KM, Inskip HM, Hanson MA: The long-term effects of prenatal development on growth and metabolism. Semin Reprod Med 2011;29:257-265.
- 34 Myatt L: Placental adaptive responses and fetal programming. J Physiol 2006;572:25–30.
- 35 Miao Z, Chen M, Wu H, Ding H, Shi Z: Comparative proteomic profile of the human placenta in normal and fetal growth restriction subjects. Cell Physiol Biochem 2014;34:1701–1710.
- Sydsjö G: Long-term consequences of non-optimal birth characteristics. Am J Reprod Immunol 36 2011;66:81-87.
- 37 Tyrrell J, Richmond RC, Palmer TM, Feenstra B, Rangarajan J, Metrustry S, Cavadino A, Paternoster L, Armstrong LL, De Silva NMG, Wood AR, Horikoshi M, Geller F, Myhre R, Bradfield JP, Kreiner-Møller E, Huikari V, Painter JN, Hottenga J-J, Allard C, Berry DJ, Bouchard L, Das S, Evans DM, Hakonarson H, Hayes MG, Heikkinen J, Hofman A, Knight B, Lind PA, McCarthy MI, McMahon G, Medland SE, Melbye M, Morris AP, Nodzenski M, Reichetzeder C, Ring SM, Sebert S, Sengpiel V, Sørensen TIA, Willemsen G, de Geus EJC, Martin NG, Spector TD, Power C, Järvelin M-R, Bisgaard H, Grant SFA, Nohr EA, Jaddoe VW, Jacobsson B, Murray JC, Hocher B, Hattersley AT, Scholtens DM, Davey Smith G, Hivert M-F, Felix JF, Hyppönen E, Lowe WL, Frayling TM, Lawlor DA, Freathy RM, Early Growth Genetics (EGG) Consortium: Genetic Evidence for Causal Relationships Between Maternal Obesity-Related Traits and Birth Weight. JAMA 2016;315:1129-1140.
- 38 Shi Z, Zhao C, Long W, Ding H, Shen R: Microarray Expression Profile Analysis of Long Non-Coding RNAs in Umbilical Cord Plasma Reveals their Potential Role in Gestational Diabetes-Induced Macrosomia. Cell Physiol Biochem 2015;36:542-554.
- Li J, Song L, Zhou L, Wu J, Sheng C, Chen H, Liu Y, Gao S, Huang W: A MicroRNA Signature in Gestational 39 Diabetes Mellitus Associated with Risk of Macrosomia. Cell Physiol Biochem 2015;37:243-252.
- 40 Gillman MW, Rifas-Shiman S, Berkey CS, Field AE, Colditz GA: Maternal Gestational Diabetes, Birth Weight, and Adolescent Obesity. Pediatrics 2003;111:e221-e226.
- 41 Boney CM: Metabolic Syndrome in Childhood: Association With Birth Weight, Maternal Obesity, and Gestational Diabetes Mellitus. Pediatrics 2005;115:e290-e296.
- 42 Schellong K, Schulz S, Harder T, Plagemann A: Birth weight and long-term overweight risk: systematic review and a meta-analysis including 643,902 persons from 66 studies and 26 countries globally. PLoS ONE 2012;7:e47776.
- 43 Curhan GC, Willett WC, Rimm EB, Spiegelman D, Ascherio AL, Stampfer MJ: Birth Weight and Adult Hypertension, Diabetes Mellitus, and Obesity in US Men. Circulation 1996;94:3246-3250.
- 44 Curhan GC, Chertow GM, Willett WC, Spiegelman D, Colditz GA, Manson JE, Speizer FE, Stampfer MJ: Birth Weight and Adult Hypertension and Obesity in Women. Circulation 1996;94:1310-1315.
- Thone-Reineke C, Kalk P, Dorn M, Klaus S, Simon K, Pfab T, Godes M, Persson P, Unger T, Hocher B: 45 High-protein nutrition during pregnancy and lactation programs blood pressure, food efficiency, and body weight of the offspring in a sex-dependent manner. Am J Physiol Regul Integr Comp Physiol 2006;291:R1025-R1030.
- 46 Djelantik AA, Kunst AE, van der Wal MF, Smit HA, Vrijkotte TGM: Contribution of overweight and obesity to the occurrence of adverse pregnancy outcomes in a multi-ethnic cohort: population attributive fractions for Amsterdam. BJOG 2012;119:283-290.
- 47 Rajia S, Chen H, Morris MJ: Maternal overnutrition impacts offspring adiposity and brain appetite markersmodulation by postweaning diet. J Neuroendocrinol 2010;22:905-914.
- 48 Calegare BFA, Fernandes L, Tufik S, D'Almeida V: Biochemical, biometrical and behavioral changes in male offspring of sleep-deprived mice. Psychoneuroendocrinology 2010;35:775-784.

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Cell Physiol Biochem 2016;39:919-938 DOI: 10.1159/000447801 © 2016 The Author(s). Published by S. Karger AG, Basel and Biochemistry Published online: August 12, 2016 www.karger.com/cpb

Reichetzeder et al.: Developmental Origins of Disease

- Togher KL, Togher KL, O'Keeffe MM, O'Keeffe MM, Khashan AS, Khashan AS, Gutierrez H, Gutierrez H, Kenny 49 LC, Kenny LC, O'Keeffe GW, O'Keeffe GW: Epigenetic regulation of the placental HSD11B2 barrier and its role as a critical regulator of fetal development. Epigenetics 2014;9:816-822.
- 50 Li J, Wang Z-N, Chen Y-P, Dong Y-P, Shuai H-L, Xiao X-M, Reichetzeder C, Hocher B: Late gestational maternal serum cortisol is inversely associated with fetal brain growth. Neurosci Biobehav Rev 2012;36:1085-1092.
- 51 Gernand AD, Schulze KJ, Stewart CP, West Jr KP, Christian P: Micronutrient deficiencies in pregnancy worldwide: health effects and prevention. Nat Rev Endocrinol 2016;12:274-289.
- 52 Chen Y, Wang G, Wang X, Ma Z, Chen Y, Chuai M, von Websky K, Hocher B, Yang X: Effects of high salt-exposure on the development of retina and lens in 5.5-day chick embryo. Cell Physiol Biochem 2014;34:804-817.
- Reichetzeder C, Chen H, Föller M, Slowinski T, Li J, Chen Y-P, Lang F, Hocher B: Maternal vitamin D 53 deficiency and fetal programming--lessons learned from humans and mice. Kidney Blood Press Res 2014;39:315-329.
- Junge KM, Bauer T, Geissler S, Hirche F, Thürmann L, Bauer M, Trump S, Bieg M, Weichenhan D, Gu L, Mallm 54 J-P, Ishaque N, Mücke O, Röder S, Herberth G, Diez U, Borte M, Rippe K, Plass C, Hermann C, Stangl GI, Eils R, Lehmann I: Increased vitamin D levels at birth and in early infancy increase offspring allergy risk evidence for involvement of epigenetic mechanisms. J Allergy Clin Immunol Pract 2016;137:610-613.
- 55 Vanhees K, Vonhögen IGC, van Schooten FJ, Godschalk RWL: You are what you eat, and so are your children: the impact of micronutrients on the epigenetic programming of offspring. Cell Mol Life Sci 2014;71:271-285.
- 56 Ganu RS, Harris RA, Collins K, Aagaard KM: Early Origins of Adult Disease: Approaches for Investigating the Programmable Epigenome in Humans, Nonhuman Primates, and Rodents. ILAR J 2012;53:306–321.
- 57 Sinclair KD, Singh R: Modelling the developmental origins of health and disease in the early embryo. Theriogenology 2007;67:43–53.
- Roseboom T, de Rooij S, Painter R: The Dutch famine and its long-term consequences for adult health. Early 58 Hum Dev 2006;82:485-491.
- 59 Gardner DS, Pearce S, Dandrea J, Walker R, Ramsay MM, Stephenson T, Symonds ME: Peri-implantation undernutrition programs blunted angiotensin II evoked baroreflex responses in young adult sheep. Hypertension 2004;43:1290-1296.
- Gardner DS, Tingey K, Van Bon BWM, Ozanne SE, Wilson V, Dandrea J, Keisler DH, Stephenson T, Symonds 60 ME: Programming of glucose-insulin metabolism in adult sheep after maternal undernutrition. Am J Physiol Regul Integr Comp Physiol 2005;289:R947-954.
- Wynn M, Wynn A: Nutrition around conception and the prevention of low birthweight. Nutr Health 61 1988;6:37-52.
- Kwong WY, Wild AE, Roberts P, Willis AC, Fleming TP: Maternal undernutrition during the preimplantation 62 period of rat development causes blastocyst abnormalities and programming of postnatal hypertension. Development 2000;127:4195-4202.
- 63 Edwards LJ, McMillen IC: Periconceptional nutrition programs development of the cardiovascular system in the fetal sheep. Am J Physiol Regul Integr Comp Physiol 2002;283:R669-679.
- 64 McEvoy TG, Robinson JJ, Aitken RP, Findlay PA, Robertson IS: Dietary excesses of urea influence the viability and metabolism of preimplantation sheep embryos and may affect fetal growth among survivors. Anim Reprod Sci 1997;47:71-90.
- Kwong WY, Miller DJ, Ursell E, Wild AE, Wilkins AP, Osmond C, Anthony FW, Fleming TP: Imprinted gene 65 expression in the rat embryo-fetal axis is altered in response to periconceptional maternal low protein diet. Reproduction 2006;132:265-277.
- Vonnahme KA, Hess BW, Hansen TR, McCormick RJ, Rule DC, Moss GE, Murdoch WJ, Nijland MJ, Skinner 66 DC, Nathanielsz PW, Ford SP: Maternal undernutrition from early- to mid-gestation leads to growth retardation, cardiac ventricular hypertrophy, and increased liver weight in the fetal sheep. Biol Reprod 2003;69:133-140.
- Dunford LJ, Sinclair KD, Kwong WY, Sturrock C, Clifford BL, Giles TC, Gardner DS: Maternal protein-energy 67 malnutrition during early pregnancy in sheep impacts the fetal ornithine cycle to reduce fetal kidney microvascular development. FASEB J 2014;28:4880-4892.

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Cell Physiol Biochem 2016;39:919-938 DOI: 10.1159/000447801 © 2016 The Author(s). Published by S. Karger AG, Basel and Biochemistry Published online: August 12, 2016 www.karger.com/cpb

