# REVIEW

**Open Access** 

# A comprehensive review on *Moringa oleifera* nanoparticles: importance of polyphenols in nanoparticle synthesis, nanoparticle efficacy and their applications

Haribalan Perumalsamy<sup>1,2,3\*†</sup>, Sri Renukadevi Balusamy<sup>4\*†</sup>, Johan Sukweenadhi<sup>5</sup>, Sagnik Nag<sup>6</sup>, Davoodbasha MubarakAli<sup>7</sup>, Mohamed El-Agamy Farh<sup>8</sup>, Hari Vijay<sup>9</sup> and Shadi Rahimi<sup>10\*</sup>

# Abstract

Moringa oleifera is one of the popular functional foods that has been tremendously exploited for synthesis of a vast majority of metal nanoparticles (NPs). The diverse secondary metabolites present in this plant turn it into a green tool for synthesis of different NPs with various biological activities. In this review, we discussed different types of NPs including silver, gold, titanium oxide, iron oxide, and zinc oxide NPs produced from the extract of different parts of M. oleifera. Different parts of M. oleifera take a role as the reducing, stabilizing, capping agent, and depending on the source of extract, the color of solution changes within NP synthesis. We highlighted the role of polyphenols in the synthesis of NPs among major constituents of *M. oleifera* extract. The different synthesis methods that could lead to the formation of various sizes and shapes of NPs and play crucial role in biomedical application were critically discussed. We further debated the mechanism of interaction of NPs with various sizes and shapes with the cells, and further their clearance from the body. The application of NPs made from M. oleifera extract as anticancer, antimicrobial, wound healing, and water treatment agent were also discussed. Small NPs show better antimicrobial activity, while they can be easily cleared from the body through the kidney. In contrast, large NPs are taken by the mono nuclear phagocyte system (MPS) cells. In case of shape, the NPs with spherical shape penetrate into the bacteria, and show stronger antibacterial activity compared to the NPs with other shapes. Finally, this review aims to correlate the key characteristics of NPs made from M. oleifera extract, such as size and shape, to their interactions with the cells for designing and engineering them for bio-applications and especially for therapeutic purposes.

**Keywords** Moringa nanoparticles, Metal nanoparticles, Green synthesis, Secondary metabolites, Application of Moringa nanoparticles, Critical parameters

<sup>†</sup>Haribalan Perumalsamy and Sri Renukadevi Balusamy have contributed equally to this work.

\*Correspondence: Haribalan Perumalsamy harijai2004@gmail.com Sri Renukadevi Balusamy renubalu@sejong.ac.kr Shadi Rahimi shadir@chalmers.se Full list of author information is available at the end of the article



© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.



# Introduction

Plant-based products have recently attracted a great interest in multiple applications including food production, health products, and bioremediations due to their sustainability, low cost, and less adverse effects on human health [1, 2]. Plant-mediated nanoparticles (NPs) are one of these products which have an increasing demand over the chemically synthetized counterparts due to their minimalistic environmental footprint [3]. *M. oleifera* is one of the plants that has been tremendously exploited for synthesis of a vast majority of nanoparticles including silver, gold, bismuth, iron, ZnO, MgO, NiO, CuO, and the rare nanometal, CeO [4–18]. Details in respect to synthesis of these various nanometals using *M. oleifera* are summarized in Table 1.

*M. oleifera* is a popular species of Genus Moringa that has been characterized by fast growth and harsh condition adaptation [19, 20]. Figure 1 depicts the global distribution of *M. oleifera*. The tree natively exists across a wide range of Asian and African countries including India, Sri Lanka, Pakistan, Bangladesh, Afghanistan, Madagascar, and Arabian Peninsula [21]. The M. oleifera had been used by ancient Egyptians in cosmetics and for skin treatment. Also, it had been acknowledged by Romans and Greeks [22]. In time being, various parts of the plant were broadly used in several environmental, industrial, dietary supplements, and medical applications [23–25]. For instance, seed powder is used in the so-called coagulation-flocculation processes for cleaning drinking water from environmental contaminants such as pesticides, dyes, and pharmaceuticals [26, 27]. Also, different parts of the plant, including seeds, flowers, and leaves are good nutritious source of protein, lipids, vitamins, beta-carotene, minerals, and other nutrients, which are being exploited in food technology [24, 28–31]. The leaves also, besides the nutritious value, contain a

Metal NPs	Plant parts	Synthesis method	Secondary metabolites	Size (nm)	Shape	Zeta potential	Applications	References
Silver NPs (AgN	NPs)							
AgNPs	Leaves	Green	_	57	Spherical		Antimicrobial	[97]
AgNPs	Leaves (freeze dried)	Green	Flavones, terpe- noids and poly- saccharides	11	Spherical	_	Antibacterial	[15]
AgNPs	Leaves (fresh)	Green	Flavones, terpe- noids and poly- saccharides	9	Spherical	_	Antimicrobial	[15]
AgNPs	Leaves	Green	_	5–80	Spherical	-	Optical limiting	[57]
AgNPs	Leaves	Green	_	96.72	Spherical	-	-	[58]
AgNPs	Leaves	Green	Phenolic com- pounds	5–10	Spherical	-	Antioxidant, cytotoxicity	[33]
AgNPs	Leaves	Green	Alkaloids, phytosterol, saponins, terpe- noids	-	-	-	Antibacterial	[58]
AgNPs	Leaves	Green	Terpenoids, flavonoids and polysac- charides	116.2 d.nm	Spherical or Polygonal	_	Antioxidant, Reduction in size of leish- maniasis	[34]
AgNPs	Leaves	Green	-	1–56.9, 2–448.1, 3–4705	-	-	Antimicrobial	[98]
AgNPs	Leaves	Green	-	32	Cubic	-	-	[59]
AgNPs	Leaves	Green	_	20–40	Spherical	_	Antimicrobial	[92]
AgNPs	Leaves	Green	_	10-100	Round	_	Cytotoxicity	[13]
AgNPs	Leaves	Green	_	30	Spherical	_	Antimicrobial	[63]
AgNPs	Leaves	Green	Flavonoid- kaempferol, phenolic- chlorogenic, tannins	11–14.3	Rod	_	Antibacterial	[35]
AgNPs	Leaves	Green	Flavonoids, phenolic com- pounds	50	Spherical and rectangular	_	Cytotoxicity	[36]
AgNPs	Leaves	Green	_	95.12	Spherical	_	-	[160]
AgNPs	Stem bark	Green	Terpenoids, flavonoids and polysac- charides	40	Spherical and Pentagon	_	Anticancer	[37]
AgNPs	Stem bark	Green	-	_	-	-	-	[60]
AgNPs	Seeds	Green	Chlorophyll a, b; carotenoids, phenol, flavo- noids	15.5±4.2	Spherical	_	Immobilization, modulators of heavy metals	[161]
AgNPs	Crude gum	Green	-	60	-	-	Antibacterial	[10]
AgNPs	Crude gum	Green	_	9	Spherical	_	Antimicrobial	[162]
AgNPs	Crude gum	Green	_	50	_	_	Antibacterial	[10]
AgNPs	Flower	Green	-	273.98	Spherical	-	-	[58]
AgNPs	Flower	Green	Flavonoids and phenolic compounds	8	Spherical	_	Antibacterial	[4]
AgNPs	Leaves	Green	-	-	-	-27	Anticancer	[92]
AgNPs/ AuNPs	Leaves	Green	-	11–25	Hexagonal, triangular and spherical	-36.7	Anticancer	[92]

Table 1	Synthesis,	physiochemical	properties,	secondary	metabolites,	and application	ons of moring	a nanoparticles
---------	------------	----------------	-------------	-----------	--------------	-----------------	---------------	-----------------

# Table 1 (continued)

Metal NPs	Plant parts	Synthesis method	Secondary metabolites	Size (nm)	Shape	Zeta potential	Applications	References
Gold nanopart	icles (AuNPs)							
AuNPs	Leaves	Green	-	_	_	-36.9	Anticancer	[92]
AuNPs	Leaves	Green	-	10–20	Spherical	-25.3	Antiproliferative, apoptosis	[65]
AuNPs	Leaves	Green	_	20-60	Spherical	-24.09	Stability	[67]
AuNPs	Pod	Green	Alkaloid, terpe- noids, flavonoid, phenols, tan- nins, saponins quinines, proteins	40–80	Distinct	-	Antibacterial, hepatoprotec- tive	[64]
AuNPs	Gum	Green	-	5–45	Spherical	-	DNA protection	[66]
Zinc oxide nan	oparticles (ZnONP	s)						
ZnONPs	Leaves	Green		52	Spherical	_	Photocatalytic, antibacterial	[83]
ZnONPs	Leaves	Green	Vitamins, flavonoids, and phenolic acids	12.27–30.51	-	-	Electrochemical activity	[42]
ZnONPs	Leaves	Green	-	10.81	Wurtzite	-	-	[85]
ZnONPs	Leaves	Green	-	17.8	Cubic	_	-	[85]
ZnONPs	Leaves	Green	Phenols and Amines	10	Irregular	-	Effective adsorption	[86]
ZnONPs	Leaves	Green	Phenolic com- pounds	25	Sphere	_	Antibacterial	[38]
ZnONPs	Flower	Green	-	13.2	_	-	Effective chelat- ing agent	[84]
ZnONPs	Seed	Green	-	13.2	_	-	Effective chelat- ing agent	[84]
ZnONPs	Crude gum	Green	-	60	-	_	Antibacterial	[10]
ZnONPs	Crude gum	Green	_	60	_	_	Antibacterial	[10]
Iron oxide nan	oparticles (FeONPs	)						
FeONPs	Leaves	Green	Polyphenols	30.5	Non-spherical	-	Antioxidant, antibacterial	[78]
FeONPs	Leaves	Green	-	37.3	Non-spherical	-	Antioxidant, antibacterial, thermal proper- ties	[78]
FeONPs	Leaves	Green	-	82±7	Spherical rod	-	Antibacterial, photocatalytic activity	[75]
FeONPs	Leaves	Green	-	$15.01 \pm 6.03$	Rod	-	Antibacterial	[76]
Titanium oxide	nanoparticles (TiC	D <sub>2</sub> NPs)						
TiO <sub>2</sub> NPs	Leaves	Green	-	12	Tetragonal	-	-	[69]
TiO <sub>2</sub> NPs	Leaves	Green	_	100	Spherical	_	Wound healing	[70]
TiO <sub>2</sub> NPs	Seeds	Green	_	90	Spherical	-	Apoptosis and anti-inflam- matory	[54]

large variety of secondary metabolites that exert antimicrobial, antioxidant, growth, and immune enhancing activities making the plant a potential alternative dietary supplement to livestock and poultry for organic meat and egg production, which has received a great public demand in recent years [32]. These constituents also exert pharmaceutical actions such as DNA protection, anti-cancer, anti-diabetic, anti-inflammatory, anti-ulcer,



Fig. 1 Geographical distribution of *M. oleifera*. *M. oleifera* is native to Asia and Africa, but grown in many other nations, as shown. Source: https://www.cabi.org/isc/datasheet/34868

cardiovascular, anti-allergic, analgesic, wound healing, and fever relieving activities [23]. With all these great benefits, it comes as no surprise why *M. oleifera* is called "The Miracle tree" or "nature gift" [22, 28, 29].

M. oleifera bioactive phytochemical profile includes quinine, saponins, flavonoids, tannin, steroids, glycosides, niazinins, niaziminins and many others [23] (Fig. 2A). This large diversity of secondary metabolites has drawn the attention towards M. oleifera as a green tool for synthesis of different NPs that exhibit biological activities. In some literature, it has been experimentally proven that secondary metabolites of M. oleifera can mediate synthesis of NPs with remarkable activities. For example, flavonoids, phenolics, polysaccharides, and terpenoids of the plant leaves, flowers, stem, peel, and pods are found to be the key players in synthesis of diverse NPs with variable sizes and shapes. The synthesized NPs showed a multiplicity of marveling biological activities including antibacterial, antioxidant, and cytotoxic and anticancer [4, 15, 33-41]. Owing to this, it is believed that the hidden power behind the biomedically active NPs is due to the plant part extracts utilized for the synthesis [4, 5, 10, 13, 42]. Besides biomedical applications, NPs synthesized by M. oleifera have also applications such as water decontamination via exerting antimicrobial and chelating activities [43].

Due to the growth flexibility, cultivation, and medicinal properties of *M. oleifera*, it has been adopted in many countries across the globe for industrial purposes [22]. Now, the global market size of *M. oleifera* products was estimated to be 5000 million USD in 2019. With increasing global demand, global market value is expected to reach 8400 million USD by 2026 [44]. Such exponential market increase emphasizes the necessity of learning all tales related to M. oleifera and its applications. A pool of literature reviews has discussed the utilization of M. oleifera extracts in medicine, industry, and environment sustainability, but few literatures have thoroughly presented the extract-based NPs synthesis, secondary metabolites (SMs) part in the synthesis process, and the biotechnological impact of synthesized NPs. Owing to overgrowing population across the world, reduced farming land, poor bioavailability of M. oleifera finds the way for the instigation of *M. oleifera* mediated synthesis positively for better future. Hence, our review will focus on exploitation of M. oleifera extracts in NPs synthesis, how M. oleifera secondary metabolites could be the driving force for NP synthesis, critical factors involved for NP synthesis, their applications, and the future prospects in this area.

# Green synthesis of metallic nanoparticles using various extracts of *M. oleifera*

In its entirety, the process of biosynthesizing metal nanoparticles with Moringa plant extracts comprises three primary stages: (1) the initiation phase, wherein the reduction of metal ions and the nucleation of reduced metal atoms transpire; (2) the growth phase, during which the aggregation of initially formed nanoparticles



Fig. 2 Metal NPs synthesis using various parts of *M. oleifera* extract. A Mechanism of gold NPs formation and bioactive compounds from *M. oleifera*. The original image was adapted with permission from Ref. [157]. 2023, Shadi Rahimi. B Significant secondary metabolites of *M. oleifera* involved in synthesis of metal NPs were highlighted

results in the spontaneous formation of larger particles (achieved through heterogeneous nucleation and growth, along with subsequent reduction of metal ions), concomitant with an elevation in the thermodynamic stability of the nanoparticles; and (3) the termination phase, dictating the ultimate morphology of the nanoparticles [45-50]. During the termination phase, nanoparticles assume the most energetically favorable configuration, and this step is significantly affected by the moringa plant extract's capacity to stabilize metal nanoparticles. For instance, nanotriangles, characterized by high surface energy, exhibit reduced stability. In the absence of adequate stability support within specific extracts, nanotriangles tend to adopt a more stable morphology, such as a truncated triangle, in an effort to minimize Gibbs free energy [51].

When a metallic salt undergoes dissociation into cations and anions, the cations undergo saturation to create hydroxyl complexes. Following the supersaturation of hydroxyl complexes, the growth of metal crystallites with oxygen species commences, leading to the development of crystalline planes with varying energy levels. The provision of heat assumes a pivotal role in supplying energy to the reaction system. This process persists until the activation of the capping agent derived from moringa plant extracts, ultimately halt the growth of high-energy atomic planes. Consequently, specific types of NPs are formed [52]. Broadly speaking, during the synthesis, the green reducing agents contribute electrons to the metal ions, thereby converting them into nanoparticles. The amine groups present in proteins and other significant secondary metabolites, along with the hydroxyl and carboxyl groups found in moringa polyphenols and amino acids, as well as the hydroxyl groups within polysaccharides and carboxyl groups in organic acids, play a crucial role in chelating metal ions. Through this mechanism, these functional groups on the secondary metabolites from the moringa

phyto-extract catalyze the formation of metallic NPs [51, 53].

Silver (Ag), gold (Au), titanium oxide ( $TiO_2$ ), iron oxide (FeO), and zinc oxide (ZnO) NPs produced from the extract of different parts of *M. oleifera* are shown in Fig. 2B and are described as following.

# Silver nanoparticles (AgNPs)

Compared to physical and chemical synthesis, biological synthesis using M. oleifera is said to be clean, easy, nontoxic, and environmentally friendly, which is pH, temperature and time dependent NP synthesis [54]. Different parts of M. oleifera plant including flower, leaf, seed, and stem were used for silver nanoparticle (AgNPs) synthesis as follows. The spherically monodispersed AgNPs prepared using Moringa flower extract showed average crystalline size of 8 nm with increased inhibition of bacterial growth such as Klebsiella pneumoniae and Staphyllococcus aureus [4]. Because of the reduction of silver ion and the creation of AgNPs, the biofabrication of AgNPs using M. oleifera leaf extract produced a reddish brown color [55]. AgNPs produced from M. oleifera leaf extracts displayed antibacterial activities when exposed to sunlight as the principal source of energy [15].

Biosynthesis of colloidal shaped AgNPs were synthesized by reduction method using *M. oleifera* leaf extract as a reducing agent, and exhibited anti-microbial properties [15]. M. oleifera AgNPs were synthesized using a clean, non-toxic and environmentally friendly approach, which possessed significant biological activity against diarrhea bacterial strains such as E. coli, and S. aureus [56]. AgNPs using fresh leaves of *M. oleifera* as a reducing and stabilizing agent, produced polydisperse nanostructures such as irregular contours, nanorods, triangles etc., when leaves were cut finely and boiled for 5 min in water and filtered [57]. Green synthesis was used to produce AgNPs from *M. oleifera* leaf extract, and phytochemicals such as reducing sugar, flavonoids, and phenolic compounds found in the extract are responsible for the conversion of silver ion to metallic silver, resulting in spherical and rectangular NPs [36]. A multivariate optimization employed in the synthesis of AgNPs using M. oleifera stem bark, leaves and flower water extracts showed spherical shaped nanostructures of 95.12 nm, 96.72 nm, and 273.98 nm, respectively [58]. A facile and dependable green production of AgNPs from moringa leaf aqueous extract yielded spherical nanostructures [13]. A simple, quick, and environmentally friendly synthesis of stable AgNPs utilizing M. oleifera leaf extract was developed. The resulting crystalline AgNPs feature centered-cubic shape and an average size of 32 nm [59]. M. oleifera leaves and seeds were employed for the simple production of AgNPs by reducing AgNO<sub>3</sub> to tannic

acid under vigorous shaking with the stabilizer sodium citrate until a dark gray colloidal solution was produced [33]. The seeds of *M. oleifera* were used for the synthesis of AgNPs, where colorless solution was turned into brown color indicating the formation of NPs [60]. *M. oleifera* stem bark extract was used for stable formation of AgNPs which can be used for the treatment of cancer [37]. AgNPs produced biologically by the reducing action of secondary metabolites found in the stem of *M. oleifera*, exhibited great potential for rapid synthesis of NP with antibacterial activity [61].

There are several simple and eco-friendly methods used for AgNPs synthesis using M. oleifera. In a work, M. oleifera leaf extract was employed to create AgNPs in a simple and environmentally friendly manner. This method is considered green chemistry because it uses natural plant extracts instead of harmful chemicals, making it more environmentally friendly. The process was found to be proficient, meaning that it was effective in producing high-quality AgNPs, which had a uniform size distribution, good stability, and high purity [62]. A non-toxic, eco-friendly, simple, and efficient synthesis of AgNPs were synthesized in cold condition using aqueous extract of *M. oleifera* leaves extract [63]. A biogenic synthesis of polyethylene glycol (PEG) induced AgNPs formation using organic compounds of *M. oleifera* aqueous extract is considered as benign, facile, biocompatible, cost-effective, and eco-friendly method [35]. Using M. oleifera, which contains flavonoid, tannin, and phenol phytochemicals as the capping and reducing agent to reduce the silver ions to AgNPs, an easy one-step green technique for AgNP synthesis was developed [35]. M. oleifera gum-based AgNPs were successfully synthesized through ecofriendly, sustainable, economical and easy to use green synthesis process [10].