- Lloyd LJ, Foster T, Rhodes P, Rhind SM, Gardner DS: Protein-energy malnutrition during early gestation 68 in sheep blunts fetal renal vascular and nephron development and compromises adult renal function. J Physiol 2012;590:377-393.
- Joles JA, Sculley DV, Langley-Evans SC: Proteinuria in aging rats due to low-protein diet during mid-69 gestation. J Dev Orig Health Dis 2010;1:75-83.
- 70 Gilbert JS, Lang AL, Grant AR, Nijland MJ: Maternal nutrient restriction in sheep: hypertension and decreased nephron number in offspring at 9 months of age. J Physiol 2005;565:137–147.
- 71 Kind KL, Moore VM, Davies MJ: Diet around conception and during pregnancy--effects on fetal and neonatal outcomes. Reprod Biomed Online 2006;12:532-541.
- 72 Harding JE: The nutritional basis of the fetal origins of adult disease. Int J Epidemiol 2001;30:15–23.
- 73 Stein AD, Zybert PA, van de Bor M, Lumey LH: Intrauterine famine exposure and body proportions at birth: the Dutch Hunger Winter. Int J Epidemiol 2004;33:831-836.
- 74 Gao F, Liu Y, Li L, Li M, Zhang C, Ao C, Hou X: Effects of maternal undernutrition during late pregnancy on the development and function of ovine fetal liver. Anim Reprod Sci 2014;147:99-105.
- 75 Bloomfield FH, Oliver MH, Giannoulias CD, Gluckman PD, Harding JE, Challis JRG: Brief undernutrition in late-gestation sheep programs the hypothalamic-pituitary-adrenal axis in adult offspring. Endocrinology 2003;144:2933-2940.
- Ong KK., Ahmed ML, Emmett PM, Preece MA, Dunger DB: Association between postnatal catch-up growth 76 and obesity in childhood: prospective cohort study. BMJ 2000;320:967.
- 77 Crowther NJ, Cameron N, Trusler J, Toman M, Norris SA, Gray IP: Influence of Catch-up Growth on Glucose Tolerance and β-Cell Function in 7-Year-Old Children: Results From the Birth to Twenty Study. Pediatrics 2008;121:e1715-e1722.
- 78 Ong KK, Petry CJ, Emmett PM, Sandhu MS, Kiess W, Hales CN, Ness AR, Dunger DB, ALSPAC study team: Insulin sensitivity and secretion in normal children related to size at birth, postnatal growth, and plasma insulin-like growth factor-I levels. Diabetologia 2004;47:1064-1070.
- 79 Jiang X, Ma H, Wang Y, Liu Y: Early life factors and type 2 diabetes mellitus. J Diabetes Res 2013;2013:485082.
- 80 Watkins AJ, Wilkins A, Cunningham C, Perry VH, Seet MJ, Osmond C, Eckert JJ, Torrens C, Cagampang FRA, Cleal J, Gray WP, Hanson MA, Fleming TP: Low protein diet fed exclusively during mouse oocyte maturation leads to behavioural and cardiovascular abnormalities in offspring. J Physiol 2008;586:2231-2244.
- 81 McPherson NO, Owens JA, Fullston T, Lane M: Preconception diet or exercise intervention in obese fathers normalizes sperm microRNA profile and metabolic syndrome in female offspring. Am J Physiol Endocrinol Metab 2015;308:E805-E821.
- 82 Bygren LO, Kaati G, Edvinsson S: Longevity determined by paternal ancestors' nutrition during their slow growth period. Acta Biotheor 2001;49:53–59.
- 83 Kaati G, Bygren LO, Edvinsson S: Cardiovascular and diabetes mortality determined by nutrition during parents' and grandparents' slow growth period. Eur J Hum Genet 2002;10:682-688.
- 84 Ashworth CJ, Toma LM, Hunter MG: Nutritional effects on oocyte and embryo development in mammals: implications for reproductive efficiency and environmental sustainability. Philos Trans R Soc Lond B Biol Sci 2009;364:3351-3361.
- 85 Hocher B: Fetal programming of cardiovascular diseases in later life - mechanisms beyond maternal undernutrition. J Physiol 2007;579:287-288.
- Barker DJ, Osmond C, Golding J, Kuh D, Wadsworth ME: Growth in utero, blood pressure in childhood and 86 adult life, and mortality from cardiovascular disease. BMJ 1989;298:564-567.
- Abitbol CL, Rodriguez MM: The long-term renal and cardiovascular consequences of prematurity. Nat Rev 87 Nephrol 2012;8:265-274.
- 88 Chen Y-P, Lu Y-P, Li J, Liu Z-W, Chen W-J, Liang X-J, Chen X, Wen W-R, Xiao X-M, Reichetzeder C, Hocher B: Fetal and maternal angiotensin (1-7) are associated with preterm birth. J Hypertens 2014;32:1833-1841.
- 89 de Jong F, Monuteaux MC, van Elburg RM, Gillman MW, Belfort MB: Systematic review and meta-analysis of preterm birth and later systolic blood pressure. Hypertension 2012;59:226-234.
- 90 Parkinson JRC, Hyde MJ, Gale C, Santhakumaran S, Modi N: Preterm birth and the metabolic syndrome in adult life: a systematic review and meta-analysis. Pediatrics 2013;131:e1240-1263.
- 91 Carmody JB, Charlton JR: Short-term gestation, long-term risk: prematurity and chronic kidney disease. Pediatrics 2013;131:1168-1179.



Cell Physiol Biochem 2016;39:919-938 DOI: 10.1159/000447801 © 2016 The Author(s). Published by S. Karger AG, Basel and Biochemistry Published online: August 12, 2016 www.karger.com/cpb

Reichetzeder et al.: Developmental Origins of Disease

- 92 Shi W, Bellusci S, Warburton D: Lung development and adult lung diseases. Chest 2007;132:651–656.
- Shah MM, Sampogna RV, Sakurai H, Bush KT, Nigam SK: Branching morphogenesis and kidney disease. 93 Development 2004;131:1449-1462.
- 94 Peixoto FO, Pereira-Terra P, Moura RS, Carvalho-Dias E, Correia-Pinto J, Nogueira-Silva C: The role of ephrins-B1 and -B2 during fetal rat lung development. Cell Physiol Biochem 2015;35:104–115.
- 95 Keijzer-Veen MG, Devos AS, Meradji M, Dekker FW, Nauta J, van der Heijden BJ: Reduced renal length and volume 20 years after very preterm birth. Pediatr Nephrol 2010;25:499-507.
- Bacchetta J, Harambat J, Dubourg L, Guy B, Liutkus A, Canterino I, Kassaï B, Putet G, Cochat P: Both 96 extrauterine and intrauterine growth restriction impair renal function in children born very preterm. Kidney Int 2009;76:445-452.
- 97 Brenner BM, Garcia DL, Anderson S: Glomeruli and blood pressure. Less of one, more the other? Am J Hypertens 1988;1:335-347.
- Luyckx VA, Brenner BM: Low birth weight, nephron number, and kidney disease. Kidney Int Suppl 98 2005:S68-77.
- 99 Koullali B, Oudijk MA, Nijman T a. J, Mol BWJ, Pajkrt E: Risk assessment and management to prevent preterm birth. Semin Fetal Neonatal Med 2016;21:80-88.
- 100 Gotsch F, Gotsch F, Romero R, Erez O, Vaisbuch E, Kusanovic JP, Mazaki-Tovi S, Kim SK, Hassan S, Yeo L: The preterm parturition syndrome and its implications for understanding the biology, risk assessment, diagnosis, treatment and prevention of preterm birth. J Matern Fetal Neonatal Med 2009;2:5-23.
- 101 Swaggart KA, Pavlicev M, Muglia LJ: Genomics of Preterm Birth. Cold Spring Harb Perspect Med 2015;5:a023127.
- 102 Cutfield WS, Hofman PL, Mitchell M, Morison IM: Could Epigenetics Play a Role in the Developmental Origins of Health and Disease? Pediatr Res 2007;61:68R-75R.
- 103 Regan FM, Cutfield WS, Jefferies C, Robinson E, Hofman PL: The impact of early nutrition in premature infants on later childhood insulin sensitivity and growth. Pediatrics 2006;118:1943-1949.
- 104 Lane RH: Fetal programming, epigenetics, and adult onset disease. Clin Perinatol 2014;41:815–831.
- 105 Tinnion R, Gillone J, Cheetham T, Embleton N: Preterm birth and subsequent insulin sensitivity: a systematic review. Arch Dis Child 2014;99:362-368.
- 106 Finken MJJ, Keijzer-Veen MG, Dekker FW, Frölich M, Hille ETM, Romijn JA, Wit JM, Dutch POPS-19 Collaborative Study Group: Preterm birth and later insulin resistance: effects of birth weight and postnatal growth in a population based longitudinal study from birth into adult life. Diabetologia 2006;49:478-485.
- 107 Barker DJP: The origins of the developmental origins theory. J Intern Med 2007;261:412-417.
- 108 Barker DJP: Fetal programming of coronary heart disease. Trends Endocrinol Metab 2002;13:364–368.
- Sullivan MC, Hawes K, Winchester SB, Miller RJ: Developmental origins theory from prematurity to adult 109 disease. J Obstet Gynecol Neonatal Nurs 2008;37:158–164.
- 110 Hattersley AT, Tooke JE: The fetal insulin hypothesis: an alternative explanation of the association of low birthweight with diabetes and vascular disease. Lancet 1999;353:1789-1792.
- 111 Calkins K, Devaskar SU: Fetal Origins of Adult Disease. Curr Probl Pediatr Adolesc Health Care 2011;41:158-176.
- 112 Freathy RM, Bennett AJ, Ring SM, Shields B, Groves CJ, Timpson NJ, Weedon MN, Zeggini E, Lindgren CM, Lango H, Perry JRB, Pouta A, Ruokonen A, Hyppönen E, Power C, Elliott P, Strachan DP, Järvelin M-R, Smith GD, McCarthy MI, Frayling TM, Hattersley AT: Type 2 diabetes risk alleles are associated with reduced size at birth. Diabetes 2009;58:1428-1433.
- 113 Pfab T, Slowinski T, Godes M, Halle H, Priem F, Hocher B: Low birth weight, a risk factor for cardiovascular diseases in later life, is already associated with elevated fetal glycosylated hemoglobin at birth. Circulation 2006;114:1687-1692.
- Li J, Wang ZN, Schlemm L, Pfab T, Xiao XM, Chen YP, Hocher B: Low birth weight and elevated head-toabdominal circumference ratio are associated with elevated fetal glycated serum protein concentrations. J Hypertens 2011;29:1712-1718.
- 115 Bird A: Perceptions of epigenetics. Nature 2007;447:396–398.
- 116 Bannister AJ, Kouzarides T: Regulation of chromatin by histone modifications. Cell Res 2011;21:381–395.
- Vieira-Coimbra M, Henrique R, Jerónimo C: New insights on chromatin modifiers and histone post-117 translational modifications in renal cell tumours. Eur J Clin Invest 2015;45:16–24.
- 118 Djupedal I, Ekwall K: Epigenetics: heterochromatin meets RNAi. Cell Res 2009;19:282–295.

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Cell Physiol Biochem 2016;39:919-938 DOI: 10.1159/000447801 © 2016 The Author(s). Published by S. Karger AG, Basel and Biochemistry Published online: August 12, 2016 www.karger.com/cpb

- 119 Zhu J: DNA methylation and hepatocellular carcinoma. J Hepatobiliary Pancreat Surg 2006;13:265–273.
- 120 Robertson KD, A.Jones P: DNA methylation: past, present and future directions. Carcinogenesis 2000;21:461-467.
- 121 Gardiner-Garden M, Frommer M: CpG islands in vertebrate genomes. J Mol Biol 1987;196:261–282.
- 122 Takai D, Jones PA: Comprehensive analysis of CpG islands in human chromosomes 21 and 22. Proc Natl Acad Sci USA 2002;99:3740-3745
- 123 Du X, Han L, Guo A-Y, Zhao Z: Features of methylation and gene expression in the promoter-associated CpG islands using human methylome data. Comp Funct Genomics 2012;2012:598987.
- 124 Ginno PA, Lott PL, Christensen HC, Korf I, Chédin F: R-loop formation is a distinctive characteristic of unmethylated human CpG island promoters. Mol Cell 2012;45:814-825.
- 125 Nikitina T, Shi X, Ghosh RP, Horowitz-Scherer RA, Hansen JC, Woodcock CL: Multiple modes of interaction between the methylated DNA binding protein MeCP2 and chromatin. Mol Cell Biol 2007;27:864-877.
- 126 Messerschmidt DM, Knowles BB, Solter D: DNA methylation dynamics during epigenetic reprogramming in the germline and preimplantation embryos. Genes Dev 2014;28:812-828.
- Monk D: Germline-derived DNA methylation and early embryo epigenetic reprogramming: The selected 127 survival of imprints. Int J Biochem Cell Biol 2015;67:128-138.
- 128 Pickard B, Dean W, Engemann S, Bergmann K, Fuermann M, Jung M, Reis A, Allen N, Reik W, Walter J: Epigenetic targeting in the mouse zygote marks DNA for later methylation: a mechanism for maternal effects in development. Mech Dev 2001;103:35-47.
- 129 Reik W, Dean W, Walter J: Epigenetic reprogramming in mammalian development. Science 2001;293:1089-1093.
- 130 Schaefer S, Nadeau JH: The genetics of epigenetic inheritance: modes, molecules, and mechanisms. Q Rev Biol 2015;90:381-415.
- 131 Kim K, Friso S, Choi S-W: DNA methylation, an epigenetic mechanism connecting folate to healthy embryonic development and aging. J Nutr Biochem 2009;20:917–926.
- 132 Liu L, Zhang X, Rong C, Rui C, Ji H, Qian Y, Jia R, Sun L: Distinct DNA methylomes of human placentas between pre-eclampsia and gestational diabetes mellitus. Cell Physiol Biochem 2014;34:1877–1889.
- 133 Attig L, Gabory A, Junien C: Early nutrition and epigenetic programming: chasing shadows. Curr Opin Clin Nutr Metab Care 2010;13:284-293.
- 134 Lillycrop KA, Phillips ES, Jackson AA, Hanson MA, Burdge GC: Dietary Protein Restriction of Pregnant Rats Induces and Folic Acid Supplementation Prevents Epigenetic Modification of Hepatic Gene Expression in the Offspring. J Nutr 2005;135:1382-1386.
- 135 Pham TD, MacLennan NK, Chiu CT, Laksana GS, Hsu JL, Lane RH: Uteroplacental insufficiency increases apoptosis and alters p53 gene methylation in the full-term IUGR rat kidney. Am J Physiol Regul Integr Comp Physiol 2003;285:R962-970.
- 136 Bogdarina I, Welham S, King PJ, Burns SP, Clark AJL: Epigenetic Modification of the Renin-Angiotensin System in the Fetal Programming of Hypertension. Circ Res 2007;100:520-526.
- 137 Weaver ICG, Cervoni N, Champagne FA, D'Alessio AC, Sharma S, Seckl JR, Dymov S, Szyf M, Meaney MJ: Epigenetic programming by maternal behavior. Nat Neurosci 2004;7:847-854.
- 138 Altobelli G, Bogdarina IG, Stupka E, Clark AJL, Langley-Evans S: Genome-wide methylation and gene expression changes in newborn rats following maternal protein restriction and reversal by folic acid. PLoS ONE 2013:8:e82989.
- 139 Sinclair KD, Allegrucci C, Singh R, Gardner DS, Sebastian S, Bispham J, Thurston A, Huntley JF, Rees WD, Maloney CA, Lea RG, Craigon J, McEvoy TG, Young LE: DNA methylation, insulin resistance, and blood pressure in offspring determined by maternal periconceptional B vitamin and methionine status. Proc Natl Acad Sci USA 2007;104:19351-19356.
- 140 Jones PA, Liang G: Rethinking how DNA methylation patterns are maintained. Nat Rev Genet 2009;10:805-811.
- 141 Burdge GC, Slater-Jefferies J, Torrens C, Phillips ES, Hanson MA, Lillycrop KA: Dietary protein restriction of pregnant rats in the F0 generation induces altered methylation of hepatic gene promoters in the adult male offspring in the F1 and F2 generations. Br J Nutr 2007;97:435-439.
- 142 Benyshek DC, Johnston CS, Martin JF: Glucose metabolism is altered in the adequately-nourished grand-offspring (F3 generation) of rats malnourished during gestation and perinatal life. Diabetologia 2006;49:1117-1119.



Cell Physiol Biochem 2016;39:919-938 DOI: 10.1159/000447801 © 2016 The Author(s). Published by S. Karger AG, Basel and Biochemistry Published online: August 12, 2016 www.karger.com/cpb

- 143 Chen Y-P, Xiao X-M, Li J, Reichetzeder C, Wang Z-N, Hocher B: Paternal body mass index (BMI) is associated with offspring intrauterine growth in a gender dependent manner. PLoS ONE 2012;7:e36329.
- 144 Li J, Tsuprykov O, Yang X, Hocher B: Paternal programming of offspring cardiometabolic diseases in later life. J Hypertens DOI:10.1097/HJH.000000000001051.
- 145 Anderson LM, Riffle L, Wilson R, Travlos GS, Lubomirski MS, Alvord WG: Preconceptional fasting of fathers alters serum glucose in offspring of mice. Nutrition 2006;22:327-331.
- 146 Ng S-F, Lin RCY, Laybutt DR, Barres R, Owens JA, Morris MJ: Chronic high-fat diet in fathers programs β-cell dysfunction in female rat offspring. Nature 2010;467:963-966.
- 147 Bakke JL, Lawrence NL, Robinson S, Bennett J: Observations on the untreated progeny of hypothyroid male rats. Metabolism 1976;25:437-444.
- 148 Gabory A, Attig L, Junien C: Sexual dimorphism in environmental epigenetic programming. Mol Cell Endocrinol 2009;304:8-18.
- 149 Tarrade A, Panchenko P, Junien C, Gabory A: Placental contribution to nutritional programming of health and diseases: epigenetics and sexual dimorphism. J Exp Biol 2015;218:50-58.
- 150 Bermejo-Alvarez P, Rizos D, Lonergan P, Gutierrez-Adan A: Transcriptional sexual dimorphism during preimplantation embryo development and its consequences for developmental competence and adult health and disease. Reproduction 2011;141:563-570.
- 151 Gabory A, Roseboom TJ, Moore T, Moore LG, Junien C: Placental contribution to the origins of sexual dimorphism in health and diseases: sex chromosomes and epigenetics. Biol Sex Differ 2013;4:5.
- 152 Mittwoch U: Blastocysts prepare for the race to be male. Hum Reprod 1993;8:1550–1555.
- 153 Bermejo-Alvarez P, Rizos D, Rath D, Lonergan P, Gutierrez-Adan A: Sex determines the expression level of one third of the actively expressed genes in bovine blastocysts. Proc Natl Acad Sci USA 2010;107:3394-3399.
- 154 Hocher B, Schlemm L, Haumann H, Jian Li null, Rahnenführer J, Guthmann F, Bamberg C, Kalk P, Pfab T, Chen Y-P: Offspring sex determines the impact of the maternal ACE I/D polymorphism on maternal glycaemic control during the last weeks of pregnancy. J Renin Angiotensin Aldosterone Syst 2011;12:254-261.
- 155 Hocher B, Chen Y-P, Schlemm L, Burdack A, Li J, Halle H, Pfab T, Kalk P, Lang F, Godes M: Fetal sex determines the impact of maternal PROGINS progesterone receptor polymorphism on maternal physiology during pregnancy. Pharmacogenet Genomics 2009;19:710-718.
- 156 Hocher B, Schlemm L, Haumann H, Poralla C, Chen Y-P, Li J, Guthmann F, Bamberg C, Kalache KD, Pfab T: Interaction of maternal peroxisome proliferator-activated receptor gamma2 Pro12Ala polymorphism with fetal sex affects maternal glycemic control during pregnancy. Pharmacogenet Genomics 2010;20:139–142.
- 157 Clifton VL: Review: Sex and the human placenta: mediating differential strategies of fetal growth and survival. Placenta 2010;31:S33-39.
- 158 Ishikawa H, Rattigan A, Fundele R, Burgoyne PS: Effects of sex chromosome dosage on placental size in mice. Biol Reprod 2003;69:483-488.
- 159 Gabory A, Ferry L, Fajardy I, Jouneau L, Gothié J-D, Vigé A, Fleur C, Mayeur S, Gallou-Kabani C, Gross M-S, Attig L, Vambergue A, Lesage J, Reusens B, Vieau D, Remacle C, Jais J-P, Junien C: Maternal diets trigger sex-specific divergent trajectories of gene expression and epigenetic systems in mouse placenta. PLoS ONE 2012:7:e47986.
- 160 Maric C: Mechanisms of fetal programming of adult hypertension: role of sex hormones. Hypertension 2007:50:605-606.
- 161 Ojeda NB, Grigore D, Yanes LL, Iliescu R, Robertson EB, Zhang H, Alexander BT: Testosterone contributes to marked elevations in mean arterial pressure in adult male intrauterine growth restricted offspring. Am J Physiol Regul Integr Comp Physiol 2007;292:R758-763.
- 162 Ojeda NB, Grigore D, Robertson EB, Alexander BT: Estrogen protects against increased blood pressure in postpubertal female growth restricted offspring. Hypertension 2007;50:679-685.
- 163 Tomat AL, Salazar FJ: Mechanisms involved in developmental programming of hypertension and renal diseases. Gender differences. Horm Mol Biol Clin Investig 2014;18:63-77.
- 164 Hocher B, Slowinski T, Bauer C, Halle H: The advanced fetal programming hypothesis. Nephrol Dial Transplant 2001;16:1298-1299.
- 165 Hocher B: More than genes: the advanced fetal programming hypothesis. J Reprod Immunol 2014;104– 105:8-11.



Cellular Physiology and Biochemistry Published online: August 12, 2016

 Cell Physiol Biochem 2016;39:919-938

 DOI: 10.1159/000447801
 © 2016 The Author(s). Published by S. Karger AG, Basel

 Published online: August 12, 2016
 www.karger.com/cpb

- 166 Parkhurst SM, Ish-Horowicz D: wimp, a dominant maternal-effect mutation, reduces transcription of a specific subset of segmentation genes in Drosophila. Genes Dev 1991;5:341–357.
- 167 Hocher B, Slowinski T, Stolze T, Pleschka A, Neumayer HH, Halle H: Association of maternal G protein beta3 subunit 825T allele with low birthweight. Lancet 2000;355:1241–1242.
- 168 Masuda K, Osada H, Iitsuka Y, Seki K, Sekiya S: Positive association of maternal G protein beta3 subunit 825T allele with reduced head circumference at birth. Pediatr Res 2002;52:687–691.
- 169 Wang X, Zuckerman B, Pearson C, Kaufman G, Chen C, Wang G, Niu T, Wise PH, Bauchner H, Xu X: Maternal cigarette smoking, metabolic gene polymorphism, and infant birth weight. JAMA 2002;287:195–202.
- 170 Yadav U, Kumar P, Yadav SK, Mishra OP, Rai V: Polymorphisms in folate metabolism genes as maternal risk factor for neural tube defects: an updated meta-analysis. Metab Brain Dis 2015;30:7–24.
- 171 Liu J, Zhang Y, Jin L, Li G, Wang L, Bao Y, Fu Y, Li Z, Zhang L, Ye R, Ren A: Variants in maternal COMT and MTHFR genes and risk of neural tube defects in offspring. Metab Brain Dis 2015;30:507–513.
- 172 van Beynum IM, Kapusta L, den Heijer M, Vermeulen SHHM, Kouwenberg M, Daniëls O, Blom HJ: Maternal MTHFR 677C>T is a risk factor for congenital heart defects: effect modification by periconceptional folate supplementation. Eur Heart J 2006;27:981–987.
- 173 Fu Y, Wang L, Yi D, Jin L, Liu J, Zhang Y, Ren A: Association between maternal single nucleotide polymorphisms in genes regulating glucose metabolism and risk for neural tube defects in offspring. Birth Defects Res Part A Clin Mol Teratol 2015;103:471–478.
- 174 Cohen IL, Liu X, Lewis MES, Chudley A, Forster-Gibson C, Gonzalez M, Jenkins EC, Brown WT, Holden JJA: Autism severity is associated with child and maternal MAOA genotypes. Clin Genet 2011;79:355–362.
- 175 Torres-Espínola FJ, Altmäe S, Segura MT, Jerez A, Anjos T, Chisaguano M, Carmen López-Sabater M, Entrala C, Alvarez JC, Agil A, Florido J, Catena A, Pérez-García M, Campoy C: Maternal PPARG Pro12Ala polymorphism is associated with infant's neurodevelopmental outcomes at 18 months of age. Early Hum Dev 2015;91:457–462.
- 176 Miodovnik A, Diplas AI, Chen J, Zhu C, Engel SM, Wolff MS: Polymorphisms in the maternal sex steroid pathway are associated with behavior problems in male offspring. Psychiatr Genet 2012;22:115–122.
- 177 Tsai H-J, Liu X, Mestan K, Yu Y, Zhang S, Fang Y, Pearson C, Ortiz K, Zuckerman B, Bauchner H, Cerda S, Stubblefield PG, Xu X, Wang X: Maternal cigarette smoking, metabolic gene polymorphisms, and preterm delivery: new insights on GxE interactions and pathogenic pathways. Hum Genet 2008;123:359–369.
- 178 Danileviciute A, Grazuleviciene R, Paulauskas A, Nadisauskiene R, Nieuwenhuijsen MJ: Low level maternal smoking and infant birthweight reduction: genetic contributions of GSTT1 and GSTM1 polymorphisms. BMC Pregnancy Childbirth 2012;12:161.
- 179 Wu T, Hu Y, Chen C, Yang F, Li Z, Fang Z, Wang L, Chen D: Passive smoking, metabolic gene polymorphisms, and infant birth weight in a prospective cohort study of Chinese women. Am J Epidemiol 2007;166:313– 322.
- 180 Cassina M, Salviati L, Di Gianantonio E, Clementi M: Genetic susceptibility to teratogens: state of the art. Reprod Toxicol 2012;34:186–191.
- 181 Kusinski LC, Stanley JL, Dilworth MR, Hirt CJ, Andersson IJ, Renshall LJ, Baker BC, Baker PN, Sibley CP, Wareing M, Glazier JD: eNOS knockout mouse as a model of fetal growth restriction with an impaired uterine artery function and placental transport phenotype. Am J Physiol Regul Integr Comp Physiol 2012;303:R86-93.
- 182 Kulandavelu S, Whiteley KJ, Qu D, Mu J, Bainbridge SA, Adamson SL: Endothelial nitric oxide synthase deficiency reduces uterine blood flow, spiral artery elongation, and placental oxygenation in pregnant mice. Hypertension 2012;60:231–238.
- 183 Hocher B, Haumann H, Rahnenführer J, Reichetzeder C, Kalk P, Pfab T, Tsuprykov O, Winter S, Hofmann U, Li J, Püschel GP, Lang F, Schuppan D, Schwab M, Schaeffeler E: Maternal eNOS deficiency determines a fatty liver phenotype of the offspring in a sex dependent manner. Epigenetics 2016;11:539–552.
- 184 Costantine MM, Ghulmiyyah LM, Tamayo E, Hankins GDV, Saade GR, Longo M: Transgenerational effect of fetal programming on vascular phenotype and reactivity in endothelial nitric oxide synthase knockout mouse model. Am J Obstet Gynecol 2008;199:250.e1-7.
- 185 Burdge GC, Lillycrop KA: Nutrition, epigenetics, and developmental plasticity: implications for understanding human disease. Annu Rev Nutr 2010;30:315–339.
- 186 Hitchler MJ, Domann FE: An epigenetic perspective on the free radical theory of development. Free Radic Biol Med 2007;43:1023–1036.