In conclusion, different *M. oleifera* parts, such as the flower, leaf, seed, and stem, were employed for the synthesis of AgNPs, and depending on the synthesis technique, AgNPs of various shapes and sizes could be produced (Fig. 3).

# Gold nanoparticles (AuNPs)

The synthesis of gold nanoparticles (AuNPs) using pods of *M. oleifera* uses secondary metabolites for NP synthesis. Compared to the ethanol and chloroform extracts, aqueous extracts of pods were highly suitable for AuNP synthesis because of the presence of saponins, flavonoids, tannins, steroids, and alkaloids [64]. One-pot green synthesis of AuNPs using leaves of *M. oleifera* resulted in red-brown color formation within few seconds of mixing leaf extract with the HAuCl<sub>4</sub> solution. This is due to the excitation of surface plasmon vibrations in AuNPs [65]. A simple, easy to handle,



Fig. 3 A Different *M. oleifera* extract produces AgNPs in various shapes. **B** TEM image of AgNPs biosynthesized with *M. oleifera* leaf extract. The original image was adapted from Ref. [158]

biocompatible AuNPs synthesized using gum extracts of *M. oleifera* resulted in the color change from pale yellow to ruby red color formation due to the action of gum extract as the reducing and stabilizing agent for AuNPs synthesis. The presence of leaf proteins and metabolites in the gum extract of *M. oleifera* is what causes the decrease of gold ions and stabilization of AuNPs [66]. An eco-friendly approach was to synthesize AuNPs by chloroauric acid from *M. oleifera* leaf extract as the reducing agent, thus resulting in the stable and spherical NP production [67]. The creation of pink hue in the stable AuNPs from aqueous 1 M chloroauric acid and *M. oleifera* flower aqueous extract was attributed to the plasmon vibrations in the AuNPs being excited by a redox reaction mechanism. The capping agents and molecules such as aliphatic and trace aromatic compounds present in the flowers of *M. oleifera* are responsible for the AuNPs formation [68].

In summary, AuNPs could be synthesized using pods, leaves, flower, and gum extracts of *M. oleifera* and based on the extract source, the AuNP solutions with different colors are produced (Fig. 4).



**Fig. 4** A AuNPs synthesis using different *M. oleifera* extracts as reducing agents. Different *M. oleifera* extracts as the reducing agents caused various color change in AuNPs. **B** TEM images and size distribution of AuNPs biosynthesized by aqueous flower extracts of *M. oleifera*. The original image was adapted with permission from Ref. [68]. 2023, Shadi Rahimi

# Titanium oxide nanoparticles (TiO<sub>2</sub> NPs)

Useful in a wide range of applications, the biosynthesis of titanium oxide nanoparticles (TiO<sub>2</sub>NPs) using Moringa extract is simple, affordable, time-saving, and environmentally benign [69]. The environmentally friendly green chemistry that produces NPs using leaf extract will improve economic viability and sustainable management. Effective TiO<sub>2</sub>NPs synthesis was achieved either with aqueous extract [70–73] or ethanolic extract [69] of *M. oleifera* under various condition. Most of previous results reported using leaves of Moringa [69–72], but there are also those who used seeds [54] and sticks [73], to make TiO<sub>2</sub> NPs.

Previous research used ethanolic leaf extract of Moringa to react with titanium tetra isopropoxide (TTIP) under continuous stirring at 50 °C for 4 h, resulting in tetragonal TiO<sub>2</sub> NPs with an average size of 12 nm [69]. Sivaranjani [70]and Philominathan et al., [70] mixed an aqueous leaf extract of Moringa with a titanium dioxide solution, leading to the formation of spherical TiO<sub>2</sub>NPs with an average size of 100 nm. Kandeil et al. [54] reported similar characteristics of TiO<sub>2</sub>NPs produced by the high-energy ball mills method using Moringa seed extract. Pushpamalini et al. [71] employed a mixture of Moringa aqueous leaf extract, TTIP, and double distilled water to synthesize spherical TiO<sub>2</sub>NPs with an average size of 6.6 nm [71].

Umekar et al. [73] prepared phytoreduced graphene oxide-titanium dioxide nanocomposites (r-GO/TiO<sub>2</sub> NCs) using synthesized graphene oxide, TiO<sub>2</sub>NPs, and *M. oleifera* stick extract through a precipitation technique. Continuous agitation for 8 h at room temperature was required to achieve a uniform dispersion and anchoring of TiO<sub>2</sub>NPs onto the graphene oxide sheet. Furthermore, Satti et al. [72] reported the collection of irregular-shaped TiO<sub>2</sub>NPs with sizes ranging from 20 to 100 nm after continuous stirring of Moringa plant aqueous extract with a TiO<sub>2</sub> salt solution for 24 h at room temperature.

In general, leaves, seeds, and sticks of *M. oleifera* were used for  $TiO_2$  NP synthesis. TTIP and titanium dioxide solution were used for  $TiO_2NP$  synthesis at various shapes and sizes (Fig. 5).

# Iron oxide nanoparticles (FeONPs)

The primary forms of iron oxide (FeO), one of the functional magnetic NPs, are magnetite (Fe<sub>3</sub>O<sub>4</sub>), hematite ( $\alpha$ -Fe<sub>2</sub>O<sub>3</sub>), and maghemite ( $\gamma$ -Fe<sub>2</sub>O<sub>3</sub>). Iron oxide nanoparticles (FeO-NPs), which have a variety of uses including antimicrobial food coatings, catalysis and cosmetics, have received a great deal of attention due to their special qualities and applications [74]. Several reports stated the successful production of iron or iron oxide NPs using Moringa leaves [43, 75–77], fruit [78], seeds extract [27, 43, 79] and its oils [80].

FeONPs were synthesized using a cost-effective method utilizing *M. oleifera* for biomedical applications. This means that it does not require expensive equipment or harmful chemicals, which can significantly reduce the cost of production. Additionally, *M. oleifera* is an easily accessible and abundant plant in many low and middle-income countries, making it a cost-effective source for the synthesis of these NP [81]. FeONPs synthesized using *M. oleifera* have potential biomedical applications, particularly in targeted drug delivery systems. For more effective and efficient medication delivery, these NPs can be functionalized with







Fig. 5 A Different *M. oleifera* extract produced TiO<sub>2</sub> NPs in various sizes. B Spherical TiO2-NPs with a homogeneous nanometric size distribution of 90 nm (range 40–140 nm). The original image was adapted with permission from Ref. [54]. 2023, Shadi Rahimi

certain ligands or antibodies to target cells or tissues in the body. Additionally, FeONPs possess magnetic properties that can be applied to MRI and hyperthermia therapy for the treatment of cancer [81]. Aqueous extracts of the fruit and leaves of M. oleifera were used to create iron nanoparticles (NPs) in a green manner (MOL). MOF-Fe and MOL-Fe have erratic shapes, as seen in TEM and SEM photos (particle size of 45 nm). By adding 0.5 M of Fe-salt dropwise to the aqueous plant extract in a sonicated reactor, biogenic Fe NPs were created. Iron was present, as shown by the formation of NPs with a black hue. Fe NPs displayed peaks about 400 nm, while the Fe-salt solution displayed a peak around 300 nm. The identical spectra at 300 nm were also seen in the nanosuspension of Fe NPs, confirming the inclusion of Fe NPs in the suspension [78]. Meanwhile, Katata-Seru et al. (2018) reported that Fe NPs based on MOL and aqueous extracts of M. oleifera seeds (MOS) exhibited absorbance at 210 and 240 nm in the UV-Vis spectrum. The size range was determined to be between 2.6 and 6.2 nm, and HRTEM (High-Resolution Transmission Electron Microscopy) revealed that MOS-FeNPs are spherically structured with thick surface layers. This could be because the layer of MOS has thickened. The MOL-FeNPs' spherical form and diameter of 3.4 and 7.4 nm were confirmed by HRTEM analysis [43]. Another report found that adding 20 mL of PMC (Psidium guajava-*M. oleifera* composite) extract to the solution of  $FeCl_3$ at a volume ratio of 8:2 (V/V) for 2 min resulted in a color change from orange to dark brown, confirming the synthesis of PMC-NPs. After another hour of stirring, the homogenous solution was transferred to a hot air oven set at 100 °C for 24 h. The generated NPs have a non-uniform rod-like morphology [75]. In the other hand, the mixture's color changes from green to black, making it easy to see that FeO NPs are forming. FeO NPs produced from the leaf extract *of M. oleifera* showed spherical agglomerations with an average diameter of under 100 nm [77].

Without using organic solvents, iron oxide (maghemite,  $\gamma$ -Fe<sub>2</sub>O<sub>3</sub>) NPs were made using a modified sol-gel approach and functionalized with Mo saline solution. By using the Scherrer equation, the average NP size was determined to be 15 nm. The average size determined by Scherrer's equation is corroborated by the average size seen in TEM micrographs (between 10 and 20 nm). The NPs have cubic shape [27]. Iron oxide nanorods (FeO-NRs) were reported to be produced biochemically from FeCl<sub>3</sub> and *M. oleifera*. The formation of FeO-NRs is caused by the visible color changing. X-ray diffraction spectroscopy was used to observe the crystallinity of FeO-NRs, and the results revealed a pattern consistent with the spinel cubic lattice in the tetrahedral hematite structure. The size of the crystallite was estimated to be between 40 and 90 nm. FeO-NRs were found to have a rod-like shape, with an average particle size of 15.01 ± 6.03 nm [76].

The use of methyl ester of M. oleifera oil over Fe-Co impregnated Alumina support at 650 °C under N<sub>2</sub> environment resulted in an eco-friendly green precursor for the production of multi-walled carbon nanotubes (MW CNTs). The diameters of as-synthesized nanotubes ranged from 19 to 22 nm. The interior diameters of the nanotubes ranged from 14 to 18 nm, whereas the exterior diameters of the produced nanotubes ranged from 42 to 50 nm [80]. In order to remediate surface water with high turbidity, the interaction of functionalized magnetic NPs with proteins from M. oleifera seeds was studied. At room temperature  $(25 \pm 2 \text{ °C})$  in an environment of oxygen, Fe<sub>3</sub>O<sub>4</sub> NPs were made utilizing the co-precipitation method in aqueous solution with ferric salts as the precursors. The functionalized nanoparticles were called MOFe<sub>3</sub>O<sub>4</sub>. For Fe<sub>3</sub>O<sub>4</sub> and MOFe<sub>3</sub>O<sub>4</sub>, the Scherrer equation calculated the average crystallite sizes to be 17 nm and 35 nm, respectively [79]. The soluble proteins and other substances in the MO saline extract may be responsible for this increase in crystallite size following functionalization. According to Cheng et al. [82], ferromagnetic magnetite often has NPs with diameters smaller than 50 nm and a black coloring [82], and this has been observed in this study [79].

In summary, leaves, fruit, seed, and oil of *M. oleifera* were used for FeONPs. UV–Vis spectrum and the color change of solution after synthesis could affirm the formation of Fe NPs.

# Zinc oxide nanoparticles (ZnO)

The precipitation method was reported for synthesizing zinc oxide nanoparticles (ZnO NPs) using M. oleifera leaves extract as the natural precursor. The NPs have hexagonal wurtzite structure of an average grain size of 52 nm [83]. ZnO NPs with particle size ranging from 12.27 and 30.51 nm were effectively synthesized by the extract of *M. oleifera* leaves. It was discovered that annealing in air at 500 °C is essential for the manufacture of pure wurtzite ZnO phase [42]. Similar to this, ZnONPs were effectively produced employing a green method and an efficient chelating reduction/oxidizing agent-leaves from M. oleifera. According to Debby Scherrer's equation for ZnONPs, the average crystallite size was 10.81 nm [84]. In a different investigation, *M. oleifera* leaf extract was used to successfully produce ZnONPs. Cu was successfully doped into the ZnO lattice, as evidenced by all of the techniques used in the characterization [85]. Cu dopes the NPs and increases their effectiveness as an adsorbent for the removal of Congo red dye from aqueous solutions in the manufacture of ZnONPs using M. oleifera leaf extract. Congo red is adsorbed on both types of NPs with a pseudo-second-order kinetics, with the Cu-ZnONPs being shown to be more effective as an adsorbent than the undoped ZnONPs [86]. Another group studied the structural and optical differences between ZnONPs made from leaves and those made from different extracts. While the Tauc approximation yields direct band gap measurements are 3.25, 3.18, and 3.12 eV for ZnONPs synthesized by the leaves, seeds, and flowers, respectively, the calculated average crystallite sizes of ZnONPs are found to be 10.8, 13.9, and 13.2 nm. Regarding average crystallite size, smaller sizes are generally desirable as they can enhance the NPs properties such as surface area, reactivity, and optical behavior. When considering the direct band gap values, a larger band gap signifies a wider energy range where the material does not allow electron conduction. This can be advantageous in applications where the material needs to exhibit higher electrical resistivity or ultraviolet (UV) light absorption. Based on those factors, the ZnONPs from M. oleifera leaves has the best results [84]. A simple and eco-friendly synthesis of ZnONPs using root extract of M. oleifera resulted in the average NPs size of around ~25 nm with 2.5 of the peak width. The morphology of the particles was analyzed by TEM and showed that they were spherical in shape [38]. The synthesized NPs utilized crude of Moringa gum, were found to be about 60 nm and 50 nm for ZnONPs and AgNPs, respectively, which is considered an ideal size range for nanomaterials. The synthesized NPs exhibited absorption band in the range of 220-300 nm, a good Plasmon resonance band (PRB) of Ag and Zn NPs. On the other hand, the purified gum

powder was found to have a much larger size. This difference in size can be attributed to the fact that during the synthesis process, the gum acts as a reducing and capping agent, which helps in controlling the size and morphology of the NPs. In contrast, the purified gum powder does not have such properties, which results in larger particle size [10].

In short, leaves, flowers, seeds, root, and crude gum of Moringa were used for ZnONP synthesis. Interestingly, the leave extract could produce smaller size ZnONP compared to flowers, and seeds extracts.

# **Phytochemicals in metal NPs formation**

The utilization of plant extracts provides advantages over other biosynthetic strategies, including accessibility, secure handling, and broad metabolite sustainability. Terpenoids, flavones, ketones, aldehydes, and amides are often the primary phytochemicals involved in the synthesis of  $TiO_2NPs$  [69]. This is in synergy with reports of a smooth, green and one-pot synthesis method using Moringa leaf extract to synthesize  $TiO_2NPs$  [70–72]. The ethanol extract of M. oleifera seeds showed 11 major peaks in the chromatogram of extract. The extract's primary chemical components include phenolic compounds (aceteugenol and eugenol), anti-inflammatory terpenes and sesquiterpenes (cadinese, humulene, copaene, and ocimene), fatty acids (trans-13-octadecenoic and hexadecanoic acid), and others that are not known to be present in any known quantities. As the reducing agent in the production of biomolecules such as proteins, carbohydrates, lipids, and nucleic acids, FeO-NPs and FeO-NRs' Fourier-transform infrared spectroscopy (FT-IR) spectra study revealed various vibrational bands of these molecules [54].

Many studies have used the FT-IR spectra of Moringa extracts and nanosuspensions to provide evidence for the green synthesis of Fe or FeONPs. The spectra show peaks that correspond to O-H bonds, which are present in polysaccharides, proteins, and polyphenols, as well as water molecules. Nanosuspensions refer to a type of nanoparticle formulation where the drug particles are suspended in a liquid medium, usually water, with the help of a stabilizer. The particle size of the drug in nanosuspensions is usually in the range of 10-1000 nm [87], C=O (carbonyl group), C-O and C-H stretching [43, 75, 76, 78]. Other ending vibration of C-N (of aromatic amines) and C-O-C functional groups were also observed [75]. The observed vibration bands below 600  $\text{cm}^{-1}$  give birth to the stretching mode of the Fe-O bond [88], evidence of the successful immobilization of Moringa metabolites on the surface of the formed NPs [75]. XRD analysis could confirm the presence of hematite ( $\alpha$ -Fe<sub>2</sub>O<sub>3</sub>) and ferric oxyhydroxide

(FeOOH) in the Fe NPs [78]. Following the synthesis, there was a reduction in the extract's total phenolic concentration, which showed that the polyphenols were involved in the procedure [78].

The presence of distinct functional groups from diverse biomolecules in the leaves extract of M. oleifera plant that are responsible for the production and stability of ZnS nanostructures, was detected using FT-IR spectroscopy on the biosynthesized ZnSNPs. The presence of certain biomolecules in the 90 percent ethanolic extracts of M.oleifera leaves was discovered and quantified using the results of a study that used thin-layer chromatography densitometry. The average levels of the biomolecules crypto-chlorogenic acid, astragalin and isoquercetin identified in the study were 0.042%, 0.0634% and 0.0467% dry weight, respectively. When compared to the chemical synthesis approach, the biomolecules found in the leaves extract of M. oleifera plant acted as reducing and stabilizing agents, resulting in a 4 h reaction time. The measured negative zeta potential values of 45-55 mV confirm the biosynthesized NPs' remarkable stability over a 3-month timeframe. This demonstrates the innovative nature of the biosynthetic process for ZnSNP production [89].

For each of the primary family bioactive chemicals, a mechanism of ZnONP synthesis via the chemical interaction of Zinc nitrate precursor with M. oleifera bioactive compounds is proposed: vitamin supplements, flavonoids, and phenolic acids [42]. FT-IR analysis showed significant changes in the functional groups of various polyphenols present in the root extract [38]. The purified gum from the M. oleifera plant contained a variety of flavonoids, proteins, and carbohydrates, including D-galactose, D-xylose, D-glucuronic acid, D-mannose, L-arabinose, and L-rhamnose [10]. One major bioactive compound present in extract solution from different parts of plant was used to propose the mechanism of ZnONPs formation. For the leaves extract, a vitamin (L-ascorbic acid) was chosen, a flavonoid (quercetin) for the flowers, and a fatty acid (oleic acid) for the seeds. The nitrate ion ionizes the bioactive chemicals first via these pathways, and subsequently the resultant cations engage with  $Zn^{2+}$  via electrostatic attractions. A follow-up study is required to look at the possibilities of controlling the shape or growth of NPs. [84].

In short, among the major constituents of *M. oleifera* extract, the participation of polyphenols from *M. oleifera* extract in the synthesis of NPs was proved (Fig. 6).

# Applications of *M. oleifera* NPs

Several applications of *M. oleifera* NPs have been extensively studied (Fig. 7). In this section, we will discuss each application in detail.