Cell Physiol Biochem 2016;39:919-938 DOI: 10.1159/000447801 © 2016 The Author(s). Published by S. Karger AG, Basel and Biochemistry Published online: August 12, 2016 www.karger.com/cpb

Reichetzeder et al.: Developmental Origins of Disease

- 187 Sohal RS, Allen RG, Nations C: Oxygen free radicals play a role in cellular differentiation: an hypothesis. J Free Radic Biol Med 1986;2:175-181.
- 188 Cambonie G, Comte B, Yzydorczyk C, Ntimbane T, Germain N, Lê NLO, Pladys P, Gauthier C, Lahaie I, Abran D, Lavoie J-C, Nuyt AM: Antenatal antioxidant prevents adult hypertension, vascular dysfunction, and microvascular rarefaction associated with in utero exposure to a low-protein diet. Am J Physiol Regul Integr Comp Physiol 2007;292:R1236-1245.
- 189 Chatterjee PK, Cuzzocrea S, Brown PA, Zacharowski K, Stewart KN, Mota-Filipe H, Thiemermann C: Tempol, a membrane-permeable radical scavenger, reduces oxidant stress-mediated renal dysfunction and injury in the rat. Kidney Int 2000;58:658-673.
- 190 Dolinsky VW, Rueda-Clausen CF, Morton JS, Davidge ST, Dyck JRB: Continued Postnatal Administration of Resveratrol Prevents Diet-Induced Metabolic Syndrome in Rat Offspring Born Growth Restricted. Diabetes 2011:60:2274-2284.
- 191 Vieira-Filho LD, Cabral EV, Santos FTJ, Coimbra TM, Paixão ADO: Alpha-tocopherol prevents intrauterine undernutrition-induced oligonephronia in rats. Pediatr Nephrol 2011;26:2019-2029.
- 192 Itani N, Skeffington KL, Beck C, Niu Y, Giussani DA: Melatonin rescues cardiovascular dysfunction during hypoxic development in the chick embryo. J Pineal Res 2016;60:16-26.
- 193 Chmelíková E, Jeseta M, Sedmíková M, Petr J, Tůmová L, Kott T, Lipovová P, Jílek F: Nitric oxide synthase isoforms and the effect of their inhibition on meiotic maturation of porcine oocytes. Zygote 2010;18:235-244.
- 194 Rosselli M, Keller RJ, Dubey RK: Role of nitric oxide in the biology, physiology and pathophysiology of reproduction. Hum Reprod Update 1998;4:3-24.
- 195 Xu X, Du C, Li H, Du J, Yan X, Peng L, Li G, Chen ZJ: Association of VEGF Genetic Polymorphisms with Recurrent Spontaneous Abortion Risk: A Systematic Review and Meta-Analysis. PLoS One 2015;10:e0123696.
- 196 Howell CY, Bestor TH, Ding F, Latham KE, Mertineit C, Trasler JM, Chaillet JR: Genomic Imprinting Disrupted by a Maternal Effect Mutation in the Dnmt1 Gene. Cell 2001;104:829-838.
- 197 Ishitani A, Sageshima N, Hatake K: The involvement of HLA-E and -F in pregnancy. J Reprod Immunol 2006;69:101-113.
- 198 Manzon L, Altarescu G, Tevet A, Schimmel MS, Elstein D, Samueloff A, Grisaru-Granovsky S: Vitamin D receptor polymorphism FokI is associated with spontaneous idiopathic preterm birth in an Israeli population. Eur J Obstet Gynecol Reprod Biol 2014;177:84-88.
- 199 Thulluru HK, Park C, Dufort D, Kleiverda G, Oudejans C, van Dijk M: Maternal Nodal inversely affects NODAL and STOX1 expression in the fetal placenta. Front Genet 2013;4:170.
- 200 Gammie SC, Huang PL, Nelson RJ: Maternal aggression in endothelial nitric oxide synthase-deficient mice. Horm Behav 2000;38:13-20.
- 201 Cieslar SRL, Madsen TG, Purdie NG, Trout DR, Osborne VR, Cant JP: Mammary blood flow and metabolic activity are linked by a feedback mechanism involving nitric oxide synthesis. J Dairy Sci 2014;97:2090-2100.
- 202 Jonas W, Mileva-Seitz V, Girard AW, Bisceglia R, Kennedy JL, Sokolowski M, Meaney MJ, Fleming AS, Steiner M, MAVAN Research Team: Genetic variation in oxytocin rs2740210 and early adversity associated with postpartum depression and breastfeeding duration. Genes Brain Behav 2013;12:681-694.
- 203 An X, Song Y, Hou J, Wang S, Gao K, Cao B: Identification of a functional SNP in the 3'-UTR of caprine MTHFR gene that is associated with milk protein levels. Anim Genet 2016;47:499-503.
- Gu L, Liu H, Gu X, Boots C, Moley KH, Wang Q: Metabolic control of oocyte development: linking maternal 204 nutrition and reproductive outcomes. Cell Mol Life Sci 2015;72:251-271.
- 205 Szyf M: The early life environment and the epigenome. Biochim Biophys Acta 2009;1790:878–885.
- 206 Kuzawa CW, Thayer ZM: Timescales of human adaptation: the role of epigenetic processes. Epigenomics 2011;3:221-234.

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and dynamic processes which require an orchestration of a variety of maternal, paternal and fetal factors for an optimal outcome. This complex interaction between mother, father, placenta and embryo/fetus ensures an optimal supply of nutrients, oxygen and endocrine signals, all fundamental elements for normal development. Disruptions in this supply system may not only have a direct impact on altering fetal growth patterns, but, as evidence suggests, can be associated with the occurrence of diseases in the later life of the offspring. In the current review, we will discuss exogenous (environmental) as well as endogenous factors (both parental and offspring genes) contributing to the complex interaction between mother, father, placenta and embryo/fetus.

Undernutrition

Epidemiological data unequivocally indicate that there is a connection between early life conditions, anthropometric measures at birth and disease susceptibility in later life [1, 2] 59 e "Barker Hypothesis", also called the "Fetal Programming Hypothesis" or the theory of the "Developmental Origins of Health and Diseases (DOHaD)", has become the foundation for this increasingly popular scientific field [2]. Barker et al. were not the first investigating this subject, but it was their groundbreaking epidemiological studies in England and Wales in the late 1980ies that inspired research worldwide. Barker et al. initially demonstrated a geographical relationship between cases of ischemic heart disease in the years 1968-1978 and child mortality rates between 1921-1925 [3]. A follow-up study showed that individuals born with a reduced birth weight had an increased risk for coronary heart (67) ase in their adult life [4, 5]. Hales et al. demonstrated in another follow-up study that there is a similar inverse correlation between birth weight and later life glucose tolerance or insulin resistance [6]. In addition, they revealed that individuals with the lowest birth weight, in comparison to heavier newborns, displayed a six fold increased risk for impaired glucose tolerance or diabetes mellitus type 2 in late adulthood [6]. Until now, these findings have been replicated in several different study populations and in different ethnic groups [7]. Based on their observations, Hales and Barker formulated the "Thrifty Phenotype Hypothesis", a more detailed hypothesis trying to outline a putative mechanism of fetal programming. According to this explanation model, gestational under nutrition induces a series of adaptive processes in the fetus, trying to maximize the chances of survival in the given nutrient-poor environment. However, if a mismatch between pre- and postnatal nutrient supply exists fetal adaptation can be deleterious, increasing the risk for diseases later in life [7, 8].

The Thrifty Phenotype Hypothesis

Research stimulated by the thrifty phenotype hypothesis has improved the understanding of the plasticity of early human development, emphasizing an important role of developmental plasticity as a possible contributing factor to later human disease [9]. Until now, the thrifty phenotype hypothesis was confirmed by a number of human studies. The link between a poor intrauterine environment, restricted fetal growth and increased adult disease risk was well demonstrated in the "Dutch famine study". In this retrospective study, children born during a war-inflicted famine between December 1944 and April 1945 were analyzed [10, 11]. During the famine, the daily caloric intake for the general population was restricted to 400 - 800 kcal per day, about half as much as before and after this period. The comparison of individuals that were in utero during this time with individuals born a year before or after the famine showed that gestational caloric restriction was associated with a decreased birth weight and an increased prevalence of impaired glucose tolerance at an age of 50 years [10]. Moreover, twin studies were able to confirm the Thrifty Phenotype Hypothesis. A Danish study examined mono- and dizygotic twins which were discordant for the occurrence of type 2 diabetes. Results of the study revealed that the diabetic KARGER

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twin was born with a significantly lower birth weight compared to the euglycemic twin sibling [12]. Other twin studies, especially studies on monozygotic twins, highlighted the importance of the intrauterine environment and developmental plasticity, regardless of the underlying genotype [13]. In addition to the connection between a sub-optimal intrauterine environment, disturbed fetal growth patterns and disease predisposition in adulthood, several more recent studies showed that postnatal nutrition is to be regarded as another critical component of the thrifty phenotype hypothesis. Crowther et al. investigated the impact of pos 44 tal weight gain in a cohort of 7-year-old South African children [14]. Results of this study showed that children born with a reduced birth weight and rapid postnatal weight gain, displayed an impaired glucose tolerance already at an age of 7 years [14]. Further studies in Finland and India replicated these findings [15-17]. Various studies have demonstrated that rapid postnatal weight gain in newborns with an initial low birth weight is mainly due to fat accumulation and not due to an increase in muscle mass [18-20]. This specific phenotype was observed in several cohorts of small for gestational age newborns [21, 22]. In addition it was demonstrated that fat accumulation is more prominent in visceral than in subcutaneous fat depots [23, 24].

Parallel to the epidemiological studies outlined above, various animal studies have been conducted over the years in order to investigate the underlying mechanisms of developmental programming more thoroughly. Preclinical results substantiated findings from observational studies and gave more insight into involved mechanisms, also substantiating the thrifty phenotype hypothesis [25, 26]. It was demonstrated that restricting the maternal diet during gestation does not just result in low birth weight but induces disproportional growth. At the expense of organs such as liver, kidney, pancreas, lung and skeletal muscle, the development of brain, heart and adrenal gland is prioritized [25, 27]. Caloric restriction during gestation was shown to reduce pancreatic beta cell mass formation in the offspring, leading to a decreased production of insulin [28]. In a postnatal calorie-rich life, this lack of insulin production can predispose for the development of diabetes [29].

Overnutrition

As worldwide obesity rates are constantly rising, the focus of research has moved from maternal undernuder tion as a predisposing factor for reduced birth weight and adult disease susceptibility, to the application of maternal overnutrition on offspring health. Interestingly, it was shown that maternal overnutrition and an increased birth weight of the newborn elicits similal ffects on offspring health as observed in low birth weight offspring [30, 31]. Being born small for gestational age (SGA) usually is associated with deficits in placental function, placental blood flow and adverse environmental influences, such as maternal undernutrition, particularly if the diet lacks sufficient protein levels [32-35]. Furthermore, literature suggests a complex genetic association, as SGA offspring more commonly occurs in women themselves born SGA [32, 36]. Increases in birth weight are typically associated with maternal obesity 51 gestational or pre-gestational diabetes [37–39]. Moreover, a very recent study provided genetic evidence for a causal relationship between maternal obesityrelated traits and offspring birth weight [37]. Large for gestational age (LGA) offspring usually displays an increased body fat mass and an increased risk for metabolic disease in later life [37, 40, 41]. Current evidence suggests that either being born with a reduced or an increased birth weight increases disease risk in later life. Meta-analysis have underlined this by demonstrating an U-shaped relationship between later life metabolic diseases and birth weight [42-44]. The overlap of the adult phenotype in SGA and LGA offspring raises the important question which mechanisms are affected in these conditions, and vice versa how these mechanisms can be triggered by conditions producing extremely disparate early life phenotypes [32]. Interestingly, specific overnutrition by feeding an isocaloric highprotein diet to rats, was shown to elicit no effects on birth weight but cause an impaired phenotype in adult animals [45]. Furthermore, it was demonstrated in animal and human KARGER

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models that the simultaneous presence of two gestational insults, maternal obesity and maternal stress, are associated with increased rates of both, SGA and LGA offspring [31, 32, 46]. Literature on this subject suggests that the variability in the observed outcomes is likely due to different dosage, timing and downstream effects of a maternal insult. Maternal obesity and pre-pregnancy high-fat intake was on one hand shown to increase the risk for SGA and preeclampsia. On the other hand, this combination of maternal insults was associated with maternal gestational overeating which is associated with gestational diabetes and LGA [30, 32, 47]. Animal stress models revealed that the timing of the maternal stressor during gestation is a key factor for the offspring to be born SGA or LGA [48]. Furthermore, placental development and function, especially an impairment of the place al barrier which normally limits fetal exposure to maternal stress hormones, is thought to play a central role in the severity of maternal stress effects [49, 50]. Taken together, both over and undernutrition are associated with developmental programming. Future studies will give a more precise picture of the complex interaction between nutrition and developmental programming. More recent studies already aimed to discern the role of micronutrient deficiency or excess in regards to developmental origins of disease [51–55]. Furthermore there are also novel approaches to integrate the role of the microbiome and its interaction with nutrition into developmental programming studies [56].