Fig. 6 Incorporation of phenolic compounds in the M. oleifera NP synthesis

# Moringa oleifera nanoparticles



Fig. 7 Several mechanisms of function from green synthesized metal NPs utilizing *M. oleifera* extract

# Anticancer

A successful synthesis of AgNPs using *M. oleifera* stem bark extract showed excellent anticancer activity against HeLa cells. The anticancer activity of AgNPs is through producing high amount of reactive oxygen species (ROS), inhibition of cell replication and apoptosis induction. Thus, these AgNPs can be suggested as a potential agent for cancer therapy [90]. The leaves mediated AgNPs enhanced antioxidant capacity, concentration of polyphenolic compounds, reducing power and scavenging activity against free radicals, and increased cytotoxicity to cancer cells due to presence of phenolic compounds [33]. One pot synthesis of AgNPs using ethanolic extract of M. oleifera produced AgNPs with selective cytotoxicity to leukemia cells (CD3+) cells but non-toxicity to normal myeloid cells. The mechanism of function was through arrested cell cycle at G1 and S phases, and induced cell apoptosis by upregulation of caspase 8 and pro-apoptotic protein, BID, genes [13]. M. oleifera leaf extract-directed synthesis of AgNPs showed selective cytotoxicity against acute myeloid leukemic cells and did not significantly affect the viability of healthy CD34+cells. The AuNPs, synthesized using the aqueous flower extract of M. oleifera, were cytotoxic to the lung cancer cells whereas nontoxic to normal healthy cells. AgNPs synthesized using M. oleifera extract were both cytotoxic and genotoxic to breast cancer cells while being safe for healthy human endothelial cells [33]. Similarly, 25 mm sized, spherical shaped, and green-synthesized NPs from M. oleifera leaf aqueous extract was investigated for their anti-cancer properties in human colon cancer cell lines such as SW480 and HTC116. As a result, there was a decreased expression of CTNNB1 and LRP6, and increased LRP5 transcript [91]. A comparative study was conducted on one-step synthesis of silver (Ag), gold (Au) and Ag/Au bimetallic nanoparticles using aqueous leaf extract of M. oleifera and their cytotoxic effect against hepatocellular carcinoma (HepG2) and breast cancer (MDA-MD-231 and MCF-7). Compared to all the metal nanoparticles tested, AuNPs have shown to possess higher cytotoxicity with IC<sub>50</sub> value of 37.22-49.94 µg/ml compared to Ag/Au bimetallic nanoparticles. However, AgNPs did not show cytotoxicity up to 60 µg/ml indicating that no synergistic effect was found when using bimetallic nanoparticles [92]. In summary, using right combination of nanoparticles mixture is crucial to exhibit anti-cancer properties.

# Anti-inflammatory

Previous review described that the green synthesized NPs from *M. oleifera* have also been shown to have antiinflammatory properties. The NPs regulate the NF $\kappa$ B pathway by decreasing the expression of I $\kappa$ B $\alpha$ , p-I $\kappa$ B $\alpha$ , and pC. The reduction of these proteins may lead to the suppression of pro-inflammatory cytokines. Additionally, M. oleifera contains bioactive compounds such as glucosinolates, flavonoids, and vitamins, which have potent antioxidant and anti-inflammatory activities, offering protection against chronic inflammatory diseases [65]. Another research article investigated the potential of M. oleifera leaves extract incorporated with Ag-NPs in treating colon cancer induced by AOM (azoxymethane), which is a carcinogenic compound that is commonly used to induce colon cancer in animal models for research purposes. The extract was able to restore hematological measurements to normal levels in both simultaneous and post-treated groups, suggesting its potential anti-cancer properties. However, the specific mechanism behind these properties was not discussed in detail [93]. A recent study compared green silver oxide nanoparticles (AgONPs) synthesized using M. oleifera stem extract with those of standard pharmacological drug, diclofenac sodium. At a concentration of 80  $\mu$ g mL<sup>-1</sup>, synthesized NPs showed significant anti-inflammatory effects compared to that of standard drug suggesting the use of green methods for the treatment of inflammation related disorders [94]. Further studies need to be conducted to reveal the anti-inflammatory effects of metal nanoparticles synthesized using M. oleifera extracts. This will help medical professionals to wisely choose drugs for treating inflammation without major side effects.

# Antimicrobial (Antibacterial, antifungal, and antiviral)

Several reports showed potential use of NPs from Moringa extract as the antimicrobial substance (Fig. 10). The production of AgNPs utilized *M. oleifera* flower extract contains natural antioxidants capable of producing antibacterial compounds against *K. pneumonia* and *S. aureus* [95]. Antibacterial activity of *M. oleifera* flower extract AgNPs was shown against various pathogens namely *S. aureus*, *B. subtilis*, *M. luteus*, *S. paratyphi*, *P. aeruginosa*, and *K. pneumoniae* [96].

The antibacterial activity of *M. oleifera* leaves AgNPs with zone of inhibition (ZOI) in the range of 6–15 nm was shown for different kinds of bacteria namely *S. aureus, C. tropicalis, C. krusei,* and *K. pneumonia* [97]. The antimicrobial activity is due to the action of *M. oleifera* extracts as the antimicrobial agent that reduces the metabolic activity of different pathogens [96]. The AgNPs derived from the leaf extract of *M. oleifera* showed strong inhibition of both gram-negative and gram-positive bacteria as well as various fungal species irrespective of fresh and freeze-dried leaf sample used. In fact, the variation in membrane stability of the bacteria has no effect on the mode of action of these AgNPs, because gram-positive bacteria have a thick peptidoglycan layer, whereas gram-negative bacteria have a hard outer membrane structure

made of lipids and lipoproteins [15]. The biosynthesis of colloidal nanosilver using *M. oleifera* leaf extract as a reducing agent was found to have strong inhibitory action against several types of bacteria, including *E. coli*, *S. aureus*, *S. typhi*, and *B. subtilis*. This inhibitory action was attributed to the smaller size NPs that were synthesized, which were found to change the local electronic structure on their surface. This change in electronic structure enhanced their chemical reactivity and bactericidal effect, allowing them to interact with bacteria at a molecular level and disrupt their cellular processes, ultimately leading to their inhibition or death. These findings suggest that biosynthesized AgNPs could be used as an effective antimicrobial agent in various applications, including packaging materials [98].

The biosynthesized TiO<sub>2</sub> NPs also showed the ability to improve resistance in the wheat plants against Bipolaris sorokiniana, which is responsible for spot blotch disease [72]. 40 mg/L foliar application of biogenic  $TiO_2$ NPs from Moringa extract induced disease tolerance in wheat plants [72]. The potential use of formulated FeONPs from Moringa extract was shown as an antimicrobial substance. Jegadeesan et al. reported that the biogenic Fe NPs made from Moringa leaves extract possessed antimicrobial activity against S. aureus and B. subtilis [78]. Other study reported that the antibacterial effect of FeONPs from Moringa seeds and leaves extract was superior on gram negative strain compared to different antibiotics (Ampicillin, Gentamycin, Erytomycin and Vancomycin), at 20 µg/mL of concentration [43]. Moreover, combination with other plant extract in addition to Moringa showed superior antibacterial activity against pathogens such as S. aureus, E. coli, Shigella, and S. typhi, at 10  $\mu$ g/mL of concentration [75]. In concurrence with that, FeO-NRs also inhibited the growth of S. aureus, P. aeruginosa, E. coli, Shigella, S. typhi, and P. multocida with a higher activity at low concentration [76]. NPs activity against microorganisms occurs due to their small size which can penetrate the bacterial cell [78]. Mirza et al. [99] reported that when the negatively charged microorganism electrostatically interacts with positively charged FeONPs, they oxidize and die. It is worth noting that, when compared to traditional antibacterial medications, the bacterial strains exhibit robust and effective susceptibility to the manufactured Fe or FeONPs at lower concentrations [43, 75, 76, 78].

The synthesized ZnONPs from Moringa leaves extract were studied for antibacterial activities against bacteria like *Bacillus subtilis* and *E. coli* at a very low concentration [83]. Similar results also reported from Moringa roots extract based ZnONPs that exhibited anti-bacterial activities against Gram-positive and Gram-negative bacteria, *B. subtilis and E. coli* [38]. Moreover, ZnONPs made from Moringa crude gum exhibited excellent antibacterial activity against *E. coli* and *S. aureus*, while good activity was observed against "super bug" methicillin-resistant *S. aureus* (MRSA) [10]. Studies indicate that silver (Ag) and ZnONPs prepared by biosynthetic techniques utilizing Moringa leaf extract have enhanced antibacterial properties compared to the same NPs prepared using other agents [100].

The hydroalcoholic extract of *M. oleifera* leaves was successfully used for the production of Cu NPs of < 50 nm exhibiting antioxidant, anti-bacterial, and anti-fungal activities. Bismuth nanoparticles produced from the hydroalcoholic extract of *M. oleifera* leaves showed antioxidant, antifungal, and antibacterial activities [41]. AuNPs derived from *M. oleifera* extract were less cytotoxic and aided in the regeneration of neural cells in vivo [101]. Similarly, ZnO NPs synthesized using *M. oleifera* leaves prevented neuroendocrine toxicity from rotenone by regulating acetylcholinesterase activity and oxidative stress [102].

# Antioxidants

Phenolic compounds such as flavones, flavonols, phenolics, phenolic acids, tannins, and proanthocyanidins derived from the shikimate, pentose phosphate, and phenylpropanoid pathways are the major secondary metabolites which confer antioxidant characteristics of moringa. The use of *M. oleifera* leaf extract in the synthesis of AgNPs is rapid, eco-friendly, non-toxic, economical, and single step procedure and highly efficient than standard pentavalent antimonial treatment against Leishmania major infection probably by boosting antioxidant activity [34]. Better radical scavenging activity against 2,2-diphenl-1-picrylhydrazyl (DPPH) shown by rGO-TiO2 (phytoreduced graphene oxide-titanium dioxide) compared to GO, rGO and TiO<sub>2</sub> separately. In this research article, M. oleifera stick extract is used as a greener reducing agent in the synthesis of rGO-TiO2 nanocomposites. The neutralization reaction of DPPH's free radical nature causes antioxidant performance, with electron transit between reactants showing rGO- $TiO_2$  as an efficient antioxidant [73]. The antioxidant properties of Moringa were also greatly influenced by environmental parameters such as annual precipitation, minimum temperature, and maximum temperature on soil. Therefore, a recent study was conducted to determine the antioxidant properties of phenolic content in NP M. oleifera [103]. Therefore, studying the antioxidant capabilities of M. oleifera NPs produced using distinct extracts derived from different places is vital.

# Water treatment

Moringa extract has been successfully used for the synthesis of various NPs for theragnostic applications (Fig. 8). Anand et al. used petals of *M. oleifera* for synthesizing stable AuNPs using a green chemistry approach. These NPs could catalytically reduce nitroaniline and nitrophenol in the presence of UV light within 3 min which indicates their possible applications in removing nitro-organic compounds from the wastewater [68].

A facile NPs of FeO used in the preparation of different adsorbents for the adsorption of cations, anions, organic components, among other substances present in contaminated water [104]. When tested on fluoride ion removal, ferric oxide NPs synthesized from Moringa extract show spontaneous reaction at 30 °C. Moreover, even though the adsorption capacity of NPs is reduced after first washing, the reuse of the same NPs is likely, which can lead to a cost reduction. The NPs produced in this study are very promising materials for fluoride ion removal due to their adsorption capacity and minimum contact time to reach equilibrium [77].

The functionalized NPs were used in the coagulation/ flocculation and sedimentation (CFS) of surface raw water under the influence of an external magnetic field. A straightforward approach for the manufacture of magnetite ( $Fe_3O_4$ ) and subsequent functionalization





by Moringa extract (MO-Fe<sub>3</sub>O<sub>4</sub>) demonstrated substantial removal efficiencies of the assessed parameters, including the removal of heavy metals such as lead (Pb), cadmium (Cd), and chromium (Cr) from water [79] or the removal of apparent color, turbidity, and compounds with UV 254 nm absorption from surface waters. Within 30 min, a magnetically activated M. oleifera coagulant can successfully remove 85% of apparent color, 90% of turbidity and 50% of compounds with UV 254 nm absorption in surface waters under the influence of an external magnetic field [27]. Moreover, other report showed removal efficiency of 97.1% for the apparent color, 96.8% for turbidity, and 58.3% for the compounds with absorption at UV 254 nm after 10 min of magnetic sedimentation, proving that the CFS process can be optimized to reduce the sedimentation time from 30 to 10 min while still achieving high removal efficiencies for these contaminants. The reuse tests revealed that the functionalized magnetic NPs may be reused up to two times in a row with no noticeable reduction in efficiency in eliminating the examined parameters. Furthermore, the residual values obtained for the turbidity and apparent color parameters following the method were adequate without the requirement for a subsequent treatment operation, such as filtering [79]. In conclusion, coagulants based on natural ingredients, associated with FeONPs, provide a good

alternative to the use of synthetic compounds in water treatment processes [27].

It was also studied whether green synthesized FeONPs from Moringa extract could be used to remove nitrate ion  $(NO_3^-)$  from surface and ground water [43] (Fig. 8B).  $NO_3^-$  removal by MOS-FeNPs was 85% which was higher than that of *M. oleifera* extract. Lower pH of aqueous solution must be maintained for higher percentage of  $NO_3^-$  removal, regardless the type of FeNPs used [43]. It has also been reported that zinc nitrate hexahydrate can be effectively chelated, oxidized, or reduced through the biosynthesis of ZnONPs using a natural extract from *M. oleifera* leaves [85]. Application of various types of moringa NPs in water treatment were summarized (Table 2).

# Factors affecting the NP efficacy using *M. oleifera* extract

Size

Size of the NPs is the foremost features decide the selection of nanocarriers to be incorporated [105]. NP size affects cellular absorption by influencing the enthalpic and entropic characteristics that determine the adhesiveness of NP on the cell surface [106]. Smaller NPs possess a high likelihood to penetrate the cell membrane in comparison to huge ones. It is also perceived that the NPs with the size of >50 nm cohered to the outer membrane caused a slight intervention. NPs with the

**Table 2** The application of NPs made from *M. oleifera* in water treatment

Particle	Activity	Size (nm)	References
AgNPs from <i>M. oleifera</i> seed	Photocatalytic activity toward organic dyes (methylene blue (> 81%), 4-nitrophenol (> 75%), orange red (> 82%) and > 80% of the Pb removal, antimicrobial activity against Gram positive ( <i>Staphylococcus aureus</i> ) and Gram negative ( <i>Escherichia coli, Pseudomonas aeruginosa</i> , and <i>Salmonella</i> enterica <i>typhimurium</i> )	4	[14]
AgNPs from <i>M. oleifera</i> flower	Antibacterial ( <i>Klebsiella pneumoniae, S. aureus</i> ), sensing copper ions from 1 to 12 mM concentrations	22	[4]
FeO-NPs from a mixture of <i>M. oleifera</i> leaf extract and <i>Psidium guajava</i> leaf extract	Photocatalytic activity toward methylene blue	82	[163]
<i>M. oleifera</i> activated carbon modified with chitosan and $Fe_3O_4$ NPs	Cr adsorption capacity of 130.80 mg/g	NA	[164]
Cellulose nanofibers from M. oleifera	Cd (>12 mg/g) and Pb (>80 mg/g) adsorption	80–160	[165]
Activated carbon from <i>M. oleifera</i> seed modified with FeONPs	94.2% oils and greases adsorption	Average pore size 3.1	[166]
Aluminium oxide NPs M. oleifera gum activated carbon	Photocatalytic 94% nitrate and 95% phosphate removal	16.12	[167]
<i>M. oleifera</i> with Fe <sub>3</sub> O <sub>4</sub> NPs	Elimination of S. aureus	NA	[168]
Ag and CuONPs from <i>M. oleifera</i> leaf and stem	Antimicrobial activity against E. coli, S. typhi, P. aerugi- nosa, Klebsiella variicola	NA	[169]
Activated carbon from <i>M. oleifera</i> seed modified with AgNPs	Methyl blue and methyl orange 90% and 77% photo- degradation	200 to 305	[170]
FeONPs from <i>M. oleifera</i> leaf and stem	65.01 mg g <sup>-1</sup> metformin adsorption capacity	NA	[171]
ZnONPs from <i>M. oleifera</i> leaves extract	Adsorption of congo red dye reached equilibrium in 120 min	10	[86]

size of 25-35 nm that cohered to the surface gave minimal deformity. Smaller NPs < 10 nm caused distortion by bending the membrane inwards stacked with several NPs [107]. Salomoni et al. (2017) evaluated the antibacterial activity of AgNPs against P. aeruginosa and E. coli. They stated that, NPs with the size < 100 nm showed better inhibitory action than that of larger NPs, >1000 nm [108]. CuONPs synthesized from *M. olifera* which had size 12-18 nm showed great antimicrobial activity [109]. AgNPs synthesized from *M. olifera* leaves of size 5-22 nm showed great antibacterial activity against S. aureus than K. pneumoniae [4]. The AgNPs synthesized from M. oleifera leaves which had size of 57 nm showed a great inhibition zone against S. aureus, C. tropicalis, and *K. pneumoniae* [97]. The secondary metabolite concentration in the Moringa extract influences the reduction/oxidation and stabilization of NPs, which ultimately determines the size of the NPs.

In addition, the removal and cellular uptake pathway in vivo can also be affected by the NPs size [110]. A study reported that the smaller the NPs, the higher the renal clearance. NPs of size less than 10 nm can effortlessly pass through the blood vessels and get eliminated by kidneys, whereas the larger NPs get arrested by the MPS cells [107]. Liu et al. [111], led an experiment with The NPs size also performs an important role in their extended circulation, bio-distribution, and elimination and permeability due to nanoscale structure [112]. In particular NPs properties, which include thermal, luminescence, electrical, magnetic, permeability, are response to the size of the particles. Altering the pH levels of the synthesis medium can influence the size of NPs since large NPs are synthesized only when pH is acidic [113].

In summary, smaller NPs show better antimicrobial activity, while they can be easily cleared from the body through the kidney (Fig. 9), while larger NPs can be taken by MPS cells.

# Shape

Shape can directly stimulate cell absorption. Choosing the ideal shape for the NPs would facilitate favourable interaction with serum proteins [114]. The study on AuNPs of four different shapes: spherical, rod-shaped, hollow particles and silica shell showed different uptake concentrations for different shapes. The uptake of spherical particles by human endothelial cells was the highest



**Fig. 9** Schematic representation of the kidney showing size dependent elimination. Smaller the NP size, larger the elimination through urine. Medium NPs partially enter, and larger size NPs stacked onto the EC membrane. *EC* Endothelial cells, *GBE* Glomerular basement membrane, *EP* Endothelial podocytes, *FS* Filtration silts

and that of hollow particles [115]. The AgNPs synthesized from *M. oleifera* leaf extract, which was spherical in shape expressed remarkable antimicrobial action, preventing the growth of *S. epidermidis, K. cloacae*, and *E. coli* [61]. The rod and cube shaped AgNPs, showed significant difference of growth reduction for the two studied fungi. The rod shape NPs had less fungi growth compared to the cubic shape [116]. Sphere shaped MnO NPs showed more antioxidant activity than NPs in cubic shape [117].

Studies tend to show that NPs shape and size play a prominent part in the pathways by which particles enter cells, cycling time, identifying the target, and capacity to overcome a biological barrier. The antibacterial activity of triangle-shaped AgNPs was lower than that of spherical particles. Furthermore, as compared to the bigger triangular-shaped AgNPs, the smaller spherical AgNPs were more successful in penetrating inside the bacteria [118]. AgNPs with spherical, triangular plate and disc shapes were experimented and result showed that a high concentration of spherical AgNPs in the ratio was resulted in a significant antibacterial activity [119].