Critical Periods for Nutritional Programming

Experimental and epidemiological data show that effects of developmental programming can be triggered throughout gestation. However, the nature of adult disease can vary according to the timing of a gestational insult [57]. Analysis of the "Dutch Hunger Winter" cohort demonstrated that offspring exposed to famine during early periods of gestation displayed an increased risk for coronary heart disease in later life, which could not be observed if famine exposure happened during later stages of pregnancy [58]. Interestingly, caloric restriction during late gestation was associand with disturbances in glucose-insulin homeostasis, clinically reflected by an increased risk of type 2 diabetes [58]. This finding could be replicated in animal models [59, 60]. Gardner et al. showed that maternal undernutrition in sheep in late gestation also led to impaired glucose-insulin homeostasis, highlighting the importance of late gestational periods in regards to programming effects on intermediary metabolism [60]. According to developmental processes in respective gestational periods, current literature suggests that nutritional insults during early gestation may influence organ development, altering fetal physiology in late gestation, and postnatal function, yet often without a measurable effect on birth weight [59, 61-64]. This was demonstrated by various studies investigating nutritional alteration in the periconceptual period, which is characterized by fertilisation, blastocystogenesis and the implantation process [61-65]. Nutritional insults set in later embryonic and early fetal life, a period which comprises intense organogenesis and placental development, display similar patterns of developmental programming [66-70]. Dietary modifications in later stages of gestation, characterized by very pronounced fetal growth and placental maturation, were shown to have a strong impact on birth weight and organ maturation with pronounced programming effects on intermediary metabolism and hormonal systems [71–75]. Furthermore, also postnatal nutrition plays an important role in developmental programming. A large body of studies have demonstrated that postnatal catch-up growth in low birth weight newborns is a crucial factor for increasing the risk of adult metabolic disturbances [20, 22, 76]. Intriguingly, also in postnatal developmental programming timing is of importance. It was shown that catch-up growth restricted to the first postnatal year did not have an effect on insulin levels, but sustained catch-up growth was associated with higher insulin levels in seven year old- and insulin resistance in eight year old children [77-79]. Next to postnatal periods, newer evidence highlights the importance of preconceptual nutrition of both the mother and the father on offspring health.

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There is a growing amount of evidence, demonstrating that dietary challenges during oozyte development or spermatogenesis can induce permanent phenotypic changes in the offspring [80-84]. Taken together, available evidence highlights the importance of different time frames before, during and after gestation in regards to developmental origins of health and disease. However, given species differences in physiology, metabolism, placental structure and function, cautious interpretation of the available studies is warranted, especially when extrapolating to human situations [72].

Prematurity

More recent data showed that fetal adaptation causing persistent functional and structural changes of the fetal organism can be induced by factors that go beyond gestational nutrition and do not necessarily have to impact on anthropometric measures at birth [85]. A very important factor in this regard is preterm birth. As outlined before, initial studies on fetal programming focused on a 'deprived' intrauterine environment as a cause for low birth weight or SGA [10, 86]. Such anthropometric measurements were then used as surrogate parameters for association analyses with later life disease [10, 86]. However, many of these epidemiologic studies based their investigations on old birth records, sometimes assessing birth weight without considering gestational age at birth [87, 88]. Thus, it is possible that a considerable amount of individuals included in these epidemiologic studies, were preterm individuals and not small for gestational age [87]. De 65g et al. demonstrated in a systematic literature review followed by a meta-analysis, that preterm birth is associated with higher blood pressure in adulthood, suggesting a relevant role of gestation 64 ge in fetal programming [89]. There is an increasing body of evidence that prematurity is associated with an increased risk for various disease in adult life [53, 87, 89–91]. It is known that preterm birth causes an interruption of normal organogenesis, especially in organ systems that display a branching morphogenesis like kidney, lung, pancreas, and the vascular system [87, 92-94]. The developing kidney is particularly vulnerable to preterm birth which causes considerable deficits in organ structure and function [93]. Prematurity was shov 31 to be associated with a lower nephron endowment [95, 96] potentially increasing the risk of hypertension, proteinuria and kidney disease in later life [97, 98]. Although underlying mechanisms of increasing adult disease susceptibility by prematurity on first glance seem to be more direct, there are similarities between being born SGA and preterm. Preterm birth cannot be simply seen as an abrupt termination of gestation, but rather as a pathologic, stressful and inflammatory event, influenced by numerous factors, ranging from ethnicity and socioeconomic status to dysfunctions in hormonal systems and gestational micronutrient deficiencies [53, 88, 99-101]. Similar to SGA infants preterm infants suffer from an adverse intrauterine environment and, by being born prematurely, are additionally exposed to an adverse neonatal environment [102]. Both, SGA and preterm born infants display similar postnatal growth patterns with about 80% of both groups exhibiting catch up growth [102]. Resembling observations in SGA newborns, prematurity predisposes to childhood adiposity, with data indicating a shift in adipose tissue distribution towards visceral fat depots [102]. Furthermore, similar to observations in SGA cohorts, a more rapid postnatal catch-up growth was shown to be associated with greater reductions in insulin sensitivity [103, 104]. A recent systematic review and meta-analysis showed that there is also an association between preterm birth and insulin sensitivity throughout life. However, the data in this regard are conflicting and observed associations might be affected by the overall heterogeneity of the study designs and analyzed populations [105]. Albeit conflicting findings, prematurity putatively increases the risk for insulin resistance which, at least in part, appears to be regulated by postnatal growth. This highlights the importance of an optimal nutritional strategy for preterm infants which yet remains to be determined [104, 1061.

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The Role of Insulin in Fetal Development and Adult Diseases

The developmental origins theory can be applied to all early life events including low birth weight and/or prematurity [107-109]. Adverse environmental exposures during fetal and neonatal life are thought to trigger compensatory persistent physiological responses. Such adaptations may modify set points of physiological systems involved in sustaining homeostasis. This can become maladaptive if a mismatch between anticipated and actual environment occurs. In this regard, a lot of research was focused on insulin resistance as the main culprit for both, altered anthropometric measurements at birth, and later life disease susceptibility. Divergent to the developmental origins theory first postulated by Barker et al., Hattersley and Tooke proposed in their "fetal insulin hypothesis" that genetically determined insulin resistance results in impaired insulin-mediated growth in the fetus as well as in an insulin resistant phenotype in adult life [110]. It is known that type 2 diabetes has a strong genetic component. Furthermore, insulin acts as key factor in fetal growth. Thus, any genetic variant that impairs insulin secretion and/or insulin sensitivity may re 10e birth weight and concomitantly result in adult life type 2 diabetes. Put differently, the "fetal insulin hypothesis" postulated that the genotype, not low birth weight, increases the risk of adult diabetes [110, 111]. The **33** othesis is supported by genetic evidence showing that single nucleotide polymorphisms associated with an increased risk for type 2 diabetes were associated with low birth weight [112]. Moreover, a study in Caucasian mothers revealed that there is a negative correlation between total glycated hemoglobin in cord blood (fetal) and birth weight [113]. The relationship between cord blood glycemia and birth weight is diametrically opposed to the well described positive correlation between maternal glycemia and birth weight which was also observed in this study [113]. When subjected to similar degrees of maternal glycemia (reflected by maternal total glycated hemoglobin), lighter fetuses appear incapable of lowering their blood glucose concentrations (reflected by the newborn's total glycated hemoglobin), as do heavier fetuses. Meanwhile, the findings of an inverse relationship between cord blood glycemia and birth weight were replicated in an Asian cohort, highlighting their validity [114]. Until now, fetal blood glucose concentrations were regarded as a passive reflection of maternal glycemia. However, the observed inverse correlation between cord blood and birth weight showed that the fetal response to similar maternal glucose levels might not behave as uniform as previously thought [113, 114]. From a hypothetical point of view such findings can be explained by both, genetics and the fetal environment, underlining that future research, integratively applying genetic and epigenetic methodology, is still needed to better characterize the association between early life and adult disease susceptibility.

Epigenetics

Although the underlying molecular mechanisms are incompletely understood so far, there is convincing evidence that developmental plasticity is mediated by epigenetic modifications of the DNA. Important epigenetic mechanisms are histone modifications, non coding RNAs and DNA methylation [115, 116]. These tools generally affect how accessible DNA is to transcription factor complexes, how efficiently transcription proceeds, and how stable already transcribed mRNA is [104]. Histone modifications consist of chemical alterations such as acetylation, phosphorylation and methylation [116] which can modulate chromatin structure, thus influencing the accessibility of the transcription machinery to the gene [117]. Non coding RNAs can trigger RNAse activity by RNA interference eliminating mRNA transcribed by target genes [38, 118]. The current 17 pest studied epigenetic mechanism is DNA methylation [119]. DNA methylation is the addition of a methyl group at the C5 position of the cytosine pyrimidine ring via DNA methyl transferase activity [120]. Methylated cytosines are generally located in cytosine-phosphate-guanine (CpG) sequences [121, 122]. Although about 70% of all genomic CpGs are methylated, there are clusters of KARGER

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CpGs, termed CpG islands, that remain unmethylated [102, 120, 123]. Such unmethylated CpG islands are associated with about 60% of all human genomic promoters [124]. Methylating a CpG site attracts methyl-binding proteins that trigger chromatin remodeling, leading to a more condensed chromatin, thus restricting access for the transcription machine [125]. Therefore, promoter regions of translated genes usually display low methylated CpG islands, whereas un-translated genes are heavily methylated.

In mammalian development, there are two main periods characterized by extensive epigenetic modifications. During the course of gametogenesis, genome-wide demethylation takes place followed by remethylation before fertilization. In early phases of embryogenesis, extensive epigenetic modifications occur, with phases of total demethylation alternating with phases of remethylation, ensuring the totipotency of the developing embryo [126]. Additionally, de- and remethylation processes after fertilization are thought to play a role in the removal of acquired epigenetic modifications, especially those acquired during gametogenesis [127–129]. However, some parental epigenetic modifications seem to escape the second wave of demethylation, underlining a potential inheritance of epigenetic modifications set during gametogenesis [102, 130].

Epigenetic mechanisms are not only important in early phases of pregnancy but throughout gestation [131, 132]. Current literature suggests that epigenetic modifications acquired during early developmental phases can be permanent [133]. It was demonstrated in a variety of experimental models and clinical studies of fetal programming that environmental conditions during gestation or shortly after birth can induce epigenetic alterations, stably changing the degree of 63 pmoter methylation and thereby permanently altering gene expression [54, 133]. Rat offspring of dams fed a low-protein diet during pregnancy exhibit decreases in promoter methylation of the glucuronid receptor and the peroxisome proliferator-activated receptor α (PPAR- α) gene in the liver [134]. Similar epigenetic changes were shown for p53 in the kidney [135], the suprarenal angiotensin II type-1b receptor [136], and for the hypothalamic glucocorticoid receptor [137]. More recent data underlined that an alteration of DNA methylation triggered by maternal undernutrition is not tissue specific but a global phenomenon, associated with widespread changes of gene expression [138]. It is not exactly known yet for how long the time window for stable epigenetic changes is opened, but current evidence suggests that the timeframe spans from conceptional to early postnatal stages [137, 139]. It has also been demonstrated that DNA methylation patterns can be transmitted from one generation to the following [140]. Moreover, it was shown that a gestational low-protein diet fed to F0 dams can still alter promoter methylation and gene expression of the F2 generation without any nutrient restriction in the F1 generation [141]. Another study even described a significant impact of a gestational/lactational low protein diet administered only to the F0 generation on the phenotype of F3 generation offspring [142].

Paternal Programming

Until now, the focus regarding fetal p⁵⁸ ramming was mostly set on maternal programming. However, there is accumulating evidence that the father also plays a relevant role in epigenetic modifications of the offspring's phenotype [143, 144]. Epidemiological data showed that the grandchildren of men, who were exposed to a restricted caloric intake during the slow growth phase just before reaching puberty, lived significantly longer than grandchildren of men, who experienced overnutrition during this phase [82]. In a more detailed analysis of t³³ e data, it was demonstrated that an excess caloric intake of the paternal grandfather was associated with a fourfold increased risk of dying from diabetes associated disease in the grandchildren's generation [83]. There is also data from animal experiments confirming an influence in terms of fetal programming on the offspring. In a study by Anderson et al. it was shown that paternal fasting before mating was associated with reduced serum glucose levels in the F1 generation [145]. Ng et al. were able to demonstrate

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that a preconceptional high fat diet of the father causes beta cell dysfunction of the pancreas in the F1 generation [146]. Apart from dietary influences, Bakke et al. demonstrated in a pioneer study that hypothyroidism of male rats before mating resulted in significant phenotypic changes of the F1 generation [147]. Paternal hypothyroidism was induced either by radiothyroidectomy or by large doses of neonatally injected thyroxine. Offspring of hypothyroid fathers displayed a slower postnatal development, reduced weaning weights and increased final body weights, and had enlarged pituitary and thyroid glands. Furthermore, female offspring from thyroidectomized fathers developed significantly smaller uteri and enlarged ovaries, whereas testes of male offspring were significantly smaller [147].

Sex Differences in Developmental Origins of Disease

The existence of sex specific differences in animal models of developmental programming is well described in currently available literature. The vast majority of non communicable diseases, which in many cases have developmental origins, often display some degree of sex bias [148]. 57st developmental programming studies have shown that the same stimulus can elicit different long term effects, depending on the sex of the offspring. The underlying mechanism of this sexual dimorphism is not well understood [149]. It was demonstrated that gene expression shows sex specific differences, which are already detectable in the preimplanted embryo, long before any gonadal development and sex hormone production [150-152]. Thus, such early phenotypic differences can only be attributed to transcriptional differences resulting from different sex chromosomes, i.e. to Y-chromosomal genes and X-chromosomal genes that to a smaller or bigger extent escape X-chromosome inactivation [150]. Moreover, it was shown that sex chromosomal differences in gene expression can influence the transcription of autosomal genes, resulting in prominent sex specific transcriptional differences [150]. Analysis of bovine blastocysts demonstrated that one third of genes, most of them of autosomal origin, displays sex specific differences in expression [153]. Mechanistically, the imprinting of X-linked genes may be involved in sex specific expression differences. In females specific imprinting ensures that the paternal allele is uniquely or preferentially expressed. As male embryos are missing the paternally inherited X-chromosome, synergistic effects of double X dosage plus imprinting mechanisms may be responsible for sex specific transcriptional differences [150]. Early gestational sexual dimorphism in protein expression may influence several molecular pathways, including glucose and protein metabolism and impact on epigenetic mechanisms, particularly DNA methylation. This might be one underlying reason for a sex specific different susceptibility to environmental stressors, leading to distinct long-term effects in the offspring [150]. Furthermore, there is evidence in literature that the fetal sex as a major genetic variant of the fetal genome may influence maternal physiology during gestation in genetically susceptible pregnant women. It was demonstrated that depending on fetal sex certain maternal genetic variants (ACE I/D; PPARG2 Pro 12 Ala; PROGINS progesteron receptor polymorphism) are associated with different outcomes in regards to maternal glycemia and blood pressure regulation, both very influential factors in fetal development [154–156]. Another crucial factor for sexual dimorphism in developmental programming is the placenta [148, 149, 151]. Being the functional link between the maternal environment and the fetus, the placenta plays a central role as a buffer for environmental effects and is capable of modulating effects of adverse intrauterine conditions [151]. As this organ derives from embryonic trophoblast cells, it bears the same sex as the embryo/fetus [151]. Depending on the sex of the fetus, the placenta displays sexual dimorphism, with different growth rates and a varying responsiveness to fetal hormones [157]. In many species male placentas usually are larger or distinctively shaped, an observation that, at least in mice, seems to be independent of androgen effects [151, 158]. More importantly, current literature suggests that female and male placentas are characterized by different molecular mechanisms to optimize the outcome of the offspring, with distinct transcriptomes, perfectly

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shaped for a proper development of the given sex [151, 159]. Concomitantly, this results in a different susceptibility to perturbations in the intrauterine environment and the ability to cope with them. The molecular mechanisms underlying this sexually distinct adaptive responses are largely unknown, however, current data indicates that the sex specific genome and epigenome are key factors [150, 151]. Finally, regarding disease susceptibility in later life, the impact of sex hormones during development and over the course of life has to be taken into account. However, only a few studies so far have examined the contribution of sex hormones in later life disease susceptibility due to developmental programming. Ojeda et al. demonstrated in a rat model of placental insufficiency that intrauterine growth retardation (IUGR) was associated with hypertension in male offspring [160, 161]. Furthermore, serum testosterone levels were twofold higher in IUGR males than in healthy controls, indicating a connection to the observed hypertension in male IUGR offspring. Castration at an age of 10 weeks abolished hypertension in male IUGR offspring with intrauterine growth retardation. No effects of castration on blood pressure were observed in healthy controls [161]. Female growth retarded offspring also developed hypertension, however this increase in blood pressure returned to normotensive values once the animals reached puberty and displayed increasing levels of estradiol [162]. Ovariectomy at an age of 10 weeks blunted this decrease in blood pressure compared to intact IUGR females. In a third group that received $17\beta\text{-estradiol}$ replacement, ovariectomy induced increases in blood pressure were attenuated [162]. Results from these and similar studies indicate that sex hormones can influence the long term consequences of developmental programming [161-163]. However, until now there is a lack of studies that evaluated this matter in different animal models with other outcomes than hypertension in a similar extensive fashion as Ojeda et al. Taken together, sexual dimorphism is tightly connected to developmental programming. The influence of sex hormones and differences in placental function are important factors in this regard. Moreover, disparities in the sex specific genome and epigenome, leading to a transcriptional sexual dimorphism which is already present in the preimplanting embryo, may play a relevant role in the varying susceptibility to environmental stressors among the sexes.

Interaction of Parental Genes, Parental Environment and Fetal Programming

As outlined before, maternal and paternal environmental factors can influence the phenotype of the offspring by inducing epigenetic adaptive mechanisms. Another factor responsible for developmental programming during intrauterine life might be related to parental genes that impact on the fetal phenotype independent of their presence in the fetal genome [164, 165]. About 25 years ago, Parkhurst et al. described a wimp mutation in Drosophila that resulted in a lethal phenotype, although the mutation was not transmitted to the offspring [166]. Hocher et al. translated this finding to mammalian/human development. They showed that a single nucleotide polymorphism (SNP) in the maternal G p 62 ein beta3subunit gene, which is involved in regulation of blood supply to the uterus, is associated with a substantial reduction in birth weight without actually being transmitted to the offspring (Fig. 1) [167]. Other studies later demonstrated similar independent associations between specific maternal genes and offspring phenotype without any transmission of the particular gene [168–55]. It was shown that maternal mutations of relevant genes involved in folate metabolism are associated with an increased risk for neural tube or congenital heart defects [170–172]. Similar findings were demonstrated for maternal polymorphisms involved in glucose metabolism [173]. Such drastic teratogenic consequences highlight the possible impact maternal genetic deficiencies can have, regardless of any transmission to the offspring. Not as drastic alterations on the offspring phenotype, that better fit to the concept of the developmental origins hypothesis were observed for maternal polymorphisms in the monoamine oxidase A, the peroxisome proliferator activated receptor gamma and cytochrome P enzyme genes controlling sex steroid biosynthesis and metabolism [174-176]. Additionally, it was demonstrated that the maternal genotype plays an important role KARGER

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Fig. 1. The Advanced Fetal Programming Hypothesis. (A) Maternal gene dysfunction may influence ovary function. Physiological ovarian function, oocyte development and maturation, including the establishment of epigenetic patterns could potentially be affected by altered maternal gene function [193-196]. (B) Maternal gene dysfunction can impact on the placenta and the embryonic/ fetal environment by altering decidual function [181, 182, 197-199]. (C) Maternal gene dysfunction may alter weaning behavior and lactation performance and thus influence the early postnatal environment [200-203]. (D) The impact of maternal gene dysfunction on embryonal/fetal/neonatal environmental factors listed in A-C may trigger stable, long lasting epigenetic adaptation in the offspring or result in developmental toxicity [204-206]. Regardless of the underlying mechanism, the phenotype of the offspring may be modified by altered maternal gene function without any transmission of the affected maternal gene.



in modifying adverse intrauterine conditions [180]. Studies showed that polymorphisms of genes encoding for xenobiotic metabolizing enzymes, such as *GSTT1*, and *GSTM1* gene or phase I/phase II enzymes such as *CYP1A1* or *EPHX1* can elicit a modifying effect on birth weight among actively or passively smoking mothers [177–179]. Taken together, there is an increasing amount of evidence indicating that parental genes may influence offspring physiology independent of the inheritance of these genes.

In a very recent study, Hocher et al. aimed to better characterize this biological phenomenon in an animal experiment. Therefore, female heterozygous endothelial nitric oxide synthase (eNOS) knockout mice were mated with male wildtype mice and their wildtype offspring was compared to wildtype offspring from wildtype mice. A heterozygous knock out in the eNOS gene was chosen because of the central role eNOS plays in controlling vascular and placental function, negatively affecting the interuterine environment [181, 182]. The partial lack of the maternal eNOS gene resulted in a reduced birth weight and a steatotic liver phenotype of wildtype offspring. Sex specific differences regarding the phenotype were observed [183]. Using a similar study design, Costantine et al. had previously also demonstrated a sex specific transgenerational effect of a maternal heterozygous eNOS deficiency on the vascular phenotype of wildtype offspring [184]. Results of both studies suggest a non-environmentally mediated mechanism of developmental programming driven by altered parental gene function. Without any transmittance of the specific gene, maternal and putatively also paternal gene dysfunction might influence oocyte or sperm maturation and later embryonic and fetal development by the induction of epigenetic modifications or developmental toxicity, finally resulting in an altered phenotype ("The Advanced Fetal Programming Hypothesis"; see Fig. 1) [165, 183]. Next to the general implications of these findings for the field of developmental programming, they also suggest to reassess murine knock out or transgenic animal models, one of the most important tools currently used in studying gene function. The presumed causality between a genetic manipulation and a resulting phenotype should be reconsidered in regards to potential confounding by

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an induction of epigenetic changes due to parental gene dysfunction that is absent in the offspring [183].

Interventional Approaches

Some studies already investigated if it is possible to ameliorate deleterious effects of fetal programming by interventional approaches. Lillycrop et al. showed in a rat model of fetal programming that a high-protein diet of pregnant dams lead to a hypomethylation of different genes in the offspring. However, if folate was supplemented simultaneous to the low protein diet, the previously observed epigenetic modifications were absent [134]. Until now these or similar results were observed by other studies [138, 185]. It was demonstrated that the DNA methylation machinery relies on ingested methyl group donors and other essential micronutrients [185]. A restriction of these factors may have far-reaching consequences on the phenotype of offspring [139, 185].

An interesting explanation model of epigenetic modifications due to environmental influences provides the "Free Radical Theory of Developement" [186, 187]. This theory is based on the biochemical link between redox buffer systems, such as glutathione, and the methyl group metabolism. It was demonstrated that unfavorable intrauterine conditions, triggered for example by gestational protein restriction, can impact on the capacity of redox buffer systems and expose the organism to increased oxidative stress [186]. As an opposing measure, the production of glutathione can be increased which requires methyl group metabolites. This increased demand for methyl group donors can result in a reduced availability of the essential methyl group donator S-adenosylmethionine (SAM), thus affecting epigenetic mechanisms including DNA and histone methylation [186]. Camboine et al. were able to show in a protein restriction model that the simultaneous feeding of a lipidperoxidation inhibitor during pregnancy and lactation is able to prevent the effects of the low protein diet on blood pressure regulation, vascular function and microvascular rarefaction and counteracts a reduction of glutathione [188]. Similar results were generated employing other antioxidants [189-192]. In summary, nutritional interventions during pregnancy affecting the methyl group metabolism might be able to prevent or alter a developmentally programmed phenotype. Future studies are needed to assess, whether such approaches could be translatable therapeutic options targeting the developmental origins of diseases.