These shape and size are likely to affect how cells recognize and react to the NPs during transport in blood, primarily in small vessels and malignant cells vessels [120]. The investigation in the uptake and transport of different shaped (spheres, rods and discs) NPs across intestinal cells clearly illustrated that rods have greater cellular uptake, while the disc shaped NPs showed greater transport among others [121]. Spheres and rods exhibit different binding patterns in a shear force.

Shape is a core factor that determines the nanomaterials (NMs) delivering drugs efficiently and accurately. The onset of adhesion and prolonged interaction post adhesion are affected by the particle shape [122]. The probability of maintaining adherent or being washed away after initial adhesion, internalisation and drug release activities are determined by the shape of the NPs. Several studies stated that the spherical NPs are better option for delivering drug as they can internalize rapidly than rod-shaped NPs [123]. Shape can also affect where NPs accumulate in the body. For example, spherical and cylindrical NPs accumulate in the liver while disc-shaped NPs accumulate in the heart [124].

A concentration dependent increase of FeCl<sub>3</sub> changed the morphology of the as-prepared moringa seeds samples, and the NMs appears small bulk or sheet shape with smooth surface. On the other hand, a few of folded nanosheets with crumpled surface seen without FeCl<sub>3</sub> can effectively prevent the nanosheets from stack, improve the open surface area, facilitate the electrolyte ion diffusion in the sheet-to-sheet, and offer more active sites to form electric double layers [125]. In general, the NPs with spherical shape can penetrate the bacteria and showed stronger antibacterial activity compared to NPs with other shapes (Fig. 10).



Fig. 10 A *M. oleifera* NPs with small size and spherical shape penetrate into the bacteria and show strong antibacterial activity. ET, Electron transport; NPs, Nanoparticles; ROS, Reactive oxygen species. **B** Antibacterial activity of spherical *M. oleifera* AgNPs compared to Vancomycin against different strains of bacteria. The original image was adapted with permission from Ref. [159]. 2023, Shadi Rahimi. **C** Antibacterial activity of spherical *M. oleifera* AgNPs. The original image was adapted from Ref. [158]

# Surface chemistry

NPs surface charge has an impact on their solubility, distribution, stability, absorption, and cellular toxicity. According to a series of studies, it is revealed that the positive ions on Ag are critical for their antimicrobial activity. The electrostatic interaction between the negatively charged cell membrane of microorganism and positively charged NPs is most likely what causes this activity [97]. Charge polarity and density both contribute to cytotoxic activity of NP. In nonphagocytic cells, positively charged ZnO, silica, silica-titania hollow and gold NPs are more cytotoxic than negatively charged ones [126]. Modulation of the surface charge on NPs impacts the immune response. Immune cells showed more inflammatory response when exposed to positively charged nanoparticles than when exposed to neutral or negatively charged NPs [127].

Particle surface can have a variety of defects, including atomic vacancies, variations in atomic coordinate, loose bonds, and structural disorder. Such surface imperfections may cause disturbed spins at the surface, which could cause N to become surface magnetised [128].

# Challenges and toxicity profiles associated with bio-synthesized nanoparticles

Nanoparticles synthesized through environmentally friendly methods are acknowledged for their extensive range of biological applications, particularly in serving as an effective mechanism for drug delivery [129, 130]. While nanoparticles derived from M. oleifera hold promise for diverse fields such as medicine and agriculture, it is essential to thoroughly assess their biosafety profile and toxicological concerns to ensure their safe application [131]. This recognition stems from their remarkable physicochemical attributes and behavioral properties. Nonetheless, as the utilization of these nanoparticles in biological contexts continues to expand, there exists considerable uncertainty regarding their safety in the human system. Due to their infinitesimal size and distinct characteristics, nanoparticles find frequent application in the realm of nanomedicine and as carriers for pharmaceutical agents. However, potential toxicity towards healthy human cells, tissues, and organs is a significant concern, influenced by factors such as crystallinity, solubility, aggregation, surface characteristics, morphology, surface area, and dosage-dependent attributes [132, 133]. It becomes imperative to investigate the nanoparticles' capacity to elicit an innate immune response. Various nanoparticles, including metal and metal-oxide nanoparticles, have been reported to induce pro-inflammatory effects in both in vitro and in vivo studies [134, 135].

Nanoparticles predominantly enter the human body through the respiratory system, where they often provoke

inflammatory responses, primarily driven by redox stress. Adding to the complexity, nanoparticles exhibit a remarkable capability to breach the blood-brain barrier, with the olfactory nerve hypothesized to serve as a conduit for their entry. This raises concerns about the potential impact of nanoparticles on the olfactory mucosa, from where they could traverse to the brain, exerting influence on both its health and functioning [136, 137]. Moreover, oxidative stress emerges as a central and extensively studied consequence of nanoparticle exposure. The heightened production of ROS, surpassing the capacity of antioxidants to neutralize them, establishes an oxidative stress condition [138, 139]. Notably, various byproducts of biological reactions, including hydroxyl radical, hydrogen peroxide, peroxynitrite, nitric oxide, and superoxide radical, represent key contributors to ROS generation. The ensuing ROS unleashes havoc on cellular components, causing damage to proteins, lipids, and crucial biomolecules. This cascade of molecular events triggers responses reminiscent of the activation of the nicotinamide adenine dinucleotide phosphate (NADPH) system [140]. Consequently, this disruption cascades through the electron transport chain, induces depolarization of the mitochondrial membrane, and alters the structural integrity of mitochondria, collectively posing significant implications for cellular and, consequently, overall health [133]. For instance, silver nanoparticles (AgNPs) when ingested potentially accumulated in all of the organs tested, and most predominantly distributed in intestine and stomach, skin and exhibits toxicity [141, 142]. In the skin, Ag induces blue-grey discoloration known as argyria. Animal studies indicated that their toxicity depends on dose-dependent exposure, causing death, weight loss, decreased activity, liver enzyme impairment, and immunological consequences [142]. Whereas, in kidney AgNPs resulted in the deposition of Ag in renal glomerular basement membrane [142, 143].

A study conducted by Vijaykumar et al., delineates, for the inaugural instance, the ecologically mindful synthesis of MgO NPs encapsulated within Moringa gum (Mgm-MgO NPs). A comprehensive evaluation was undertaken, encompassing assessments of their antioxidant efficacy, potential for hemolysis, cytotoxicity in human ovarian teratocarcinoma (PA-1) cells, phytotoxicity in mungbean (Vigna radiata), antiangiogenic attributes via the chorioallantoic membrane (CAM) chick embryo assay, and in vivo toxicity appraisals employing zebrafish embryos. Remarkably, the Mgm-MgO NPs exhibited notable antioxidant prowess. At a concentration of 500 µg/ml, these nanoparticles induced substantial hemolysis, registering at 72.54%, whereas lower concentrations demonstrated negligible hemolytic effects. The cytotoxic impact on PA-1 cells,

evaluated through the MTT assay, revealed a significant reduction in cell viability across a concentration range of 0.1–500  $\mu$ g/ml of Mgm-MgO NPs. Contrastingly, Mgm-MgO NPs displayed no discernible impact on seed germination but markedly influenced the root and shoot lengths of mungbean. Furthermore, the CAM assay, employed to scrutinize antiangiogenic potential, demonstrated no significant alterations following a 72-h exposure to Mgm-MgO NPs. Concluding the investigative spectrum, the embryotoxicity assay utilizing zebrafish embryos elucidated that Mgm-MgO NPs, within the concentration range of 0.1–500  $\mu$ g/ml, exhibited no deleterious effects on morphology, mortality, or survival rate [144].

In a study conducted by Hou and his colleagues, the impact of ZnONPs on cellular processes was investigated, revealing disruptions in DNA replication across various phases of the cell cycle pathway, including  $G_1$ , M, and  $G_2$ , alongside the impairment of mini-chromosome maintenance. The cytotoxic effects induced by nanoparticles were found to extend to multiple levels, affecting physicochemical, metabolic, and molecular pathways [145]. The size of NPs emerged as a noteworthy factor, with smaller particles exhibiting larger surface areas, enabling intricate interactions with cellular components such as carbohydrates, fatty acids, proteins, and nucleic acids. This observation suggests that particle size may play a role in influencing the cytotoxic efficacy of NPs. A significant contributor to the observed cytotoxicity was identified as the disturbance of intracellular calcium (Ca<sup>2+</sup>). Despite Ca<sup>2+</sup> being a pivotal signaling molecule involved in metabolic regulation, its elevated levels were found to induce acute toxicity on cellular mitochondria [146, 147]. This, in turn, triggered apoptosis through mechanisms such as the preferential release of cytochrome c, heightened ROS production, and the opening of the inner mitochondrial pore, ultimately culminating in the demise of the individual cell [148]. Contrastingly by a current investigation conducted by Mahfouz et al., entails the progression of a more reliable methodology for the biofabrication of zinc oxide nanoparticles (ZNPs) utilizing the green synthesis approach, employing *M. oleifera* extract (MO-ZNPs) as a highly efficient chelating agent for acrylamide (AA). The study delves into the impact of AA on glutathione redox dynamics, liver functionality, lipid profile, and zinc residues in Sprague Dawley rats. AA induced an elevation in liver enzymes, hepatosomatic index, and immunohistochemical manifestation of caspase-3 and CYP2E1 expression. Conversely, co-treatment with MO-ZNPs exhibited a stabilizing effect on the gene expression of glutathionerelated enzymes, normalization of hepatocellular enzyme levels, and restoration of hepatic tissue microarchitecture. This substantiates the proposition that MO-ZNPs

stands as a promising hepatoprotective agent, mitigating AA-induced hepatotoxicity [149].

For instance, recent findings indicate that the cytotoxic effects of nanoparticles extend beyond inducing cell death; they also encompass the suppression of cell growth when cells are arrested in specific cell cycle phases such as  $G_2/M$ , S, or  $G_0/G_1$ . Cells arrested in the cell cycle either accumulate significant damage, leading to apoptosis, or engage in repair mechanisms. The occurrence of cell cycle arrest is often cell-type specific and dependent on the particular stage of the cell cycle [150]. The primary toxicity of nanoparticles involves their interaction with DNA, while secondary genotoxicity is contributed by the ROS/RNS they produce. In the indirect primary clastogenic pathway, unsaturated aldehydes generated as a consequence of primary lipid oxidation by ROS serve as precursors for the formation of exocyclic DNA adducts. This intricate interplay underscores the multifaceted nature of nanoparticle-induced cytotoxicity, involving both direct interactions with genetic material and the generation of secondary genotoxic effects through oxidative stress pathways [140].

In many cases Moringa NPs have been found to reduce systemic toxicity caused by the effect of other nanoparticles. The primary objective of the current investigation employed by Abu Zeid et al., was twofold: firstly, to employ an environmentally benign approach for synthesizing selenium nanoparticles (SeNPs) utilizing M. oleifera leaf extract (MOLE); secondly, to discern and compare the protective attributes of the green-synthesized MOLE-SeNPs conjugate and MOLE ethanolic extract as interventions for mitigating melamine (MEL)induced nephrotoxicity in male rats. The nephrotoxic impact of MEL was characterized by an array of morphological alterations in the kidneys, coupled with an up-regulated immune-expression of proliferating cell nuclear antigen (PCNA) and proliferation-associated nuclear antigen Ki-67. Administration of either MOLE or MOLE-SeNPs significantly ameliorated MEL-induced renal functional impairments, oxidative stress, histological anomalies, adjustments in the relative mRNA expression of apoptosis-related genes, and the immuneexpression of renal PCNA and Ki-67. In conclusion, both the green-synthesized MOLE-SeNPs and MOLE exhibited nephron-protective properties against MEL-induced nephropathy in murine subjects. Remarkably, this study marks the pioneering disclosure of these effects, with a more pronounced impact observed in the MOLE group compared to the green biosynthesized MOLE-SeNPs conjugate group [151].

Numerous studies have extensively explored the integration of nanoparticles (NPs), such as copper oxide, into biomedical platforms [152]. However, there is a potential concern that these nanoparticles may hasten the process of protein oligomerization. A study conducted by Jaragh-Alhadad and Falahati aimed to elucidate the impact of CuONPs on the oligomerization of  $\beta$ 1–42 (A $\beta$ 1–42) and its associated neurotoxicity. The study uncovered crucial insights into the adverse effects of CuONPs on central nervous system proteins, which promote the formation of cytotoxic oligomers [153].

Understanding the toxicity of nanoparticles is integral to developing improved and more efficient nanomaterials. Consequently, extensive research is underway to enhance our overall understanding of the effects that nanoparticles exert on the environment and public health. This concerted effort not only seeks to mitigate potential risks but also aims to propel the development of safer materials with broader applications.

# **Future perspectives and conclusion**

In conclusion, the synthesis of NPs using *Moringa oleifera* extract stands as a remarkable testament to the plant's versatility and potential applications in nanotechnology. The diverse secondary metabolites, notably polyphenols, within *M. oleifera* have been identified as pivotal players in the synthesis process, influencing the size, shape, and biological efficacy of the resulting NPs. The distinctive roles played by these plant components, acting as reducing, stabilizing, and capping agents, have been elucidated, revealing the intricate interplay of biological molecules in the nanomaterial synthesis.

Currently, several studies are focusing on the translational clinical research of nanoparticles synthesized using Moringa extract. Some of them have shown promising effects in vivo for treating Leishmaniasis and Retinoblastoma [34, 154]. However biosynthetic techniques have limited control over the particle size, and this may be achieved by better control of temperature, and also adjusting the concentration of Moringa oleifera leaf extract. Apart from flavonoids and polyphenols, the role of alkaloids which are mainly cyclic nitrogen compounds (>12000) may also be evaluated thoroughly. Although several in vitro studies show the antimicrobial and antioxidant potential of nanoparticles synthesized using moringa extract, there is limited in vivo data which poses considerable hurdles in the translation of nanoparticles synthesized using Moringa extract. Furthermore, there is a major gap in the pharmacokinetics and underlying mechanisms responsible for cytotoxicity. Additionally, the optimal size and morphology necessary to elicit a favorable therapeutic outcome must be assessed. PEGylation has been successfully utilized to improve the bioavailability of nanoparticles synthesized using moringa extract [155]. However, studies have also shown that phenolic compounds present in the Moringa extract can

also serve as excellent capping agents [156] which may enhance the bioavailability and circulation, although it warrants further studies.

In essence, this comprehensive review underscores the multifaceted potential of *M. oleifera*-synthesized NPs in diverse biomedical applications. As we continue to unravel the intricacies of NP synthesis and their interactions with biological systems, the prospect of leveraging *M. oleifera* as a green tool for nanotechnology holds immense promise. Future research endeavors may focus on refining synthesis methods, optimizing NP characteristics, and elucidating specific molecular pathways for enhanced therapeutic outcomes. The convergence of plant-based synthesis and nanotechnology opens a captivating avenue for advancing bio-inspired materials with unprecedented functionalities, propelling us towards a future where *M. oleifera* nanoparticles play a pivotal role in transformative biomedical solutions.

#### Abbreviations

MPS	Mono nuclear phagocyte system
PEG	Polyethylene glycol
TTIP	Titanium tetra isopropoxide
HRTEM	High-resolution transmission electron microscopy
FT-IR	Fourier-transform infrared spectroscopy
AOM	Azoxymethane
ZOI	Zone of inhibition
rGO-TiO2	Phytoreduced graphene oxide-titanium dioxide
MRSA	Methicillin-resistant S. aureus
rGO-TiO2	Phytoreduced graphene oxide-titanium dioxide
DPPH	2,2-Diphenl-1-picrylhydrazyl

## Acknowledgements

No funding was received to carry out this research.

## Author contributions

H.P, SRD conceptualization, outline, literature search, writing, critical review, figures, and supervision; SR critical review, figures and supervision; JS, SN, DM writing, figures; MF & AH writing.

## Funding

Open access funding provided by Chalmers University of Technology.

#### Data availability

Data will be made available upon request.

# Declarations

### **Competing interests**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

## Author details

<sup>1</sup>Institute for Next Generation Material Design, Hanyang University, Seoul, Republic of Korea. <sup>2</sup>Center for Creative Convergence Education, Hanyang University, Seoul, Republic of Korea. <sup>3</sup>Department of Chemistry, College of Natural Sciences, Hanyang University, Seoul, Republic of Korea. <sup>4</sup>Department of Food Science and Biotechnology, Sejong University, Gwangjin-Gu, Seoul 05006, Republic of Korea. <sup>5</sup>Faculty of Biotechnology, University of Surabaya, Surabaya 60293, Indonesia. <sup>6</sup>Pharmacology Unit, Jeffrey Cheah School of Medicine and Health Sciences (JCSMHS), Monash University Malaysia, 47500 Bandar Sunway, Selangor Darul Ehsan, Malaysia. <sup>7</sup>School of Life Sciences, B.S. Abdur Rahman Crescent Institute of Science and Technology, Chennai, Tamil Nadu, India. <sup>8</sup>Department of Radiation Oncology, College of Medicine, Yonsei University, Seoul, South Korea. <sup>9</sup>Global Health Research, Saveetha Medical College, Saveetha Institute of Medical and Technical Sciences, Chennai, India. <sup>10</sup>Division of Systems and Synthetic Biology, Department of Life Sciences, Chalmers University of Technology, Gothenburg, Sweden.

# Received: 8 November 2023 Accepted: 7 February 2024 Published online: 19 February 2024

#### References

- Devkota HP, Gaire BP, Hori K, Subedi L, Adhikari-Devkota A, Belwal T, et al. The science of matcha: bioactive compounds, analytical techniques and biological properties. Trends Food Sci Technol. 2021;118:735–43.
- Nunes DAD, Salgado AM, da Gama-Rodrigues EF, Taketani RG, da Cunha CD, Sérvulo EFC. Use of plant materials for the bioremediation of soil from an industrial site. J Environ Sci Heal Part A. 2020;55:650–60. https:// doi.org/10.1080/10934529.2020.1726695.
- Srikar S, Giri D, Pal D, Mishra P, Upadhyay S. Green synthesis of silver nanoparticles: a review. Green Sustain Chem. 2016;6:34–56. https://doi. org/10.4236/gsc.2016.61004.
- Bindhu MR, Umadevi M, Esmail GA, Al-Dhabi NA, Arasu MV. Green synthesis and characterization of silver nanoparticles from Moringa oleifera flower and assessment of antimicrobial and sensing properties. J Photochem Photobiol B Biol. 2020;205:111836. https://doi.org/10. 1016/j.jphotobiol.2020.111836.
- Ezhilarasi AA, Vijaya JJ, Kaviyarasu K, Maaza M, Ayeshamariam A, Kennedy LJ. Green synthesis of NiO nanoparticles using Moringa oleifera extract and their biomedical applications: cytotoxicity effect of nanoparticles against HT-29 cancer cells. J Photochem Photobiol B Biol. 2016;164:352–60. https://doi.org/10.1016/j.jphotobiol.2016.10.003.
- Shahbazi MA, Faghfouri L, Ferreira MPA, Figueiredo P, Maleki H, Sefat F, et al. The versatile biomedical applications of bismuth-based nanoparticles and composites: therapeutic, diagnostic, biosensing, and regenerative properties. Chem Soc Rev. 2020;49:1253–321.
- Rabiee N, Bagherzadeh M, Kiani M, Ghadiri AM, Etessamifar F, Jaberizadeh AH, et al. Biosynthesis of copper oxide nanoparticles with potential biomedical applications. Int J Nanomed. 2020;15:3983–99.
- Abel S, Tesfaye JL, Nagaprasad N, Shanmugam R, Dwarampudi LP, Krishnaraj R. Synthesis and characterization of zinc oxide nanoparticles using Moringa leaf extract. J Nanomater. 2021;2021:4525770. https:// doi.org/10.1155/2021/4525770.
- Kaittanis C, Santra S, Asati A, Perez JM. A cerium oxide nanoparticlebased device for the detection of chronic inflammation via optical and magnetic resonance imaging. Nanoscale. 2012;4:2117–23.
- Irfan M, Munir H, Ismail H. Moringa oleifera gum based silver and zinc oxide nanoparticles: green synthesis, characterization and their antibacterial potential against MRSA. Biomater Res. 2021;25:17. https://doi.org/ 10.1186/s40824-021-00219-5.
- Gomez C, Hallot G, Pastor A, Laurent S, Brun E, Sicard-Roselli C, et al. Metallic bismuth nanoparticles: towards a robust, productive and ultrasound assisted synthesis from batch to flow-continuous chemistry. Ultrason Sonochem. 2019;56:167–73. https://doi.org/10.1016/j.ultso nch.2019.04.012.
- Katata-Seru L, Moremedi T, Aremu OS, Bahadur I. Green synthesis of iron nanoparticles using Moringa oleifera extracts and their applications: removal of nitrate from water and antibacterial activity against Escherichia coli. J Mol Liq. 2018;256:296–304. https://doi.org/10.1016/j. molliq.2017.11.093.
- Khor KZ, Joseph J, Shamsuddin F, Lim V, Moses EJ, Samad NA. The cytotoxic effects of moringa oleifera leaf extract and silver nanoparticles on human kasumi-1 cells. Int J Nanomed. 2020;15:5661–70.
- Mehwish HM, Rajoka MSR, Xiong Y, Cai H, Aadil RM, Mahmood Q, et al. Green synthesis of a silver nanoparticle using Moringa oleifera seed and its applications for antimicrobial and sun-light mediated photocatalytic

water detoxification. J Environ Chem Eng. 2021;9:105290. https://doi. org/10.1016/j.jece.2021.105290.

- Moodley JS, Krishna SBN, Pillay K, Sershen Govender P. Green synthesis of silver nanoparticles from Moringa oleifera leaf extracts and its antimicrobial potential. Adv Nat Sci Nanosci Nanotechnol. 2018;9:015011.
- Narendhran S, Manikandan M, Baby SP. Antibacterial, antioxidant properties of Solanum trilobatum and sodium hydroxide-mediated magnesium oxide nanoparticles: a green chemistry approach. Bull Mater Sci. 2019. https://doi.org/10.1007/s12034-019-1811-7.
- Ponnanikajamideen M, Rukumani V, Paulkumar K, Vanaja M, Samuel Rajendran R, Annadurai G. Moringa oleifera gum mediated fabrication of gold nanoparticles for effective DNA protection. J Bionanoscience. 2017;11:52–8.
- Pugazhendhi A, Prabhu R, Muruganantham K, Shanmuganathan R, Natarajan S. Anticancer, antimicrobial and photocatalytic activities of green synthesized magnesium oxide nanoparticles (MgONPs) using aqueous extract of Sargassum wightii. J Photochem Photobiol B Biol. 2019;190:86–97. https://doi.org/10.1016/j.jphotobiol.2018.11.014.
- 19. Rivas-San Vicente M, Plasencia J. Salicylic acid beyond defence: Its role in plant growth and development. J Exp Bot. 2011;62:3321–38.
- Rivas R, Oliveira MT, Santos MG. Three cycles of water deficit from seed to young plants of Moringa oleifera woody species improves stress tolerance. Plant Physiol Biochem. 2013;63:200–8.
- Anwar F, Latif S, Ashraf M, Gilani AH. Moringa oleifera: a food plant with multiple medicinal uses. Phyther Res. 2007;21:17–25. https://doi.org/10. 1002/ptr.2023.
- Leone A, Spada A, Battezzati A, Schiraldi A, Aristil J, Bertoli S. Cultivation, genetic, ethnopharmacology, phytochemistry and pharmacology of Moringa oleifera leaves: an overview. Int J Mol Sci. 2015;16:12791–835.
- Paikra BK, Dhongade HKJ, Gidwani B. Phytochemistry and pharmacology of Moringa oleifera lam. J pharmacopuncture. 2017;20:194–200.
- Saa RW, Fombang EN, Ndjantou EB, Njintang NY. Treatments and uses of Moringa oleifera seeds in human nutrition: a review. Food Sci Nutr. 2019;7:1911–9. https://doi.org/10.1002/fsn3.1057.
- Ueda Yamaguchi N, Cusioli LF, Quesada HB, Camargo Ferreira ME, Fagundes-Klen MR, Salcedo Vieira AM, et al. A review of Moringa oleifera seeds in water treatment: trends and future challenges. Process Saf Environ Prot. 2021;147:405–20.
- Baptista ATA, Coldebella PF, Cardines PHF, Gomes RG, Vieira MF, Bergamasco R, et al. Coagulation–flocculation process with ultrafiltered saline extract of Moringa oleifera for the treatment of surface water. Chem Eng J. 2015;276:166–73.
- dos Santos BS, Eyng E, Bittencourt PRS, Frare LM, de Flores ÉLM, Costa M. Electro-flocculation associated with the extract of *Moringa oleifera* Lam as natural coagulant for the removal of reactive blue 5G dye. Acta Sci Pol Hortorum Cultus Technol. 2016;38:483–8. https://doi.org/10. 4025/actascitechnol.v38i4.28796.
- Fahey JR. Microbiological monitoring of laboratory mice. In: Sundberg J, editor. Genetically engineered mice handbook. Boca Raton: CRC Press; 2016. p. 157–64.
- 29. Mahmood KT, Mugal T, Haq IU. Moringa oleifera: a natural gift-a review. J Pharm Sci Res. 2010;2:775–81.
- Ferreira PMP, Farias DF, de Oliveira JTA, de Carvalho AFU. Moringa oleifera: bioactive compounds and nutritional potential. Rev Nutr Scielo. 2008. https://doi.org/10.1590/S1415-52732008000400007.
- Busani M, Patrick JM, Arnold H, Voster M. Nutritional characterization of Moringa (Moringa oleifera Lam.) leaves. Afr J Biotechnol. 2011;10:12925–33. https://doi.org/10.5897/AJB10.1599.
- 32. Mahfuz S, Piao XS. Application of moringa (Moringa oleifera) as natural feed supplement in poultry diets. Animals. 2019;9:431.
- Shousha WG, Aboulthana WM, Salama AH, Saleh MH, Essawy EA. Evaluation of the biological activity of Moringa oleifera leaves extract after incorporating silver nanoparticles, in vitro study. Bull Natl Res Cent. 2019. https://doi.org/10.1186/s42269-019-0221-8.
- 34. El-Khadragy M, Alolayan EM, Metwally DM, El-Din MFS, Alobud SS, Alsultan NI, et al. Clinical efficacy associated with enhanced antioxidant enzyme activities of silver nanoparticles biosynthesized using moringa oleifera leaf extract, against cutaneous leishmaniasis in a murine model of leishmania major. Int J Environ Res Public Health. 2018;15:1037.
- 35. Aisida SO, Ugwoke E, Uwais A, Iroegbu C, Botha S, Ahmad I, et al. Incubation period induced biogenic synthesis of PEG enhanced Moringa

oleifera silver nanocapsules and its antibacterial activity. J Polym Res. 2019. https://doi.org/10.1007/s10965-019-1897-z.

- Paul S, Basak P, Majumder R, Mukherjee A, Ghosh J, Patra S, et al. Biochemical estimation of Moringa oleifera leaf extract for synthesis of silver nanoparticle mediated drug delivery system. J Plant Biochem Biotechnol. 2020;29:86–93. https://doi.org/10.1007/s13562-019-00517-z.
- Vasanth K, Ilango K, MohanKumar R, Agrawal A, Dubey GP. Anticancer activity of Moringa oleifera mediated silver nanoparticles on human cervical carcinoma cells by apoptosis induction. Colloids Surf B Biointerfaces. 2014;117:354–9. https://doi.org/10.1016/j.colsurfb.2014.02.052.
- Espenti CS, Rama Krishna AG, Rami Reddy YV. Green biosynthesis of ZnO nanomaterials and their anti-bacterial activity by using Moringa Oleifera root aqueous extract. SN Appl Sci. 2020;2:1–11. https://doi.org/ 10.1007/s42452-020-2945-3.
- Surendra TV, Roopan SM, Arasu MV, Al-Dhabi NA, Rayalu GM. RSM optimized Moringa oleifera peel extract for green synthesis of M. oleifera capped palladium nanoparticles with antibacterial and hemolytic property. J Photochem Photobiol B Biol. 2016;162:550–7. https://doi. org/10.1016/j.jphotobiol.2016.07.032.
- Aliyu AO, Garba S, Bognet O. Green synthesis, characterization and antimicrobial activity of vanadium nanoparticles using leaf extract of Moringa oleifera. Int J Chem Sci Res. 2017;16:231.
- Das PE, Majdalawieh AF, Abu-Yousef IA, Narasimhan S, Poltronieri P. Use of a hydroalcoholic extract of moringa oleifera leaves for the green synthesis of bismuth nanoparticles and evaluation of their anti-microbial and antioxidant activities. Materials (Basel). 2020;13:876.
- 42. Matinise N, Fuku XG, Kaviyarasu K, Mayedwa N, Maaza M. ZnO nanoparticles via Moringa oleifera green synthesis: physical properties & mechanism of formation. Appl Surf Sci. 2017;406:339–47.
- Katata-Seru L, Moremedi T, Aremu OS, Bahadur I. Green synthesis of iron nanoparticles using Moringa oleifera extracts and their applications: removal of nitrate from water and antibacterial activity against Escherichia coli. J Mol Liq. 2018;256:296–304.
- 44. Facts & Factors. Global Moringa Products Market Size Will Reach USD 8,400 Million by 2026: Facts & Factors. 2020; 1–3.
- Marslin G, Siram K, Maqbool Q, Selvakesavan RK, Kruszka D, Kachlicki P, et al. Secondary metabolites in the green synthesis of metallic nanoparticles. Materials (Basel). 2018;11:1–25.
- Mosallam FM, El-Sayyad GS, Fathy RM, El-Batal Al. Biomolecules-mediated synthesis of selenium nanoparticles using Aspergillus oryzae fermented Lupin extract and gamma radiation for hindering the growth of some multidrug-resistant bacteria and pathogenic fungi. Microb Pathog. 2018;122:108–16. https://doi.org/10.1016/j.micpath.2018.06. 013.
- Kuppusamy P, Yusoff MM, Maniam GP, Govindan N. Biosynthesis of metallic nanoparticles using plant derivatives and their new avenues in pharmacological applications – an updated report. Saudi Pharm J. 2016;24:473–84. https://doi.org/10.1016/j.jsps.2014.11.013.
- Virk P, Awad MA, Saleh Abdu-Ilah Alsaif S, Hendi AA, Elobeid M, Ortashi K, et al. Green synthesis of Moringa oleifera leaf nanoparticles and an assessment of their therapeutic potential. J King Saud Univ Sci. 2023;35:102576. https://doi.org/10.1016/j.jksus.2023.102576.
- Barman A, Kotal A, Das M. Synthesis of metal based nano particles from Moringa Olifera and its biomedical applications: a review. Inorg Chem Commun. 2023;158:111438. https://doi.org/10.1016/j.inoche.2023. 111438.
- Naren Vidaarth TM, Surendhiran S, Jagan KSG, Savitha S, Balu KS, Karthik A, et al. Surface chemistry of phytochemical enriched MgO nanoparticles for antibacterial, antioxidant, and textile dye degradation applications. J Photochem Photobiol A Chem. 2024;448:115349. https://doi. org/10.1016/j.jphotochem.2023.115349.
- Barathikannan K, Chelliah R, Selvakumar V, Elahi F, Rubab M, Sanyal S, et al. Plant-based metabolites and their uses in nanomaterials synthesis: an overview. In: Husen A, editor., et al., Secondary metabolites based green synthesis of nanomaterials and their applications. Springer Nature Singapore: Singapore; 2023. p. 1–22. https://doi.org/10.1007/ 978-981-99-0927-8\_1.
- El-Seedi HR, El-Shabasy RM, Khalifa SAM, Saeed A, Shah A, Shah R, et al. Metal nanoparticles fabricated by green chemistry using natural extracts: biosynthesis, mechanisms, and applications. RSC Adv. 2019;9:24539–59.

- Shelke DB, Islam NF, Chambhare MR, Sonawane HB, Patowary R, Prasad R, et al. Enhancing secondary metabolites and alleviating environmental stress in crops with mycogenic nanoparticles: a comprehensive review. Biocatal Agric Biotechnol. 2023;52:102805. https://doi.org/10. 1016/j.bcab.2023.102805.
- Kandeil MA, Mohammed ET, Hashem KS, Aleya L, Abdel-Daim MM. Moringa seed extract alleviates titanium oxide nanoparticles (TiO2-NPs)induced cerebral oxidative damage, and increases cerebral mitochondrial viability. Environ Sci Pollut Res. 2020;27:19169–84.
- Elumalai EK, Prasad TNVKV, Hemachandran J, Viviyan Therasa S, Thirumalai T, David E. Extracellular synthesis of silver nanoparticles using leaves of Euphorbia hirta and their antibacterial activities. J Pharm Sci Res. 2010;2:549–54.
- Amsden BG, Sukarto A, Knight DK, Shapka SN. Methacrylated glycol chitosan as a photopolymerizable biomaterial. Biomacromol. 2007;8:3758–66.
- Sathyavathi R, Bala Murali Krishna M, Rao ND. Biosynthesis of silver nanoparticles using Moringa oleifera leaf extract and its application to optical limiting. J Nanosci Nanotechnol. 2011;11:2031–5.
- Mokgweetsi P. Green synthesis of silver nanoparticles using Moringa oleifera, employing multivariate optimization methodologies. Int J Adv Res. 2018;6:953–62.
- Diana EP, Sumiati S, Nur ATN. Synthesis of silver nanoparticles using moringa oleifera I leaf extract as bioreductor. Mater Sci Forum. 2019;967:145–9.
- Kalugendo E, Kousalya P. Synthesis of silver nanoparticles using Moringa Oleifera seeds, glycyrrhiza glabra stems, and its anti-methicillin-resistant staphylococcus aureus activity. Asian J Pharm Clin Res. 2019;12:368–70.
- Eric CA, Benjamín V, Monica C, Mario AC, Francisco DM, Rogelio AR, et al. Silver nanoparticles biosynthesized by secondary metabolites from Moringa oleifera stem and their antimicrobial properties. African J Biotechnol. 2017;16:400–7.
- 62. Mohammed GM, Hawar SN. Green biosynthesis of silver nanoparticles from moringa oleifera leaves and its antimicrobial and cytotoxicity activities. Int J Biomater. 2022;2022:4136641.
- Ghosh N, Paul S, Basak P. Silver nanoparticles of Moringa Oleifera green synthesis, characterisation and its antimicrobial efficacy. J Drug Deliv Ther. 2014. https://doi.org/10.22270/jddt.v0i0.906.
- 64. Belliraj TS, Nanda A, Ragunathan R. In-vitro hepatoprotective activity of Moringa oleifera mediated synthesis of gold nanoparticles. J Chem Pharm Res. 2015;7:781–8.
- 65. Tiloke C, Phulukdaree A, Anand K, Gengan RM, Chuturgoon AA. Moringa oleifera gold nanoparticles modulate oncogenes, tumor suppressor genes, and caspase-9 splice variants in A549 cells. J Cell Biochem. 2016;13:2302–14.
- Ponnanikajamideen M, Rukumani V, Paulkumar K, Vanaja M, Rajendran RS, Annadurai G. Moringa oleifera gum mediated fabrication of gold nanoparticles for effective DNA protection. J Bionanosci. 2017;11:52–8. https://doi.org/10.1166/jbns.2017.1411.
- Chakraborty A, Das DK, Sinha M, Dey S, Bhattacharjee S. Moringa oleifera leaf extract mediated green synthesis of stabilized gold nanoparticles. J Bionanoscience. 2013;7:415–9.
- Anand K, Gengan RM, Phulukdaree A, Chuturgoon A. Agroforestry waste moringa oleifera petals mediated green synthesis of gold nanoparticles and their anti-cancer and catalytic activity. J Ind Eng Chem. 2015;21:1105–11. https://doi.org/10.1016/j.jiec.2014.05.021.
- 69. Vivek P, Preeti J. Green synthesis of TiO 2 nanoparticle using. Int Res J Eng Technol. 2017;4:1–7.
- Sivaranjani V, Philominathan P. Synthesize of Titanium dioxide nanoparticles using Moringa oleifera leaves and evaluation of wound healing activity. Wound Med. 2016;12:1–5. https://doi.org/10.1016/j.wndm. 2015.11.002.
- Pushpamalini T, Keerthana M, Sangavi R, Nagaraj A, Kamaraj P. Comparative analysis of green synthesis of TiO2 nanoparticles using four different leaf extract. Mater Today Proc. 2020;40:S180-4. https://doi.org/ 10.1016/j.matpr.2020.08.438.
- Satti SH, Raja NI, Javed B, Akram A, Mashwani ZR, Ahmad MS, et al. Titanium dioxide nanoparticles elicited agro-morphological and physicochemical modifications in wheat plants to control Bipolaris sorokiniana. PLoS ONE. 2021;16:1–19. https://doi.org/10.1371/journal.pone.0246880.