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References

Gillman MW: Prenatal famine and developmental origins of type 2 diabetes. Lancet Diabetes Endocrinol 2015;3:751-752.
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Reichetzeder et al.: Developmental Origins of Disease

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- Inadera H: Developmental origins of obesity and type 2 diabetes: molecular aspects and role of chemicals. 29 Environ Health Prev Med 2013;18:185-197.
- 30 Alfaradhi MZ, Ozanne SE: Developmental programming in response to maternal overnutrition. Front Genet 2011:2:27.
- 31 Heerwagen MJR, Miller MR, Barbour LA, Friedman JE: Maternal obesity and fetal metabolic programming: a fertile epigenetic soil. Am J Physiol Regul Integr Comp Physiol 2010;299:R711-722.
- 32 Grissom NM, Reyes TM: Gestational overgrowth and undergrowth affect neurodevelopment: similarities and differences from behavior to epigenetics. Int J Dev Neurosci 2013;31:406-414.
- 33 Godfrey KM, Inskip HM, Hanson MA: The long-term effects of prenatal development on growth and metabolism. Semin Reprod Med 2011;29:257-265.
- 34 Myatt L: Placental adaptive responses and fetal programming. J Physiol 2006;572:25-30.
- 35 Miao Z, Chen M, Wu H, Ding H, Shi Z: Comparative proteomic profile of the human placenta in normal and fetal growth restriction subjects. Cell Physiol Biochem 2014;34:1701-1710.
- 36 Sydsjö G: Long-term consequences of non-optimal birth characteristics. Am J Reprod Immunol 2011;66:81-87.
- 37 Tyrrell J, Richmond RC, Palmer TM, Feenstra B, Rangarajan J, Metrustry S, Cavadino A, Paternoster L, Armstrong LL, De Silva NMG, Wood AR, Horikoshi M, Geller F, Myhre R, Bradfield JP, Kreiner-Møller E, Huikari V, Painter JN, Hottenga J-J, Allard C, Berry DJ, Bouchard L, Das S, Evans DM, Hakonarson H, Hayes MG, Heikkinen J, Hofman A, Knight B, Lind PA, McCarthy MI, McMahon G, Medland SE, Melbye M, Morris AP, Nodzenski M, Reichetzeder C, Ring SM, Sebert S, Sengpiel V, Sørensen TIA, Willemsen G, de Geus EJC, Martin NG, Spector TD, Power C, Järvelin M-R, Bisgaard H, Grant SFA, Nohr EA, Jaddoe VW, Jacobsson B, Murray JC, Hocher B, Hattersley AT, Scholtens DM, Davey Smith G, Hivert M-F, Felix JF, Hyppönen E, Lowe WL, Frayling TM, Lawlor DA, Freathy RM, Early Growth Genetics (EGG) Consortium: Genetic Evidence for Causal Relationships Between Maternal Obesity-Related Traits and Birth Weight, JAMA 2016;315:1129-1140.
- 38 Shi Z, Zhao C, Long W, Ding H, Shen R: Microarray Expression Profile Analysis of Long Non-Coding RNAs in Umbilical Cord Plasma Reveals their Potential Role in Gestational Diabetes-Induced Macrosomia. Cell Physiol Biochem 2015;36:542-554.
- 39 Li J, Song L, Zhou L, Wu J, Sheng C, Chen H, Liu Y, Gao S, Huang W: A MicroRNA Signature in Gestational Diabetes Mellitus Associated with Risk of Macrosomia. Cell Physiol Biochem 2015;37:243-252.
- Gillman MW, Rifas-Shiman S, Berkey CS, Field AE, Colditz GA: Maternal Gestational Diabetes, Birth Weight, 40 and Adolescent Obesity. Pediatrics 2003;111:e221-e226.
- Boney CM: Metabolic Syndrome in Childhood: Association With Birth Weight, Maternal Obesity, and 41 Gestational Diabetes Mellitus. Pediatrics 2005;115:e290-e296.
- Schellong K, Schulz S, Harder T, Plagemann A: Birth weight and long-term overweight risk: systematic 42 review and a meta-analysis including 643,902 persons from 66 studies and 26 countries globally. PLoS ONE 2012;7:e47776.
- Curhan GC, Willett WC, Rimm EB, Spiegelman D, Ascherio AL, Stampfer MJ: Birth Weight and Adult 43 Hypertension, Diabetes Mellitus, and Obesity in US Men. Circulation 1996;94:3246-3250.
- Curhan GC, Chertow GM, Willett WC, Spiegelman D, Colditz GA, Manson JE, Speizer FE, Stampfer MJ: Birth 44 Weight and Adult Hypertension and Obesity in Women. Circulation 1996;94:1310-1315.
- 45 Thone-Reineke C, Kalk P, Dorn M, Klaus S, Simon K, Pfab T, Godes M, Persson P, Unger T, Hocher B: High-protein nutrition during pregnancy and lactation p 21 ams blood pressure, food efficiency, and body weight of the offspring in a sex-dependent manner. Am J Physiol Regul Integr Comp Physiol 406;291:R1025-R1030.
- Djelantik AA, Kunst AE, van der Wal MF, Smit HA, Vrijkotte TGM: Contribution of overweight and 46 obesity to the occurrence of adverse pregnancy outcomes in a multi-ethnic cohort: population attributive fractions for Amsterdam. BJOG 2012;119:283-290.
- Rajia S, Chen H, Morris MJ: Maternal overnutrition impacts offspring adiposity and brain appetite markers-47 21 Julation by postweaning diet. J Neuroendocrinol 2010;22:905-914.
- Calegare BFA, Fernandes L, Tufik S, D'Almeida V: Biochemical, biometrical and behavioral changes in male 48 offspring of sleep-deprived mice. Psychoneuroendocrinology 2010;35:775-784.

KARGER

	Cell Physiol Biochem 2016;39:919-938 DOI: 10.1159/000447801 © 2016 The Author(s). Pl	ublished by S. Karger AG, Basel	
	and Biochemistry Published online: August 12, 2016 www.karger.com/cpb		932
	Reichetzeder et al.: Developmental Origins of Disease		
1	48		
	Togher KL, Togher KL, O'Keeffe MM, O'Ke <mark>35</mark> MM, Khashan AS, Khashan AS, Gutierre		
	LC, Kenny LC, O'Keeffe GW, O'Keeffe GW: Epigenetic regulation of the placental HSD	11B2 barrier and its	
1242001	24) as a critical regulator of fetal development. Epigenetics 2014; 9:816–822. Li J, Wang Z-N, Chen Y-P, Dong Y-P, Shuai H-L, Xiao X-M, Reichetzeder C, Hocher B: La	te gestational maternal	
	serum cortisol is inversely associated with fetal brain growth. Neurosci Biobehav R		
	Gernand AD, Schulze KJ, Stewart CP, West Jr KP, Christian P: Micronutrient deficienc		
	9 prldwide: health effects and prevention. Nat Rev Endocrinol 2016;12:274–289. 9		
	Chen Y, Wang G, Wang X, Ma Z, Chen Y, Chuai M, von Websky K, Hocher B, Yang X: Efi	1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	
	salt-exposure on the development of retina and lens in 5.5-day chick embryo. Cell P	hysiol Biochem	
2000 d	3 14;34:804–817. Reichetzeder C, Chen H, Föller M, Slowinski T, Li J, Chen Y-P, Lang F, Hocher B: Mater	nal vitamin D	
	deficiency and fetal programminglessons learned from humans and mice. Kidney		
	2014;39:315–329.		
54	Junge KM, Bauer T, Geissler S, Hirche F, Thürmann L, Bauer M, Trump S, Bieg M, Wei	chenhan D, Gu L, Mallm	
	J-P, Ishaque N, Mücke O, Röder S, Herberth G, Diez U, Borte M, Rippe K, Plass C, Herm		
	R, Lehmann I: Increased vitamin D levels at birth and in early infancy increase offsp		
10101	20 lence for involvement of epigenetic mechanisms. J Allergy Clin Immunol Pract 2 Vanhees K, Vonhögen IGC, van Schooten FJ, Godschalk RWL: You are what you eat, a		
	the impact of micronutrients on the epigenetic programming of offspring. Cell Mol I	and a second	
	285.		
56	Ganu RS, Harris RA, Collins K, Aagaard KM: Early Origins of Adult Disease: Approac	nes for Investigating the	
	22 grammable Epigenome in Humans, Nonhuman Primates, and Rodents. ILAR J 20		
	Sinclair KD, Singh R: Modelling the developmental origins of health and disease in t	he early embryo.	
	Theriogenology 2007;67:43–53. Roseboom T, de Rooij S, Painter R: The Dutch famine and its long-term consequence	e for adult health. Farly	
	Hum Dev 2006;82:485–491.	s for addit flearth. Early	
	Gardner DS, Pearce S, Dandrea J, Walker R, Ramsay MM, Stephenson T, Symonds ME	: Peri-implantation	
1	undernutrition programs blunted angiotensin II evoked baroreflex responses in you	ing adult sheep.	
	Hypertension 2004;43:1290-1296.		
	Gardner DS, Tingey K, Van Bon BWM, Ozanne SE, Wilson V, Dandrea J, Keisler DH, St		
	ME: Programming of glucose-insulin metabolism in adult sheep after maternal undo Physiol Regul Integr Comp Physiol 2005; 289:R947-954.	ernutrition. Am J	
	Wynn M, Wynn A: Nutrition around conception and the prevention of low birthweig	ht. Nutr Health	
	23 8;6:37-52.	The second se	
62	Kwong WY, Wild AE, Roberts P, Willis AC, Fleming TP: Maternal undernutrition duri	ng the preimplantation	
	period of rat development causes blastocyst abnormalities and programming of pos	stnatal hypertension.	
	Development 2000;127:4195–4202. Edwar 41 J, McMillen IC: Periconceptional nutrition programs development of the c	ardiovascular sustam	
	in the fetal sheep. Am J Physiol Regul Integr Comp Physiol 2002;283:R669-679.	aruiovascular system	
	McEvoy TG, Robinson JJ, Aitken RP, Findlay PA, Robertson IS: Dietary excesses of uro	a influence the viability	
	and metabolism of preimplantation sheep embryos and may affect fetal growth amo		
	Reprod Sci 1997;47:71–90.		
	Kwong WY, Miller DJ, Ursell E, Wild AE, Wilkins AP, Osmond C, Anthony FW, Fleming		
	expression in the rat embryo-fetal axis is altered in response to periconceptional m. Reproduction 2006;132:265–277.	aternal low protein diet.	
	Keproduction 2006;132:265–277. Vonnahme KA, Hess BW, Hansen TR, McCormick RJ, Rule DC, Moss GE, Murdoch WJ,	Niiland MI. Skinner	
	DC, Nathanielsz PW, Ford SP: Maternal undernutrition from early- to mid-gestation		
3	retardation, cardiac ventricular hypertrophy, and increased liver weight in the fetal	sheep. Biol Reprod	
	2003;69:133-140.		
	Dunford LJ, Sinclair KD, Kwong WY, Sturrock C, Clifford BL, Giles TC, Gardner DS: Ma		
	malnutrition during early pregnancy in sheep impacts the fetal ornithine cycle to re	duce fetal kidney	
	microvascular development. FASEB J 2014;28:4880–4892.		
V/	ARGER		
1/.	NINGEIN		