- Umekar MS, Chaudhary RG, Bhusari GS, Mondal A, Potbhare AK, Mahmood S. Phytoreduced graphene oxide-titanium dioxide nanocomposites using Moringa oleifera stick extract. Mater Today Proc. 2020;29:709–14. https://doi.org/10.1016/j.matpr.2020.04.169.
- Prucek R, Tuček J, Kilianová M, Panáček A, Kvítek L, Filip J, et al. The targeted antibacterial and antifungal properties of magnetic nanocomposite of iron oxide and silver nanoparticles. Biomaterials. 2011;32:4704–13.
- Madubuonu N, Aisida SO, Ali A, Ahmad I, Zhao TK, Botha S, et al. Biosynthesis of iron oxide nanoparticles via a composite of Psidium guavaja-Moringa oleifera and their antibacterial and photocatalytic study. J Photochem Photobiol B Biol. 2019;199:111601. https://doi.org/ 10.1016/j.jphotobiol.2019.111601.
- Aisida SO, Madubuonu N, Alnasir MH, Ahmad I, Botha S, Maaza M, et al. Biogenic synthesis of iron oxide nanorods using Moringa oleifera leaf extract for antibacterial applications. Appl Nanosci. 2020;10:305–15. https://doi.org/10.1007/s13204-019-01099-x.
- Silveira C, Shimabuku QL, Fernandes Silva M, Bergamasco R. Iron-oxide nanoparticles by the green synthesis method using Moringa oleifera leaf extract for fluoride removal. Environ Technol (United Kingdom). 2018;39:2926–36.
- Jegadeesan GB, Srimathi K, Santosh Srinivas N, Manishkanna S, Vignesh D. Green synthesis of iron oxide nanoparticles using Terminalia bellirica and Moringa oleifera fruit and leaf extracts: Antioxidant, antibacterial and thermoacoustic properties. Biocatal Agric Biotechnol. 2019;21:101354. https://doi.org/10.1016/j.bcab.2019.101354.
- Mateus GAP, Paludo MP, Dos Santos TRT, Silva MF, Nishi L, Fagundes-Klen MR, et al. Obtaining drinking water using a magnetic coagulant composed of magnetite nanoparticles functionalized with Moringa oleifera seed extract. J Environ Chem Eng. 2018;6:4084–92. https://doi. org/10.1016/j.jece.2018.05.050.
- Kalaiselvan S, Angulakshmi VS, Mageswari S, Karthikeyan S. Carbon nanotubes from plant derived hydrocarbon - an efficient renewable precursor. J Environ Nanotechnol. 2018;7:41–6.
- Kiwumulo HF, Muwonge H, Ibingira C, Lubwama M, Kirabira JB, Ssekitoleko RT. Green synthesis and characterization of iron-oxide nanoparticles using Moringa oleifera: a potential protocol for use in low and middle income countries. BMC Res Notes. 2022;15:149. https://doi. org/10.1186/s13104-022-06039-7.
- 82. Cheng G, Zheng SY. Construction of a high-performance magnetic enzyme nanosystem for rapid tryptic digestion. Sci Rep. 2014;4:1–10.
- Pal S, Mondal S, Maity J, Mukherjee R. Synthesis and characterization of zno nanoparticles using Moringa Oleifera leaf extract: Investigation of photocatalytic and antibacterial activity. Int J Nanosci Nanotechnol. 2018;14:111–9.
- Ngom I, Ngom BD, Sackey J, Khamlich S. Biosynthesis of zinc oxide nanoparticles using extracts of Moringa Oleifera: structural & optical properties. Mater Today Proc. 2019;36:526–33. https://doi.org/10.1016/j. matpr.2020.05.323.
- Ngom I, Ndiayea NM, Bakayoko AFM, Ngom BD, Maaza M. On the use of Moringa oleifera leaves extract for the biosynthesis of NiO and ZnO nanoparticles. MRS Adv. 2020;5:1145–55.
- Srivastava V, Choubey AK. Kinetic and isothermal study of effect of transition metal doping on adsorptive property of zinc oxide nanoparticles synthesized via green route using Moringa oleifera leaf extract. Mater Res Express. 2019. https://doi.org/10.1088/2053-1591/ab7158.
- Gondwal M, Pant GJN. Synthesis and catalytic and biological activities of silver and copper nanoparticles using cassia occidentalis. Int J Biomater. 2018. https://doi.org/10.1155/2018/6735426.
- Kombaiah K, Vijaya JJ, Kennedy LJ, Bououdina M, Ramalingam RJ, Al-Lohedan HA. Okra extract-assisted green synthesis of CoFe2O4 nanoparticles and their optical, magnetic, and antimicrobial properties. Mater Chem Phys. 2018;204:410–9. https://doi.org/10.1016/j.match emphys.2017.10.077.
- Sur UK, Ankamwar B. Optical, dielectric, electronic and morphological study of biologically synthesized zinc sulphide nanoparticles using: Moringa oleifera leaf extract and quantitative analysis of chemical components present in the leaf extract. RSC Adv. 2016;6:95611–9.
- Abdal Dayem A, Hossain MK, Bin Lee S, Kim K, Saha SK, Yang G-M, et al. The role of reactive oxygen species (ROS) in the biological activities of metallic nanoparticles. Int J Mol Sci. 2017;18:120.

- 91. Althomali A, Daghestani MH, Basil Almukaynizi F, Al-Zahrani SA, Awad MA, Merghani NM, et al. Anti-colon cancer activities of green-synthesized Moringa oleifera-AgNPs against human colon cancer cells. Green Process Synth. 2022;11:545–54.
- Gupta S, Hemlata H, Tejavath KK. Synthesis, characterization and comparative anticancer potential of phytosynthesized mono and bimetallic nanoparticles using Moringa oleifera aqueous leaf extract. Beilstein Arch. 2020;1:1–16.
- Sahu RK, Aboulthana WM, Mehta DK. Phyto-phospholipid complexation as a novel drug delivery system for management of cancer with better bioavailability: current perspectives and future prospects. Anticancer Agents Med Chem. 2021;21:1403–12.
- Muhammad S, Ali A, Shah J, Hamza M, Kashif M, Ajat Khel BK, et al. Using Moringa oleifera stem extract for green synthesis, characterization, and anti-inflammatory activity of silver oxide nanoparticles. Nat Appl Sci Int J. 2023;4:80–97.
- Bindhu MR, Umadevi M, Esmail GA, Al-Dhabi NA, Arasu MV. Green synthesis and characterization of silver nanoparticles from Moringa oleifera flower and assessment of antimicrobial and sensing properties. J Photochem Photobiol B Biol. 2020;205:781–8.
- Kalaiselvi V, Mathammal R, Vijayakumar S, Vaseeharan B. Microwave assisted green synthesis of hydroxyapatite nanorods using Moringa oleifera flower extract and its antimicrobial applications. Int J Vet Sci Med. 2018;6:286–95. https://doi.org/10.1016/j.ijvsm.2018.08.003.
- Prasad TNVKV, Elumalai EK. Biofabrication of Ag nanoparticles using Moringa oleifera leaf extract and their antimicrobial activity. Asian Pac J Trop Biomed. 2011;1:439–42.
- Narwade B, Prasad N, Lokhande SM, Madavi AB, Sahoo AK. Extracellular biosynthesis of silver nanoparticles using Moringa oleifera leaves extract and its antimicrobial efficacy in packaging materials. Res J life Sci Chem Sci. 2018;4:188–202.
- Mirza AU, Kareem A, Nami SAA, Khan MS, Rehman S, Bhat SA, et al. Biogenic synthesis of iron oxide nanoparticles using Agrewia optiva and Prunus persica phyto species: Characterization, antibacterial and antioxidant activity. J Photochem Photobiol B Biol. 2018;185:262–74. https://doi.org/10.1016/j.jphotobiol.2018.06.009.
- Jadhav V, Bhagare A, Ali IH, Dhayagude A, Lokhande D, Aher J, et al. Role of Moringa oleifera on green synthesis of metal/metal oxide nanomaterials. J Nanomater. 2022. https://doi.org/10.1155/2022/2147393.
- Boruah JS, Devi C, Hazarika U, Bhaskar Reddy PV, Chowdhury D, Barthakur M, et al. Green synthesis of gold nanoparticles using an antiepileptic plant extract: in vitro biological and photo-catalytic activities. RSC Adv. 2021;11:28029–41.
- 102. Akintunde JK, Farai TI, Arogundade MR, Adeleke JT. Biogenic zinc-oxide nanoparticles of Moringa oleifera leaves abrogates rotenone induced neuroendocrine toxicity by regulation of oxidative stress and acetylcho-linesterase activity. Biochem Biophys Rep. 2021;26:100999.
- Olaoye AB, Ologunde CA, Molehin OR, Nwankwo I. Comparative antioxidant analysis of Moringa oleifera leaf extracts from South Western States in Nigeria. Futur J Pharm Sci. 2021;7:68. https://doi.org/10.1186/ s43094-021-00204-8.
- Gulshan F, Kameshima Y, Nakajima A, Okada K. Preparation of aluminairon oxide compounds by gel evaporation method and its simultaneous uptake properties for Ni2+, NH4+ and H2PO4-. J Hazard Mater. 2009;169:697–702.
- 105. Zhang J, Han X, Li X, Luo Y, Zhao H, Yang M, et al. Core-shell hybrid liposomal vesicles loaded with panax notoginsenoside: preparation, characterization and protective effects on global cerebral ischemia/reperfusion injury and acute myocardial ischemia in rats. Int J Nanomed. 2012;7:4299–310.
- Hoshyar N, Gray S, Han H, Bao G. The effect of nanoparticle size on in vivo pharmacokinetics and cellular interaction. Nanomedicine (Lond). 2016;11:673–92.
- Contini C, Di Nuzzo M, Barp N, Bonazza A, De Giorgio R, Tognon M, et al. The novel zoonotic COVID-19 pandemic: an expected global health concern. J Infect Dev Ctries. 2020;14:254–64.
- Salomoni R, Léo P, Montemor AF, Rinaldi BG, Rodrigues M. Antibacterial effect of silver nanoparticles in Pseudomonas aeruginosa. Nanotechnol Sci Appl. 2017;10:115–21.
- 109. Anand K, Tiloke C, Phulukdaree A, Ranjan B, Chuturgoon A, Singh S, et al. Biosynthesis of palladium nanoparticles by using Moringa oleifera

flower extract and their catalytic and biological properties. J Photochem Photobiol B Biol. 2016;165:87–95. https://doi.org/10.1016/j.jphot obiol.2016.09.039.

- 110. Augustine R, Hasan A, Primavera R, Wilson RJ, Thakor AS, Kevadiya BD. Cellular uptake and retention of nanoparticles: insights on particle properties and interaction with cellular components. Mater Today Commun. 2020;25:101692.
- 111. Liu Y, Perumalsamy H, Kang CH, Kim SH, Hwang JS, Koh SC, et al. Intracellular synthesis of gold nanoparticles by Gluconacetobacter liquefaciens for delivery of peptide CopA3 and ginsenoside and antiinflammatory effect on lipopolysaccharide-activated macrophages. Artif Cells Nanomed Biotechnol. 2020;48:777–88.
- Bourquin J, Milosevic A, Hauser D, Lehner R, Blank F, Petri-Fink A, et al. Biodistribution, clearance, and long-term fate of clinically relevant nanomaterials. Adv Mater. 2018;30:1704307. https://doi.org/10.1002/ adma.201704307.
- Dikshit PK, Kumar J, Das AK, Sadhu S, Sharma S, Singh S, et al. Green synthesis of metallic nanoparticles: applications and limitations. Catalysts. 2021;11:902.
- Yagublu V, Karimova A, Hajibabazadeh J, Reissfelder C, Muradov M, Bellucci S, et al. Overview of physicochemical properties of nanoparticles as drug carriers for targeted cancer therapy. J Funct Biomater. 2022;13:196.
- Bartczak D, Muskens OL, Nitti S, Sanchez-Elsner T, Millar TM, Kanaras AG. Interactions of human endothelial cells with gold nanoparticles of different morphologies. Small. 2012;8:122–30.
- Alananbeh KM, Refaee WJA, Al-Qodah Z. Antifungal effect of silver nanoparticles on selected fungi isolated from raw and waste water. Indian J Pharm Sci. 2017;79:559–67.
- Fatiqin A, Amrulloh H, Simanjuntak W. Green synthesis of MgO nanoparticles using Moringa oleifera leaf aqueous extract for antibacterial activity. Bull Chem Soc Ethiop. 2021;35:161–70.
- 118. Imran Din M, Rani A. Recent advances in the synthesis and stabilization of nickel and nickel oxide nanoparticles: a green adeptness. Int J Anal Chem. 2016. https://doi.org/10.1155/2016/3512145.
- 119. Espinosa-Cristóbal LF, Holguín-Meráz C, Zaragoza-Contreras EA, Martínez-Martínez RE, Donohue-Cornejo A, Loyola-Rodríguez JP, et al. Antimicrobial and substantivity properties of silver nanoparticles against oral microbiomes clinically isolated from young and youngadult patients. J Nanomater. 2019;2019:3205971. https://doi.org/10. 1155/2019/3205971.
- Moghimi SM, Hunter AC, Murray JC. Long-circulating and target-specific nanoparticles: theory to practice. Pharmacol Rev. 2001;53:283–318.
- Banerjee A, Qi J, Gogoi R, Wong J, Mitragotri S. Role of nanoparticle size, shape and surface chemistry in oral drug delivery. J Control Release. 2016;238:176–85.
- Tan J, Shah S, Thomas A, Ou-Yang HD, Liu Y. The influence of size, shape and vessel geometry on nanoparticle distribution. Microfluid Nanofluidics. 2013;14:77–87.
- 123. Chithrani BD, Ghazani AA, Chan WCW. Determining the size and shape dependence of gold nanoparticle uptake into mammalian cells. Nano Lett. 2006;6:662–8. https://doi.org/10.1021/nl0523960.
- Devarajan PV, Jindal AB, Patil RR, Mulla F, Gaikwad RV, Samad A. Particle shape: a new design parameter for passive targeting in splenotropic drug delivery. J Pharm Sci. 2010;99:2576–81. https://doi.org/10.1002/ jps.22052.
- 125. Cai Y, Luo Y, Dong H, Zhao X, Xiao Y, Liang Y, et al. Hierarchically porous carbon nanosheets derived from Moringa oleifera stems as electrode material for high-performance electric double-layer capacitors. J Power Sour. 2017;353:260–9. https://doi.org/10.1016/j.jpowsour.2017.04.021.
- 126. Frühbeis C, Fröhlich D, Krämer-Albers E-M. Emerging roles of exosomes in neuron-glia communication. Front Physiol. 2012;3:119.
- 127. Vangeel L, Chiu W, De Jonghe S, Maes P, Slechten B, Raymenants J, et al. Remdesivir, molnupiravir and nirmatrelvir remain active against SARS-CoV-2 omicron and other variants of concern. Antiviral Res. 2022;198:105252.
- Issa N, Mayer RE, Schuller M, Wang E, Shapiro MB, DaRosa DA. Teaching for understanding in medical classrooms using multimedia design principles. Med Educ. 2013;47:388–96. https://doi.org/10.1111/medu. 12127.

- 129. Lewinski N, Colvin V, Drezek R. Cytotoxicity of nanopartides. Small. 2008;4:26–49.
- 130. Ettlinger R, Lächelt U, Gref R, Horcajada P, Lammers T, Serre C, et al. Toxicity of metal-organic framework nanoparticles: from essential analyses to potential applications. Chem Soc Rev. 2022;51:464–84.
- 131. Hamdy NM, Boseila AA, Ramadan A, Basalious EB. Iron oxide nanoparticles-plant insignia synthesis with favorable biomedical activities and less toxicity, in the "era of the-green": a systematic review. Pharmaceutics. 2022;14:844.
- Zhou Y, Chen Y, Rocha A, Sanchez CJ, Liang H. Assessment of toxicity of nanoparticles using insects as biological models. In: Ferrari E, Soloviev M, editors. Nanoparticles in biology and medicine: methods and protocols. New York: Springer US; 2020. p. 269–79. https://doi.org/10.1007/ 978-1-0716-0319-2\_20.
- Dantas GPF, Ferraz FS, Andrade LM, Costa GMJ. Male reproductive toxicity of inorganic nanoparticles in rodent models: a systematic review. Chem Biol Interact. 2022;363:110023. https://doi.org/10.1016/j.cbi.2022. 110023.
- Stalder T, Zaiter T, El-Basset W, Cornu R, Martin H, Diab-Assaf M, et al. Interaction and toxicity of ingested nanoparticles on the intestinal barrier. Toxicology. 2022;481:153353.
- Habas K, Demir E, Guo C, Brinkworth MH, Anderson D. Toxicity mechanisms of nanoparticles in the male reproductive system. Drug Metab Rev. 2021;53:604–17. https://doi.org/10.1080/03602532.2021.1917597.
- 136. Yokel R, Grulke E, Macphail R. Metal-based nanoparticle interactions with the nervous system: the challenge of brain entry and the risk of retention in the organism. Wiley Interdiscip Rev Nanomed Nanobio-technol. 2013;5:346–73.
- Srivastava A, Babu A, Filant J, Moxley KM, Ruskin R, Dhanasekaran D, et al. Exploitation of exosomes as nanocarriers for gene-, chemo-, and immune-therapy of cancer. J Biomed Nanotechnol. 2016;12:1159–73.
- Makhdoumi P, Karimi H, Khazaei M. Review on metal-based nanoparticles: role of reactive oxygen species in renal toxicity. Chem Res Toxicol. 2020;33:2503–14. https://doi.org/10.1021/acs.chemrestox.9b00438.
- 139. Wang Z, Tang M. Research progress on toxicity, function, and mechanism of metal oxide nanoparticles on vascular endothelial cells. J Appl Toxicol. 2021;41:683–700.
- Horie M, Tabei Y. Role of oxidative stress in nanoparticles toxicity. Free Radic Res. 2021;55:331–42. https://doi.org/10.1080/10715762.2020. 1859108.
- 141. van der Zande M, Vandebriel RJ, Van Doren E, Kramer E, Herrera Rivera Z, Serrano-Rojero CS, et al. Distribution, elimination, and toxicity of silver nanoparticles and silver ions in rats after 28-day oral exposure. ACS Nano. 2012;6:7427–42. https://doi.org/10.1021/nn302649p.
- 142. Gaillet S, Rouanet J-M. Silver nanoparticles: their potential toxic effects after oral exposure and underlying mechanisms—a review. Food Chem Toxicol. 2015;77:58–63.
- Creasey M, Moffat DB. The deposition of ingested silver in the rat kidney at different ages. Experientia. 1973;29:326–7. https://doi.org/10.1007/ BF01926506.
- 144. Vijayakumar S, Chen J, González Sánchez ZI, Tungare K, Bhori M, Durán-Lara EF, et al. Moringa oleifera gum capped MgO nanoparticles: synthesis, characterization, cyto- and ecotoxicity assessment. Int J Biol Macromol. 2023;233:123514.
- Hou J, Liu H, Zhang S, Liu X, Hayat T, Alsaedi A, et al. Mechanism of toxic effects of Nano-ZnO on cell cycle of zebrafish (Danio rerio). Chemosphere. 2019;229:206–13.
- Lovisolo D, Gilardino A, Ruffinatti FA. When neurons encounter nanoobjects: spotlight on calcium signalling. Int J Environ Res Public Health. 2014;11:9621–37.
- Huang YW, Cambre M, Lee HJ. The toxicity of nanoparticles depends on multiple molecular and physicochemical mechanisms. Int J Mol Sci. 2017;18:2702.
- 148. Engin AB, Engin A. Chapter 9 nanoparticles and neurotoxicity: dual response of glutamatergic receptors. In: Sharma A, Sharma HSBT, editors. Nanoneuroprotection and nanoneurotoxicology. Amsterdam: Elsevier; 2019. p. 281–303.
- 149. Mahfouz H, Dahran N, Abdel-Rahman Mohamed A, Abd El-Hakim YM, Metwally MMM, Alqahtani LS, et al. Stabilization of glutathione redox dynamics and CYP2E1 by green synthesized Moringa oleifera-mediated zinc oxide nanoparticles against acrylamide induced hepatotoxicity

in rat model: morphometric and molecular perspectives. Food Chem Toxicol. 2023;176:113744.

- 150. Gao F, Ma NJ, Zhou H, Wang Q, Zhang H, Wang P, et al. Zinc oxide nanoparticles-induced epigenetic change and G2/M arrest are associated with apoptosis in human epidermal keratinocytes. Int J Nanomed. 2016;11:3859–74.
- 151. Abu-Zeid EH, Abdel Fattah DM, Arisha AH, Ismail TA, Alsadek DM, Metwally MMM, et al. Protective prospects of eco-friendly synthesized selenium nanoparticles using Moringa oleifera or Moringa oleifera leaf extract against melamine induced nephrotoxicity in male rats. Ecotoxicol Environ Saf. 2021;221:112424. https://doi.org/10.1016/j.ecoenv.2021. 112424.
- 152. El-Batal AI, Al-Hazmi NE, Mosallam FM, El-Sayyad GS. Biogenic synthesis of copper nanoparticles by natural polysaccharides and Pleurotus ostreatus fermented fenugreek using gamma rays with antioxidant and antimicrobial potential towards some wound pathogens. Microb Pathog. 2018;118:159–69.
- 153. Jaragh-Alhadad LA, Falahati M. Copper oxide nanoparticles promote amyloid-β-triggered neurotoxicity through formation of oligomeric species as a prelude to Alzheimer's diseases. Int J Biol Macromol. 2022;207:121–9. https://doi.org/10.1016/j.ijbiomac.2022.03.006.
- Mudigunda SV, Pemmaraju DB, Sankaranarayanan SA, Rengan AK. Bioactive polymeric nanoparticles of moringa oleifera induced phytophotothermal sensitization for the enhanced therapy of retinoblastoma. Pharmaceutics. 2023;15:475.
- 155. Al-Qaraleh SY, Al-Zereini WA, Oran SA, Al-Sarayreh AZ, Al-Dalain SM. Evaluation of the antioxidant activities of green synthesized selenium nanoparticles and their conjugated polyethylene glycol (PEG) form in vivo. OpenNano. 2022;8:100109.
- Shalaby EA, Shanab SMM, El-Raheem WMA, Hanafy EA. Biological activities and antioxidant potential of different biosynthesized nanoparticles of Moringa oleifera. Sci Rep. 2022;12:18400. https://doi.org/10.1038/ s41598-022-23164-2.
- Tiloke C, Anand K, Gengan RM, Chuturgoon AA. Moringa oleifera and their phytonanoparticles: Potential antiproliferative agents against cancer. Biomed Pharmacother. 2018;108:457–66. https://doi.org/10.1016/j. biopha.2018.09.060.
- Mohammed ABA, Mohamed A, El-Naggar NEA, Mahrous H, Nasr GM, Abdella A, et al. Antioxidant and antibacterial activities of silver nanoparticles biosynthesized by Moringa oleifera through response surface methodology. J Nanomater. 2022. https://doi.org/10.1155/2022/99843 08.
- 159. Abdel-Rahman LH, Al-Farhan BS, Abou El-ezz D, Abd-El Sayed MA, Zikry MM, Abu-Dief AM. Green biogenic synthesis of silver nanoparticles using aqueous extract of Moringa oleifera: access to a powerful antimicrobial, anticancer, pesticidal and catalytic agents. J Inorg Organomet Polym Mater. 2022;32:1422–35. https://doi.org/10.1007/ s10904-021-02186-9.
- Bhusnure OG, Jadhav PP, Hindole SS, Gholve SB, Giram PS, Kuthar VS. Green synthesis of silver nanoparticle using Moringa oleifera lam extract for pharmacological activity. Int J Pharm Pharm Res. 2017;10:59–66.
- 161. Azeez L, Adejumo AL, Lateef A, Adebisi SA, Adetoro RO, Adewuyi SO, et al. Zero-valent silver nanoparticles attenuate Cd and Pb toxicities on Moringa oleifera via immobilization and induction of phytochemicals. Plant Physiol Biochem. 2019;139:283–92. https://doi.org/10.1016/j. plaphy.2019.03.030.
- 162. Merugu R, Garimella S, Velamakanni R, Vuppugalla P, Chitturi KL, Jyothi M. Synthesis, characterization and antimicrobial activity of bimetallic silver and copper nanoparticles using fruit pulp aqueous extracts of Moringa oleifera. Mater Today Proc. 2021;44:153–6. https://doi.org/10. 1016/j.matpr.2020.08.549.
- 163. Madubuonu N, Aisida SO, Ali A, Ahmad I, Zhao T, Botha S, et al. Biosynthesis of iron oxide nanoparticles via a composite of Psidium guavaja-Moringa oleifera and their antibacterial and photocatalytic study. J Photochem Photobiol B Biol. 2019;199:111601.
- Bahador F, Foroutan R, Esmaeili H, Ramavandi B. Enhancement of the chromium removal behavior of Moringa oleifera activated carbon by chitosan and iron oxide nanoparticles from water. Carbohydr Polym. 2021;251:117085.

- 165. Vázquez-Guerrero A, Cortés-Martínez R, Alfaro-Cuevas-Villanueva R, Rivera-Muñoz EM, Huirache-Acuña R. Cd(II) and Pb(II) adsorption using a composite obtained from Moringa oleifera Lam. Cellulose nanofibrils impregnated with iron nanoparticles. Water. 2021;13:89.
- 166. Santos TM, da Silva JV, da Silva GF, Pontes LAM. Development of a lowcost adsorbent obtained from Moringa oleifera and functionalized with iron nanoparticles for removal of oil from produced water. Biointerface Res Appl Chem. 2021. https://doi.org/10.33263/BRIAC115.1321413231.
- 167. Velu M, Balasubramanian B, Velmurugan P, Kamyab H, Ravi AV, Chelliapan S, et al. Fabrication of nanocomposites mediated from aluminium nanoparticles/Moringa oleifera gum activated carbon for effective photocatalytic removal of nitrate and phosphate in aqueous solution. J Clean Prod. 2021;281:124553.
- de Oliveira AM, Mateus GAP, dos Santos TRT, de Filho BAA, Gomes RG, Bergamasco R. Functionalized magnetite nanoparticles with Moringa oleifera with potent antibacterial action in wastewater. Environ Technol. 2021;42:4296–305. https://doi.org/10.1080/09593330.2020.1754923.
- Gou J, Lu Y, Xie M, Tang X, Chen L, Zhao J, et al. Antimicrobial activity in Asterceae: the selected genera characterization and against multidrug resistance bacteria. Heliyon. 2023;9:e14985.
- Noor A, Ullah R, Tuzen M, Hassan A, Ullah S. Silver-coated activated carbon nanocomposite by Moringa oleifera for dye removal by multivariate optimization approach. Biomass Convers Biorefinery. 2023. https:// doi.org/10.1007/s13399-023-03847-0.
- 171. Cusioli LF, Quesada HB, de Castro ALBP, Gomes RG, Bergamasco R. Development of a new low-cost adsorbent functionalized with iron nanoparticles for removal of metformin from contaminated water. Chemosphere. 2020;247:125852.

# Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

# Journal of Nanobiotechnology







# Journal of Nanobiotechnology

Home About Articles Submission Guidelines Awards Journal Webinars

# About -

<u>Contact</u>

**Editorial Board** 

Companion journals

# **Editorial Board**

# **Editor-in-Chief**

Weibo Cai, BS, MS, PhD, University of Wisconsin - Madison, United States of America

# **Deputy Editors**

Neus Feliu, BS, MS, PhD, *Hamburg University, Germany* Jinyao Liu, *Shanghai Jiao Tong University, China* Jin Xie, *University of Georgia, United States of America* 

# **Associate Editors**

Guojun Chen, McGill University, Canada Tianfeng Chen, Jinan University, China Liang Cheng, Soochow University, China Leonardo Fernandes Fraceto, São Paulo State University, Brazil Sanket Goel, Birla Institute of Technology and Science Pilani, Hyderabad Campus, India Shreya Goel, BTech, MTech, MS, PhD, University of Utah, United States of America Zi (Sophia) Gu, BS, PhD, University of New South Wales, Australia Hao Hong, Nanjing University, China Quanyin Hu, BS, MS, PhD, University of Wisconsin, Madison, United States of America Hyung-Jun Im, Seoul National University, Republic of Korea Arnold Ju, University of Sydney, Australia Fei Liu, Brigham and Women's Hospital, United States of America Yanyan Liu, Fudan University, China Bella Manshian, BS, PhD, KU Leuven, Belgium Dalong Ni, Shanghai Jiao Tong University School of Medicine, China Pablo del Pino, University of Santiago de Compostela, Spain Ruirui Qiao, BS, MS, PhD, The University of Queensland, Australia Lang Rao, PhD, Shenzhen Bay Laboratory, China Laura Rodriguez-Lorenzo, PhD, International Iberian Nanotechnology Laboratory, Portugal Huilin Shao, National University of Singapore, Singapore Sixiang Shi, The University of Utah, United States of America Yang Shi, BS, PhD, RWTH Aachen University Clinic, Germany Shiyang Tang, University of Southampton, United Kingdom Zhongmin Tang, Tongji University, China Wei Tao, Harvard Medical School, United States of America Gang Wei, Qingdao University, China

Chun Xu, University of Sydney, Australia Guocan Yu, Tsinghua University, China Leshuai Zhang, PhD, DABT, Medical College of Soochow University, China Lijie Grace Zhang, The George Washington University, United States of America

# **Editorial Board**

Dario D Anselmetti, University of Bielefeld, Germany Zhenan Bao, PhD, Stanford University, United States of America Xuesi Chen, Changchun Institute of Applied Chemistry, Chinese Academy of Sciences, China Matthew Cooper, BSc (Hons), PhD, The University of Queensland, Australia Haixin Cui, Chinese Academy of Agricultural Sciences, China Hongjie Dai, Stanford University, United States of America Thomas P. Davis, University of Queensland, Australia Philip Demokritou, Harvard University, United States of America Tejal Desai, University of California, San Francisco, United States of America Alberto Escarpa, PhD, University Of Alcala, Spain Chunhai Fan, Shanghai Jiao Tong University School of Medicine, China Katherine W Ferrara, Stanford University, United States of America Ljiljana Fruk, PhD, University of Cambridge, United Kingdom Subhasis Ghoshal, McGill University, Canada Zijian Guo, PhD, Nanjing University, China Peng Huang, PhD, Shenzhen University, China Taeghwan Hyeon, PhD, Seoul National University, Republic of Korea Dawei Jiang, PhD, Huazhong University of Science and Technology, China Lei Jiang, Technical Institute of Physics and Chemistry, Chinese Academy of Sciences, China Laura L Kiessling, Massachusetts Institute of Technology, United States of America David Kisailus, BS, MS, PhD, University of California, Irvine, United States of America Hua Kuang, Jiangnan University, China Kit S Lam, MD, PhD, University of California, Davis, United States of America Twan Lammers, DSc, PhD, RWTH Aachen University, Germany Shyh-Dar Li, University of British Columbia, Canada Chwee T Lim, National University Singapore, Singapore Changsheng Liu, East China University of Science and Technology, China Zhuang Liu, Soochow University, China Jonathan F Lovell, BS, MS, PhD, University of Buffalo, United States of America G. Q. Max Lu, BEng, MEng, PhD, University of Surrey, United Kingdom Naomi Matsuura, University of Toronto, Canada

Chad A Mirkin, BS, PhD, Northwestern University, United States of America David J Mooney, Harvard University, United States of America Piero Morandini, PhD, University of Milan, Italy Jwa-Min Nam, PhD, Seoul National University, Republic of Korea Aydogan Ozcan, BS, MS, PhD, University of California Los Angeles, United States of America Fernando Patolsky, University of Tel Aviv, Israel Martin Pomper, UT Southwestern Medical Center, United States of America Kanyi Pu, Nanyang Technological University, Singapore Mahendra Rai, Sant Gadge Baba Amravati University, India Jianlin Shi, Shanghai Institute of Ceramics, Chinese Academy of Sciences, China Mikhail G Shapiro, California Institute of Technology, United States of America Molly Shoichet, PhD, University of Toronto, Canada Stefaan De Smedt, PhD, Ghent University, Belgium Stefaan Soenen, KU Leuven, Belgium Jie Song, Shanghai Jiao Tong University, China Francesco Stellacci, Ecole Polytechnique Fédérale de Lausanne (EPFL), Switzerland Weihong Tan, Institute of Cancer and Basic Medicine, Chinese Academy of Sciences, China Ben Zhong Tang, Hong Kong University of Science and Technology, Hong Kong Li Tang, Ecole Polytechnique Fédérale de Lausanne, Switzerland Nguyen TK Thanh, BSc, MSc, PhD, University College London, United Kingdom Zhong-Qun Tian, Xiamen University, China Matthew V Tirrell, University of Chicago, United States of America Jason Unrine, University of Kentucky, United States of America Nicolas Voelcker, Monash University, Australia Li-Jun Wan, Institute of Chemistry, Chinese Academy of Sciences, China Lihong V Wang, California Institute of Technology, United States of America Xudong Wang, University of Wisconsin-Madison, United States of America Zhenyu Wang, Jiangnan University, China Zhong Lin Wang, Georgia Institute of Technology, United States of America Karen L Wooley, Texas A&M University, United States of America Joseph C Wu, MD, PhD, Stanford University, United States of America Jianping Xie, National University of Singapore, Singapore Bengang Xing, Nanyang Technological University, Singapore Hao Yan, Arizona State University, United States of America Peidong Yang, University of California, Berkeley, United States of America Kenji Yasuda, Waseda University, Japan Yadong Yin, PhD, University of California Riverside, United States of America

Junyoung Yoon, Ewha Womans University, Republic of Korea Fan Zhang, Fudan University, China Liangfang Zhang, PhD, University of California San Diego, United States of America Dongyuan Zhao, Fudan University at Jiangwan Campus, China Yuliang Zhao, National Center for Nanoscience and Technology, China Gang Zheng, University of Toronto, Canada

Submit manuscript Journal of Nanobiotechnology



Editorial Board

Manuscript editing services

Instructions for Editors

Sign up for article alerts and news from this journal

# **Annual Journal Metrics**

# **Citation Impact 2023**

Journal Impact Factor: 10.6 5-year Journal Impact Factor: 11.4 Source Normalized Impact per Paper (SNIP): 1.296 SCImago Journal Rank (SJR): 1.840

# Speed 2024

Submission to first editorial decision (median days): 10 Submission to acceptance (median days): 100

Usage 2024 Downloads: 4,328,955 Altmetric mentions: 1,121

More about our metrics

# Follow

 $( \mathbf{y} )$ 

# **Companion Journals**

Journal of Nanobiotechnology is a companion journal to: Journal of Nanobiotechnology Cancer Nanotechnology ISSN: 1477-3155

# Contractanetschnology

# Submission anguines: journalsubmissions@springernature.com

Read more on our blogs	<u>Policies</u>	Support and Contact		
Receive BMC newsletters	<u>Accessibility</u>	Leave feedback		
Manage article alerts	Press center	<u>Careers</u>		
Language editing for authors				
Scientific editing for authors				
Follow BMC				
By using this website, you agree to our Terms and Conditions, Your US state privacy rights, Privacy statement and Cookies				

By using this website, you agree to our <u>lerms and Conditions</u>, <u>Your US state privacy rights</u>, <u>Privacy statement</u> and <u>Coo</u>policy. <u>Your privacy choices/Manage cookies</u> we use in the preference centre.

# **SPRINGER NATURE**

© 2025 BioMed Central Ltd unless otherwise stated. Part of Springer Nature.

Search Replace journals Set published About BMC Login Menu

Journal of Nanobiotechnology

Home About Articles Submission Guidelines Awards Journal Webinars

Articles 
Calections
Suggisments
Environments

4



Scheme 1. Schematic illustration describing preparation of uPA-sEVs-siSrc

A proof-of-concept study on bioorthogonal-based pretargeting and s	ignal amplify radiotheranostic strategy			
Radiotheranostics differs from the vast majority of other cancer therapies in its capacity for simultaneous imaging and therapy, and it is becoming more widely implemented. A balance between diagnostic and tre				
Hongshang Yang, Xinying Zeng, Jia Liu, Jingchao Li, Yun Li, Qinglin Zhang, Linlin Shu, Huanhuan Liu, Xueqi Wang, Yuanyuan Liang, Ji Hu, Lumei Huang, Zhide Guo and Xiasthong Zhang				
Journal of Nanobiotechnology 2024 22:101				
Research Published on: 10 March 2024				
> Full Text > PDF				
Inject [ <sup>68</sup> Ga]Ga-PSMA-Tz	Inject Pd@Au-PEG-TCO	Inject [ <sup>177</sup> Lu]Lu-PSMA-Tz		



dified Fe ions-doped carbon dots as multifu latform for ferroptosis and photodynamic synergistic therapy of mel hotodynamic therapy (PDT), the lack of efficient ferroptosis inducers an

oite the higher sensitivity of melanoma towards ferroptosis and photodyn billty of photosensitizers restrict their synergistic ... montoric inducer: and the

Jacomy of processionane in the system of the grant of the system of the





combat influenza A virus in mice by activ g the RIG-I-like receptor signaling pathway and various animal species. Infection with IAV c UIC-36 encoparticles combat influenza A virus in mice by activating the IRG-18a recepto the Influenza A virus (in bit is acrossic spheroid that infects humans and various animal ison and is often accompanied by ponumonia characturized ... Raijing Sa, Komen Li, In Saa, Javei M, Jijing Tati Tatinong Lu and Tatenin Hu Jaund of Vanobecharoogy 2024 22:99 Research | Published on: 9 March 2024 bull Tati IDE





The prognosis for hepatosellular carcinoma (CC) remains suboptimal, disasterinardo by high neuronean and metassis rates. Abboays medialismuschemp has down pointed in combat. Gowing Du, Ying Lu, Ku, Chen Du, Marin Day, Ja Xu, In Liu, Su Liu, Sjin Chen, Yi Wang, Zhigang Wang, Hatao Ran, Junni Wang and Dajing Guo



Gaurento lasadar merupanya mangkalanya yaptam nemodela satasimmune micraentotement to regenerate alvealar bone in periodentiti via tele micro las //scholentotement alvealarya micro alle status //scholentotement alvealarya bone status - scholentotementotementotementotementotementotementotementotementotemento bone status - scholentotementotementotementotementotementotementotemento bone status - scholentotementotementotementotementotementotementotemento bone status - scholentotementotementotementotementotementotementotemento bone status - scholentotementotementotementotementotementotementotementotementotemento bone status - scholentotementotementotementotementotementotementotemento bone status - scholentotementotementotementotementotementotementotemento scholentotementotementotementotementotementotementotementotementotemento scholentotementotementotementotementotementotementotementotementotementotemento scholentotemen

Journal of Nanobiotechnology 2024 22:94 Research Published on: 6 March 2024

Full Text PDE



Glone-targeted exaliplatin/furthin clathrate revensing the limit unsequencial minoremitorement through highding fraction tendent Glones are used develop restances to tennocolomical (MIX). This deviations glones accesses interleakin-10 (b. 10) and transforming growth bectory (TGP-()), recurring regulatory (TGP-ag) and highling the Acc. Xia Li Ying Chang, Zihki Yang, Gelreg A, Monglei Haar, Wellang Ye, Mao Lia, Bangle Zhang, Daoshou Lia and Sprain Zhou minorestational transforming and transforming and the Acc.