	Cell Physiol Biochem 2016;39:919–938 DOI: 10.1159/000447801 © 2016 The Author(s). Published by S. Karger AG, Basel Published online: August 12, 2016 www.karger.com/cpb	935
	Reichetzeder et al.: Developmental Origins of Disease	
119 32 J: DNA methylation and hepa	tocellular carcinoma. J Hepatobiliary Pancreat Surg 2006;13:265–273.	
and an	thylation: past, present and future directions. Carcinogenesis	
	CpG islands in vertebrate genomes. J Mol Biol 1987;196:261–282.	
	e analysis of CpG islands in human chromosomes 21 and 22. Proc Natl	
27 d Sci USA 2002;99:3740-374 123 Du X, Han L, Guo A-Y, Zhao Z: Feat	ures of methylation and gene expression in the promoter-associated CpG	
	data. Comp Funct Genomics 2012;2012:598987	
	C, Korf I, Chédin F: R-loop formation is a distinctive characteristic of	
그는 것 이 것 같은	promoters. Mol Cell 2012;45:814–825. /itz-Scherer RA, Hansen JC, Woodcock CL: Multiple modes of interaction	
	ding protein MeCP2 and chromatin. Mol Cell Biol 2007;27:864–877.	
126 Messerschmidt DM, Knowles BB,	Solter D: DNA methylation dynamics during epigenetic reprogramming in	
	1 embryos. Genes Dev 2014;28:812–828.	
127 Monk D: Germline-derived DNA i survival of imprints. Int J Biocher	nethylation and early embryo epigenetic reprogramming: The selected	
	Bergmann K, Fuermann M, Jung M, Reis A, Allen N, Reik W, Walter J:	
	zygote marks DNA for later methylation: a mechanism for maternal	
15 cts in development. Mech Dev		
129 Reik W, Dean W, Walter J: Epigen 2001;293:1089–1093.	etic reprogramming in mammalian development. Science	
ener – Martin Roman (Martin Carlos Carlos Construction)	ics of epigenetic inheritance: modes, molecules, and mechanisms. Q Rev	
Biol 2015;90:381-415.		
	thylation, an epigenetic mechanism connecting folate to healthy	
	g. J Nutr Biochem 2009;20:917–926.	
	Qian Y, Jia R, Sun L: Distinct DNA methylomes of human placentas tional diabetes mellitus. Cell Physiol Biochem 2014;34:1877–1889.	
	nutrition and epigenetic programming: chasing shadows. Curr Opin Clin	
2 tr Metab Care 2010;13:284-2)3.	
	AA, Hanson MA, Burdge GC: Dietary Protein Restriction of Pregnant Rats	
the Offspring. J Nutr 2005;135:13	ntation Prevents Epigenetic Modification of Hepatic Gene Expression in 182–1386	
	Γ, Laksana GS, Hsu JL, Lane RH: Uteroplacental insufficiency increases	
apoptosis and alters p53 gene m	thylation in the full-term IUGR rat kidney. Am J Physiol Regul Integr	
Comp Physiol 2003;285:R962-97		
	urns SP, Clark AJL: Epigenetic Modification of the Renin-Angiotensin of Hypertension. Circ Res 2007;100:520–526.	
	ne FA, D'Alessio AC, Sharma S, Seckl JR, Dymov S, Szyf M, Meaney MJ:	
Epigenetic programming by mate	rnal behavior. Nat Neurosci 2004;7:847–854.	
	E, Clark AJL, Langley-Evans S: Genome-wide methylation and gene	
expression changes in newborn i ONE 2013;8:e82989.	ats following maternal protein restriction and reversal by folic acid. PLoS	
	Gardner DS, Sebastian S, Bispham J, Thurston A, Huntley JF, Rees WD,	
	Evoy TG, Young LE: DNA methylation, insulin resistance, and blood	
	by maternal periconceptional B vitamin and methionine status. Proc Natl	
15 d Sci USA 2007;104:19351-1	9356. v DNA methylation patterns are maintained. Nat Rev Genet 2009;10:805–	
811.	6	
141 Burdge GC, Slater-Jefferies J, Torr	ens C, Phillips ES, Hanson MA, Lillycrop KA: Dietary protein restriction tion induces altered methylation of hepatic gene promoters in the adult	
	enerations. Br J Nutr 2007;97:435–439.	
	JF: Glucose metabolism is altered in the adequately-nourished	
grand-offspring (F3 generation) 2006;49:1117–1119.	f rats malnourished during gestation and perinatal life. Diabetologia	2:51 PM
		178:31
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	Reichetzeder et al.: Developmental Origins of Disease	550
	14	
143	Chen Y-P, Xiao X-M, Li J, Reichetzeder C, Wang Z-N, Hocher B: Paternal body mass index (BMI) is associated	
1.4.4	with offspring intrauterine growth in a gender dependent manner. PLoS ONE 2012;7:e36329. Li J, Tsuprykov O, Yang X, Hocher B: Paternal programming of offspring cardiometabolic diseases in later	
144	life. J Hypertens DOI:10.1097/HJH.000000000001051.	
145	Anderson LM, Riffle L, Wilson R, Travlos GS, Lubomirski MS, Alvord WG: Preconceptional fasting of fathers	
	alters serum glucose in offspring of mice. Nutrition 2006;22:327-331.	
146	Ng S-F, Lin RCY, Laybutt DR, Barres R, Owens JA, Morris MJ: Chronic high-fat diet in fathers programs β-cell	
1.47	dysfunction in female rat offspring. Nature 2010;467:963–966.	
147	Bakke JL, Lawrence NL, Robinson S, Bennett J: Observations on the untreated progeny of hypothyroid male rats. Metabolism 1976;25:437-444.	
148	Gabory A, Attig L, Junien C: Sexual dimorphism in environmental epigenetic programming. Mol Cell	
	Endocrinol 2009;304:8–18.	
149	Tarrade A, Panchenko P, Junien C, Gabory A: Placental contribution to nutritional programming of health	
	and diseases: epigenetics and sexual dimorphism. J Exp Biol 2015;218:50–58.	
150	Bermejo-Alvarez P, Rizos D, Lonergan P, Gutierrez-Adan A: Transcriptional sexual dimorphism during	
	preimplantation embryo development and its consequences for developmental competence and adult health and disease. Reproduction 2011;141:563–570.	
151	· 같은 사업에서 가장 같은 것입니다. 이상 전 전 전 전 전 전 전 전 전 전 전 전 전 전 전 전 전 전	
	dimorphism in health and diseases: sex chromosomes and epigenetics. Biol Sex Differ 2013;4:5.	
152	Mittwoch U: Blastocysts prepare for the race to be male. Hum Reprod 1993;8:1550–1555.	
153	Bermejo-Alvarez P, Rizos D, Rath D, Lonergan P, Gutierrez-Adan A: Sex determines the expression level of	
	one third of the actively expressed genes in bovine blastocysts. Proc Natl Acad Sci USA 2010;107:3394– 3399.	
154		
1.04	T, Chen Y-P: Offspring sex determines the impact of the maternal ACE I/D polymorphism on maternal	
	glycaemic control during the last weeks of pregnancy. J Renin Angiotensin Aldosterone Syst 2011;12:254–	
	261.	
155	Hocher B, Chen Y-P, Schlemm L, Burdack A, Li J, Halle H, Pfab T, Kalk P, Lang F, Godes M: Fetal sex	
	determines the impact of maternal PROGINS progesterone receptor polymorphism on maternal physiology 22 ing pregnancy. Pharmacogenet Genomics 2009;19:710–718.	
156	22 ther B, Schlemm L, Haumann H, Poralla C, Chen Y-P, Li J, Guthmann F, Bamberg C, Kalache KD, Pfab T:	
	Interaction of maternal peroxisome proliferator-activated receptor gamma2 Pro12Ala polymorphism with	
	38 sex affects maternal glycemic control during pregnancy. Pharmacogenet Genomics 2010;20:139–142.	
157		
1.50	survival. Placenta 2010;31:S33-39.	
158	Ishikawa H, Rattigan A, Fundele R, Burgoyne PS: Effects of sex chromosome dosage on placental size in mice. Biol Reprod 2003;69:483–488.	
159	그렇게 한 것 같이 있는 것 같은 것 같이 있는 것 같이 아니는 것을 잘 잘 알았다. 것 것 것 것 것 같은 것 것 것 것 것 것 같은 것 것 것 것 같이 있는 것 것 것 것 것 것 것 것 것 것 것 것 것 것 것 것 것 것	
	Attig L, Vambergue A, Lesage J, Reusens B, Vieau D, Remacle C, Jais J-P, Junien C: Maternal diets trigger	
	sex-specific divergent trajectories of gene expression and epigenetic systems in mouse placenta. PLoS ONE	
	2012;7:e47986.	
160	Maric C: Mechanisms of fetal programming of adult hypertension: role of sex hormones. Hypertension	
161	2007;50:605–606. Ojeda NB, Grigore D, Yanes LL, Iliescu R, Robertson EB, Zhang H, Alexander BT: Testosterone contributes	
101	to marked elevations in mean arterial pressure in adult male intrauterine growth restricted offspring. Am J	
	Physiol Regul Integr Comp Physiol 2007;292:R758-763.	
162	Ojeda NB, Grigore D, Robertson EB, Alexander BT: Estrogen protects against increased blood pressure in	
	postpubertal female growth restricted offspring. Hypertension 2007;50:679–685.	
163	Tomat AL, Salazar FJ: Mechanisms involved in developmental programming of hypertension and renal	
164	diseases. Gender differences. Horm Mol Biol Clin Investig 2014;18:63–77. Hocher B. Slowinski T. Bayer C. Holle H: The advanced fatal programming hypothesis. Nephrel Dial	
104	Hocher B, Slowinski T, Bauer C, Halle H: The advanced fetal programming hypothesis. Nephrol Dial Transplant 2001;16:1298–1299.	
165		
	105:8-11.	
K	ARGER	
TM	TI Val la IX	

Cellular Physiology

Cell Physiol Biochem 2016;39:919-938 DOI: 10.1159/000447801 © 2016 The Author(s). Published by S. Karger AG, Basel

and Biochemistry Published online: August 12, 2016 www.karger.com/cpb

Reichetzeder et al.: Developmental Origins of Disease

166 Parkhurst SM, Ish-Horowicz D: wimp, a dominant maternal-effect mutation, reduces transcription of a specific subset of segmentation genes in Drosophila. Genes Dev 1991;5:341–357.

167 Hocher B, Slowinski T, Stolze T, Pleschka A, Neumayer HH, Halle H: Association of maternal G protein beta3 subunit 825T allele with low birthweight. Lancet 2000;355:1241–1242.

- 168 Masuda K, Osada H, Iitsuka Y, Seki K, Sekiya S: Positive association of maternal G protein beta3 subunit 825T allele with reduced head circumference at birth. Pediatr Res 2002;52:687–691.
- 169 Wang X, Zuckerman B, Pearson C, Kaufman G, Chen C, Wang G, Niu T, Wise PH, Bauchner H, Xu X: Maternal cigarette smoking, metabolic gene polymorphism, and infant birth weight. JAMA 2002;287:195–202.
- 170 Yadav U, Kumar P, Yadav SK, Mishra OP, Rai V: Polymorphisms in folate metabolism genes as maternal risk factor for neural tube defects: an updated meta-analysis. Metab Brain Dis 2015;30:7–24.
- 171 Liu J, Zhang Y, Jin L, Li G, Wang L, Bao Y, Fu Y, Li Z, Zhang L, Ye R, Ren A: Variants in maternal COMT and MTHFR genes and risk of neural tube defects in offspring. Metab Brain Dis 2015;30:507–513.
- 172 van Beynum IM, Kapusta L, den Heijer M, Vermeulen SHHM, Kouwenberg M, Daniëls O, Blom HJ: Maternal MTHFR 677C>T is a risk factor for congenital heart defects: effect modification by periconceptional folate supplementation. Eur Heart J 2006;27:981–987.
- 173 Fu Y, Wang L, Yi D, Jin L, Liu J, Zhang Y, Ren A: Association between maternal single nucleotide polymorphisms in genes regulating glucose metabolism and risk for neural tube defects in offspring. Birth Defects Res Part A Clin Mol Teratol 2015;103:471–478.
- 174 Cohen IL, Liu X, Lewis MES, Chudley A, Forster-Gibson C, Gonzalez M, Jenkins EC, Brown WT, Holden JJA: Autism severity is associated with child and maternal MAOA genotypes. Clin Genet 2011;79:355–362.
- 175 Torres-Espínola FJ, Altmäe S, Segura MT, Jerez A, Anjos T, Chisaguano M, Carmen López-Sabater M, Entrala C, Alvarez JC, Agil A, Florido J, Catena A, Pérez-García M, Campoy C: Maternal PPARG Pro12Ala polymorphism is associated with infant's neurodevelopmental outcomes at 18 months of age. Early Hum Dev 2015;91:457–462.
- 176 Miodovnik A, Diplas AI, Chen J, Zhu C, Engel SM, Wolff MS: Polymorphisms in the maternal sex steroid pathway are associated with behavior problems in male offspring. Psychiatr Genet 2012;22:115–122.
- 177 Tsai H-J, Liu X, Mestan K, Yu Y, Zhang S, Fang Y, Pearson C, Ortiz K, Zuckerman B, Bauchner H, Cerda S, Stubblefield PG, Xu X, Wang X: Maternal cigarette smoking, metabolic gene polymorphisms, and preterm delivery: new insights on GxE interactions and pathogenic pathways. Hum Genet 2008;123:359–369.
- 178 Danileviciute A, Grazuleviciene R, Paulauskas A, Nadisauskiene R, Nieuwenhuijsen MJ: Low level maternal smoking and infant birthweight reduction: genetic contributions of GSTT1 and GSTM1 polymorphisms. BMC Pregnancy Childbirth 2012;12:161.
- 179 Wu T, Hu Y, Chen C, Yang F, Li Z, Fang Z, Wang L, Chen D: Passive smoking, metabolic gene polymorphisms, and infant birth weight in a prospective cohort study of Chinese women. Am J Epidemiol 2007;166:313– 322.
- 180 Cassina M, Salviati L, Di Gianantonio E, Clementi M: Genetic susceptibility to teratogens: state of the art. Reprod Toxicol 2012;34:186–191.
- 181 Kusinski LC, Stanley JL, Dilworth MR, Hirt CJ, Andersson IJ, Renshall LJ, Baker BC, Baker PN, Sibley CP, Wareing M, Glazier JD: eNOS knockout mouse as a model of fetal growth restriction with an impaired uterine artery function and placental transport phenotype. Am J Physiol Regul Integr Comp Physiol 2012;303:R86-93.
- 182 Kulandavelu S, Whiteley KJ, Qu D, Mu J, Bainbridge SA, Adamson SL: Endothelial nitric oxide synthase deficiency reduces uterine blood flow, spiral artery elongation, and placental oxygenation in pregnant mice. Hypertension 2012;60:231–238.
- 183 Hocher B, Haumann H, Rahnenführer J, Reichetzeder C, Kalk P, Pfab T, Tsuprykov O, Winter S, Hofmann U, Li J, Püschel GP, Lang F, Schuppan D, Schwab M, Schaeffeler E: Maternal eNOS deficiency determines a fatty liver phenotype of the offspring in a sex dependent manner. Epigenetics 2016;11:539–552.
- 184 Costantine MM, Ghulmiyyah LM, Tamayo E, Hankins GDV, Saade GR, Longo M: Transgenerational effect of fetal programming on vascular phenotype and reactivity in endothelial nitric oxide synthase knockout mouse model. Am J Obstet Gynecol 2008;199:250.e1-7.
- 185 Burdge GC, Lillycrop KA: Nutrition, epigenetics, and developmental plasticity: implications for understanding human disease. Annu Rev Nutr 2010;30:315–339.
- 186 Hitchler MJ, Domann FE: An epigenetic perspective on the free radical theory of development. Free Radic Biol Med 2007;43:1023–1036.

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