Journal of Nanobiotechnology 2024 22:93

Research | Published on: 5 March 2024 Full Text PDF



Spile structure of gold nambranchus induces haptatosticity in mouse haptatoryte organoid models Gold namografice (GRP) have item entennishy recognized as an active cardidate for a large variety of biomedical applications. However commerion of operCV (SPIC of GRP has been have indexed al... Rul Zhang, Dan Li, Rubo Zhao, Dandan Luo, Yetring Hu, Shengyan Wang, Xiaolu Zhuo, M. Zubair labai, Han Zhang, Qiangian Han and Xiar Journal of Nanobiotechnology 2024 22:92 Research Published on: 5 March 2024

Full Text PDE

Nano-enabled agrochemicals: mitigating havey metal taskisty and whanking ong adeptability for sustainable ong production The primary factors that restrict agricultural productivity and jopandoa human and food skrifty are havey metal (HMA), including assence, and akmirum, which adversally impact toro yields and q. Alazar Ghoshan, Kolazane Timumendan, Necia Pakhina, Meisam Zargar, Soyed Mehal Razvi and Moxian Chan Jammir di Manobinetnology 2024 42:31

Review | Published on: 5 March 2024 Full Text PDE

Micorhidics-enabled fluorinated assembly of EGCG ligands-aFDC ranoparticles for synergistic tumor cells and enhausted it cells regulation is cancer immune decipient with bibbr (C) derived endution offers a versatile means of developing novel immuncherapies that targets programmed death-ligand 1 (POL T)regularement death-ligand and the synthesis and targets and targets and the synthesis and targets and targets and the synthesis and targets and the synthesis and targets and the synthesis and targets and t

Research Published on: 4 March 2024

Full Text PDF



m, lead







٩

RETRACTED ARTICLE: An overview on mRNA-based vaccines to prevent monkeypox infection

The human monkeypox virus (Mpox) is classified as a member of the Pouvinidae family and belongs to the Orthopowinus genus. Mpox possesses double stranded DNA, and there are two known genetic clades: those or ... Mohammad Natami, Amirasaan Gorgatada, Araslan Gholipour, Sayedeh Narges Fatemi, Nima Firouzeh, Maryam Zokaei, Saad Hasan Mohammed Ali, Hadis Kheradjoo, Somayeh Sedighi, Omid Gholizadeh and Shaylan Kalavi

Journal of Nanobiotechnology 2024 22:86 Review | Published on: 1 March 2024

Full Text PDE

Promoting collateral formation in type 2 diabetes mellitus using ultra-small nanodots with autophagy activation and ROS scavenging Impaired collateral formation is a major factor contributing to poor prognosis in type 2 diabetes mellitus (T2DM) patients with atheror disease. However, the current pharmacological trea...

Ynuan Wang, Felfel LI, Linshuang Mao, Yu Liu, Shual Chen, Jingmeng Liu, Ke Huang, Qiujing Chen, Jianrong Wu, Lin Lu, Yuanyi Zheng, Welfeng Shen, Tao Ying, Yang Dai and Ying Shen

Journal of Nanobiotechnology 2024 22:85 Research | Published on: 1 March 2024

Full Text PDE





A novel extracellular vesicles production system harnessing matrix homeostasis and macrophage reprogramming intigates catecartivitis Observativitis (DA) is a degenerated dease that opplicativity inpairs quality of IRs. There is a pressing need for innovative DA therapies: While small extracellar vesicles (DR) also porning implements. Tarred Wang, Hongel Zhao, Yi Zhang, Yanhi Liu, Jalin Liu, Ge Chen, Ka Duan, Zhong Li, Hoi Po James Hui and Jayuan Yan Jamari di Novobiectrinology 2014 22.27 Research | nellitivitied on: 28 February 2024

Miscalls as an Escherickie odi mechanism for the accumulation and disposal of fluorescent cadmain subjuide assoparticles.
Bacterial logisynthmic of fluorescent anosparticles of quantum data, (2Da i ban bacc.)
Friege Valenzale backs, Ricells There College, Javiera Barnos Zulfage, Claudio Dietz Varget, Claudio A. Navaro and José M. Pérez Donoso
Jacunal of Navelineschoology 1202.12 273
Research | Published on: 27 February 2024
Listing of Listing of Listing of Listing Schwarz, Sc





Correction: Encapsulation of the dual FLAP/mPEGS-1 inhibitor BRP-187 into acetalated dextran and PLGA nanoparticles improves its cellular bioactivity Blerina Shkodra, Christian Kretzer, Paul M. Jordan, Paul Klemm, Andreas Koeberle, David Pretzel, Erden Banoglu, Stefan Lorkowski, Maria Wallert, Stephanie Höppener, Steffi Stumpf, Antje Vollrath, Stephanie Schubert, Oliver Werz and Ulrich S. Schubert Journal of Nanobiotechnology 2024 22:77 Correction | Published on: 27 February 2024 The <u>original article</u> was published in *Journal of Nonobion* <u>Full Text</u> <u>PDF</u> 2020 18/71 Inhable hybrid nanovaccines with virus klowinestic structure boast protective immune response spainst SARS-CoV-2 eviants Severe scalar expiratory syndrome consumus 2 (SARS-CoV-2), with different antigenic valuants, has posed a significant threat to public health. It is urgent to develop inhable vaccines, instead of injectab. Shuqi Wang, Pelyang Ding, Lingli Shen, Daopeng Fan, Hanghang Cheng, Jian Huo, Xin Wei, Hua He and Gaiping Zhang Journal of Nanobiotechnology 2024 22:76 Research | Published on: 27 February 2024

Full Text PDF

Highly sensitive SERS platform for pathogen analysis by cyclic DNA nanostructure@AuNP tags and cascade primer exchange The capacity to identify small amounts of pathogens in real samples is extremely useful. Herein, we proposed a sensitive platform for detecting patho using cyclic DNA nanostructure@AuNP tags (CDNA) and a c\_ Viru Xiao, Shihua Luo, Jiuxiang Qiu, Ye Zhang, Weijiang Liu, Yunhu Zhao, YiTong Zhu, Yangsi Deng, Mengdi Lu, Suling Liu, Yong Lin, Alwei Huang, Wen Wang, Xueijao Hu and Bing Gu Journal of Nanobiotechnology 2024 22:75 Research | Published on: 26 February 2024 Full Text PDF

Apoptotic extracillular walches derived from hyposis-preconditioned meanchymal stem cells within a modified gelatine hydrogel promote extendenderal regeneration by enhancing stem cell activity and regulating immunity. Date to its unique structure, activatar cartilage has immited abilities to undergo self-repair after injury Additionally, the repair of articular cartilage after injury has always been differendering in the cell activity and regulating immunity. Zhengang Ding, Zheng Yan, Xui Yuan, Guangahao Tian, Jiang Wu, Liwei Fu, Han Yin, Songlin He, Chao Ning, Yazhe Zheng, Zhichao Zhang, Xiang Su, Libo Hao, Xinng Ng, Wang Nua, Li Wileimi Gu. Journal of Nanobiotechnology 2024 22:74 Research | Published on: 23 February 2024

Full Text PDF



The formation of blood vessel system under a relatively higher Cu<sup>2+</sup> ion level is an indispensable precondition for tumor proliferation and migration, which was assisted in forming the tumor immune microenvironmen... Wenling Li, Huan Xin, Wenjuan Gao, Pengjun Yuan, Fekue Ni, Jingyi Ma, Jingrui Sun, Jianmin Xiao, Geng Tian, Lu Liu and Guilong Zhang

Journal of Nanobiotechnology 2024 22:73 Research | Published on: 19 February 2024

Full Text PDF

# M2 marcophage-derived exosomal mIR-26b-5p regulates marcophage polarization and chondrocyte hypertrophy by targeting TLR3 and COLIDA1 to alleviate outeoarthritis

allendae onterstriftig Octoaenthis (DA) is one of the most pervalent chronic musculosheital diseases among the elderly population. In this study, macrephage-derived exoco were located and is distributed. Incomers were subjected ... Yufun Qan, Genglei Chu, Lei Zhang, Zhika Wu, Quyuan Wang, Jiong siong sion and Feng Zhou

Journal of Nanobiotechnology 2024 22:72

Research | Published on: 19 February 2024 Full Text PDE







rfacial effects and biomedical applications in cancer treatment Two-dimensional nanomaterials (2D NMs), characterized by a large number of atoms or molecules arranged in one dimension (typically thickness) while having tiny dimensions in the other two dimensions, have emer...

Yan Wang, Xiao Zhang and Hua Yue

Journal of Nanobiotechnology 2024 22:67

Review Published on: 18 February 2024





rutich diet o

Binding 

Mannal Brach chi dia promota aanju dha linatada develapmanti a dharping through milluderind antxicultur walchs carrying mili 2463-25 In tantanti develapming in anta higi higi profondily influenced by multiple biological camponens of braza mili, in which mili-devined estacultur (mW) contain a large amount of verticaly trac. Dongstong Lu, Yoi Lu, Luyaan Kang, Xangyu Zhang, Jie Hu, Hao Ye, Bingu Huang, Yujun Wu, Johiao Zhao, Zhaola Dai, Junjun Wang and Dandan H

Iournal of Nanobiotechnology 2024 22:65

esearch Published on: 16 February 2024

Full Text PDF Maternal resistant starch (RS) diet Growth perform Intestinal developm Breastfeeding 1 Diarrhea in weanling -derived extracellular (mEVs) vesicles Wnt Omevs Absorption 00 V **B**-catenin GSK3 miR-146a-5p DVL2 AXIN2

NEDD4L

**B-cat** 



vity of copper oxide nanoparticles derived from Zizyphus spin Sozan E. El-Abeid, Mohamed A. Mosa, Mohamed A. M. El-Tabakh, Ahmed M. Saleh, Moi med A. El-Khateeb and Maha S. A. Haridy

Journal of Nanobiotechnology 2024 22:64 Correction | Published on: 16 February 2024

The original Full Text PDE

#### Recent adces in two-dim insional nai naterials for s able wearable electronic devi

Recent advances in two-dimensional nanomatrials for startistudie warable electronic devices the valesported advances of neutra terminal-test significantly bootest the market potential for was nanomatrials show great promise for flexible, warable electronic. Jang Ina and Mingdong Dong Januard of Nanobeshowskog 2004 22251 Review | Published on 15 February 2024

Full Text PDF



Exosomes s hsa\_circ\_00 ted by Fu ance to oxaliplatin and 5-FU by delivering has dec 0000055 A Wage number of inductorium nucleatum Fin) are present in colorectal cancer (OCC) tissues of patients who relapse after chemotherapy, and Fin bas been reported by promite a calcipation and Fin chemosestatures L. Bragging Huk, Chemchen Zhou, Watao Xiu, Huk Wang, Yuwen Dong, Yinu Zhou, Jie Ding, Xiao Zhang, Jian Xu and Yanhong Gu Jamand of Rensolectioning 2020 22-22. Research J. Published on: 15 Reinary 2020





The quest for nand -powered var

The spectra of an application protection is called a manufacturing of the spectra nes play an important role in preventing

Journal of Nanobiotechnology 2024 22:61 Review | Published on: 14 February 2024

Full Text PDE



sheno Zhano







# M2 microglie durind easemes prenete vascular emodaling in diabetic retingentity Diabetic retinopathy (DR) is a vision-threatening diabetic complication that is characterised by microwasculature impairment and immune dystanction. The present titudy demonstrated that KM microglia intensively... Xongening Wang, Changlin Xu, Cumini Bian, Penglei Ga, Ja Lai, Jangdan Wang, Tanhao Xiao, Yuanyuan Fan, Opiyuan Gu, Hong Ying Li, Angyi Xu, Zahong Hu and Phing Xu.

Journal of Nanobiotechnology 2024 22:56

Research Published on: 9 February 2024



Polydopamine assisted aptame-carrying tetrahedral DNA microslectoride sensor for ultrasmittine electrochemical detection of eassames Exconses are nanocale estrachilar and exides (D-160 m) with endocreme origin exceeded by almost all types of calls, which are considered to be in interestibility commission. Canceroux excessions senses are Exceen Sang. Tengha Zhang, Slan Liu, the Sheng and Zaming Hu Journal of Nanobiotechnology 2024 22:55 Research | Published on: 8 February 2024 The <u>Connection to this article</u> has been published in Full Text PDE



Fabrication of levelfasach-loaded parcine acallular domail matrix hydrogal and functional assessment in uninary tract infection Bacterial cyclics, a commonly occurring uninary tract infection (JTII) is removined for its estemake prevalence and tendency to recur. Depite the estemake unitation of involvement as a convertional thema. Yi Yang, Guang-Wei Yang, Jian-Juan Lu, Hao-Ran Chen, Ya-Qin Guo, Ning Yang, Yun-Zhu Zhu, Xiao-Qiang Liu, Ting-Ting Su, Yan-Yan Liu, Liang Yu, Ya-Sheng Li, Li-Fen Hu and Ja-Bin Li

Native matrix

2.04

Journal of Nanobiotechnology 2024 22:52

Original Paper | Published on: 7 February 2024 Pig skin

Derma

Extracellular matrix Collagenous fiber 1%Triton 12h 0.1%SDS 6h Pepsin pH2-3





SPRINGER NATURE © 2025 BioMed Central Ltd unless otherwise stated. Part of <u>Springer Nature</u>.

Scimago Journal & Countr	y Rank						Enter Journal Title, ISSN or Publisher Name
	Home	Journal Rankings	Journal Value	Country Rankings	Viz Tools	Help	About Us

# Journal of Nanobiotechnology 8

COUNTRY	SUBJECT AREA AND CATEGORY	PUBLISHER	SJR 2024	_
United Kingdom Universities and research institutions in United Kingdom	Biochemistry, Genetics and Molecular Biology Molecular Medicine	BioMed Central Ltd	2.282 Q1	
Media Ranking in United Kingdom	Engineering Bioengineering Engineering Biomedical Engineering		H-INDEX 125	
	Immunology and Microbiology Applied Microbiology and Biotechnology			
	Materials Science Nanoscience and Nanotechnology			
	Medicine Medicine (miscellaneous)			
	Pharmacology, Toxicology and Pharmaceutics Pharmaceutical Science			
PUBLICATION TYPE	ISSN	COVERAGE	INFORMATION	
Journals	14773155	2003-2025	Homepage	
			How to publish in this journal	

SCOPE

Journal of Nanobiotechnology is an open access peer-reviewed journal communicating scientific and technological advances in the fields of medicine and biology, with an emphasis in their interface with nanoscale sciences. The journal provides biomedical scientists and the international biotechnology business community with the latest developments in the growing field of Nanobiotechnology. The journal consists of the following sections: -Nanomaterials in Biology, Medicine, Food, Environment and Agriculture -Nanoparticles in Biology, Medicine, Food, Environment and Agriculture -Nanosensors in Biology, Medicine, Food, Environment and Agriculture -Nanoensors in Biology, Medicine, Food, Environment and Environment -Nanoensors in Biology, Medicine, Food, Environment and Environment -Nanoensors in Biology, Medicine, Food, Environment -Nanoensors in Biology, Medicine, Food, Environment -Nanoensors in Biology, Berger - Source - Sourc

 $\ensuremath{\bigcirc}$  Join the conversation about this journal



# G SCImago Graphica

Explore, visually communicate and make sense of data with our new data visualization tool.



# Metrics based on Scopus® data as of March 2025

V	Vefa	7 months ago
v	veia	7 monuns ago

# Is it free?

reply

# Melanie Ortiz 7 months ago

Dear Vefa, Thank you for contacting us. We suggest you visit the journal's homepage or contact the journal's editorial staff , so they could inform you more deeply. Best Regards, SCImago Team

~

~

#### Μ Megersa Abdisa Debelo 3 years ago

HOW CAN I PUBLISH PAPAER IN YOUR INSTITUTION?

reply

# (Č

#### Melanie Ortiz 3 years ago

Dear Megersa, thank you very much for your comment, we suggest you look for the author's instructions/submission guidelines in the journal's website. Best Regards, SCImago Team

V

# vyjainto kumar ray 5 years ago

Sir,

Is your journal Scopus indexed ? If yes, please let me know the name of the journal and also kindly let me know how to publish my research papers in your journal. How many days does it take to accept a research paper and publish it ? Also if possible, please write the names of other journals having Scopus index value.

Thanking you.

Vyjainto Kumar Ray

# India.

reply



# Melanie Ortiz 5 years ago

Dear Vyjainto,

SCImago Team

SCImago Team

SCImago Team

Thank you very much for your comment. We are sorry to tell you that SCImago Journal &

Country Rank is not a journal. SJR is a portal with scientometric indicators of journals indexed in Elsevier/Scopus. All the metadata have been provided by Scopus /Elsevier in their last update sent to SCImago, including the Coverage's period data. The SJR for 2019 was released on 11 June 2020. We suggest you consult the Scopus database directly to see the current index status as SJR is a static image of Scopus, which is changing every day.

Best Regards, SCImago Team



~

Leave a comment

Name

Email (will not be published)

Submit

The users of Scimago Journal & Country Rank have the possibility to dialogue through comments linked to a specific journal. The purpose is to have a forum in which general doubts about the processes of publication in the journal, experiences and other issues derived from the publication of papers are resolved. For topics on particular articles, maintain the dialogue through the usual channels with your editor.





# Source details

Journal of Nanobiotechnology Open Access	Cite 16	Score 2024 (j
Years currently covered by Scopus: from 2003 to 2025		
Publisher: Springer Nature E-ISSN: 1477-3155	sjr 2.2	<sup>2024</sup> <sup>(i)</sup>
Subject area: (Medicine: Medicine (miscellaneous)) (Pharmacology, Toxic	ology and Pharmaceutics: Pharmaceutical Science	
(Engineering: Biomedical Engineering) (Immunology and N View all V	Microbiology: Applied Microbiology and Biotechnology SNII	<sup>p</sup> 2024 (j)
Source type: Journal		
View all documents > Set document alert Save to source l	list	
CiteScore CiteScore rank & trend Scopus content cov	verage	
CiteScore 2024 ~ Cite	eScoreTracker 2025 🛈	
37.067 Citations 2021 - 2024	29.051 Citations to date	
16.8 = 2.200 Documents 2021 - 2024	2.051 Documents to date	
Calculated on 05 May, 2025	pdated on 05 May, 2025 • Updated monthly	
CiteScore rank 2024 🕤		

Q

Category	Rank Percen	tile
Medicine Medicine (miscellaneous)	#11/433	97th
Pharmacology, Toxicology and Pharmaceutics Pharmaceutical	#6/185	97th

View CiteScore methodology ightarrow CiteScore FAQ ightarrow Add CiteScore to your site  $\mathscr{S}$ 

# About Scopus

What is Scopus	
Content coverage	
Scopus blog	
Scopus API	
Privacy matters	
Language 日本語版を表示する 查看简体中文版本 查看繁體中立版本	
宣目系脰中又似乎	
Просмотр версии на русском языке	

# **Customer Service**

Help Tutorials

Contact us

# ELSEVIER

Terms and conditions  $\ensuremath{\nearrow}$   $\ensuremath{\mathsf{Privacy}}$  policy  $\ensuremath{\nearrow}$   $\ensuremath{\mathsf{Cookies}}$  settings

All content on this site: Copyright © 2025 Elsevier B.V. inite licensors, and contributors. All rights are reserved, including those for text and data mining, AI training, and similar technologies. For all open access content, the relevant licensing terms apply. We use cookies to help provide and enhance our service and tailor content.By continuing, you agree to the use of cookies <math>inite definition definition definition definition definition definition definition definition. The second definition definition definition definition definition definition definition definition definition.

*R***ELX**<sup>™</sup>